Biochemical characteristics & secretory activity of *Aeromonas* species isolated from children with gastroenteritis in Chennai

S. Ananthan & S.V. Alavandi

Department of Microbiology, Dr ALM Post-Graduate Institute of Basic Medical Sciences University of Madras, Chennai

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An attempt was made to delineate the phenotypic markers for the detection of enterotoxigenic strains of Aeromonas. Eighteen Aeromonas species comprising one isolate of A. hydrophila, six isolates of A. sobria and 11 isolates of A. caviae were obtained from 379 children suffering from acute diarrhoea in Chennai. Nine of these isolates inclusive of three A. sobria and six A. caviae were found to produce secretory response in vitro in the rabbit intestinal mucosa mounted in the Ussing chambers as revealed by significant increases in the short circuit current. Eleven strains hydrolysed aesculin, 8 fermented arabinose, 6 produced acetyl methyl carbinol, 14 produced lysine decarboxylase, 3 fermented salicin, 9 produced β -haemolysin, 9 produced CAMP-like factor and only two isolates took up congo red dye. None of these phenotypic traits were found to correlate with the in vitro secretory activity.

Key words Aeromonas - diarrhoea - secretory response

Aeromonas species have been implicated in diarrhoea the world over¹. This species produces a variety of virulence factors such as haemolysins, cytotoxins, siderophores, enterotoxins etc². Some studies have reported that enterotoxigenicity is related to species, particularly to A. hydrophila and A. sobria³ and that only the β-haemolytic strains were enterotoxigenic⁴. Some others have suggested certain phenotypic characteristics such as production of acetyl methyl carbinol (VP), lysine decarboxylase, haemolysis on sheep blood agar, CAMP-like factor, inability to hydrolyse arabinose, susceptibility to cephalothin as potential virulence markers and that these tests would identify up to 97 per cent of enterotoxigenic isolates⁵⁻⁸. A. caviae, because of their VP negativity and non-haemolytic nature would be

excluded if these criteria are employed. But A. caviae have been also recognised an important diarrhoeagenic aeromonad in children and aged individuals9 based on its enterotoxigenicity in suckling mice and cytotoxic response in HEp-2 cells in vitro. The present study was conducted to delineate important phenotypic marker(s), if any, to identify enterotoxigenic strains in a routine diagnostic laboratory. The Aeromonas isolates obtained from children suffering from acute diarrhoea in Chennai were biochemically characterised and the enterotoxigenicity of these isolates determined by their ability to induce secretory response in Ussing chambers. The phenotypic characteristics were examined for correlation with secretory activity.

Material & Methods

Isolation and identification: The study was conducted from February 1997 to April 1998 at Chennai. Rectal swabs were collected in duplicate from 379 children under 5 yr of age suffering from acute diarrhoea, admitted to the Institute of Child Health and Hospital for Children, Chennai, and transported to the laboratory in Cary Blair medium. One swab was inoculated onto Inositol Brilliant green bile salts (IBB) agar (Himedia, Mumbai) and the other was inoculated into alkaline peptone water (APW) for enrichment and incubated at 37°C for 18-24 h. A loopful from the APW was inoculated onto IBB and 5 per cent sheep blood agar (SBA). The non-inositol fermenting colonies on IBB were subcultured on nutrient agar and incubated at 37°C overnight. The colonies on nutrient agar and SBA were tested for oxidase and positive isolates were identified as described earlier¹⁰. The biochemical tests, esculin hydrolysis, production of acetyl methyl carbinol (VP), lysine decarboxylase (LDC), ornithine decarboxylase (ODC), arginine dihydrolase (ADH), fermentation of arabinose, sucrose and salicin, and detection of extracellular enzyme activities for protease, lipase and amylase were carried out by conventional methods11.

In vitro secretory activity: Cell free culture filtrates were prepared by growing the isolates in brain heart infusion (BHI) broth at 37°C for 18 h and the cultures were centrifuged at 7000 g at 4°C and the supernatants were filtered through 0.22 µm syringe filters (Millipore Inc, USA). These culture supernatants were tested for secretory activity in the intestinal mucosa mounted in the Ussing chambers (World Precision Instruments, USA) as earlier described¹². Briefly, the rabbit ileal tissue cut in a rectangular shape, obtained from a freshly sacrificed animal was mounted in the Ussing chambers, the tissue was bathed in Ringers solution on both serosal and mucosal sides and aerated with 5 per cent CO, and 95 per cent O, mixture. The instrument was balanced and the readings of short circuit current (Isc) and potential difference (PD) were recorded. 400 µl of culture filtrate was added on both serosal and mucosal sides and PD and Isc were measured every 15 min for 90 min. An enterotoxigenic strain of Vibrio cholerae was used as a positive control. The cell free culture filtrates of isolates showing increases in the Isc were regarded as positive for in vitro secretory activity¹³.

Production of haemolysin: Production of haemolysin by the Aeromonas isolates was tested as described earlier⁵. 50 μl of cell free culture filtrates of the Aeromonas isolates prepared as described above were mixed with 50 μl of 2 per cent suspension of sheep RBCs in PBS in a round bottomed 96 well microtitre plate and examined for haemolysis after incubation at 37°C for 1 h and a further incubation at 4°C for 18 h.

CAMP-like factor: The ability of Aeromonas strains to act synergistically with a β -haemolytic Staphylococcus aureus to produce more potent haemolysis indicated by arrowhead shaped zones of haemolysis was determined by the methods described earlier¹⁴.

Congo red uptake: Aeromonas isolates were inoculated onto trypticase soy agar containing 50 µg of congo red dye per ml of the medium and incubated for 18 h at 37°C. The plates were examined for isolates showing various shades of orange under obliquely reflected light on a black background which indicated uptake of the dye by the organisms¹⁵.

Statistical analysis: The relationship between various biochemical characteristics singly and in various combinations with secretory activity was examined by simple correlation analysis using SPSS computer software.

Results & Discussion

In the present study we were able to recover Aeromonas species from 18 paediatric patients suffering from diarrhoea. The 18 isolates were identified to genus level by the criteria of Aerokey II¹⁶. A. caviae was recovered from 11 patients, while A. sobria was isolated from six patients and A. hydrophila from one, indicating the predominance of A. caviae in paediatric patients as reported by earlier workers¹⁷.

Out of 18 isolates, 6 produced VP, 11 hydrolysed aesculin, 8 fermented arabinose, 3 fermented salicin, 14 produced lysine decarboxylase, 13 produced arginine dihydrolase, 9 produced β-haemolysin, 9 exhibited CAMP-like factor, 2 took up congo red dye, 8 produced lipase, 13 produced protease, 14 produced amylase and 3 isolates showed susceptibility to cephalothin (Table).

Test								-	Isolates											Total	
	A.h 8143	A.s 38a	A.s 374	A.s	A.s 6143	A.s 811	A.s 1262	A.c 359	A.c.	A.c.	A.c 96	A.c 106	A.c 113	A.c 1154	A.c 1233	A.c 1235	A.c 1264	A.c 121 (A.h (n=1) (A.s (n=6) (A.c (n=11)
Oxidase	.+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	_	9	=
Motility	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	9	Ξ
TSI	K/A,g	K/A,g	K/A,g K/A,g	4 /	K	K/A,g	¥¥	¥ ¥	ΑA	A/A	A/A	ΑV	A/A	K/A	A/A	ΚΆ	A/A	A/A			
Indole	+	+	+	+	•	+	•	+	+	+		+	+	•	+	+	•	+	_	4	00
VP	+	+	+		+	+	+	1		•			•	•		•		•	-	2	0
Aesculfa hydrolysis	+	•			+		,	+	+	+	+	+	+	+	+		,	+	-	_	6
Res to O/129-150 µg	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	9	=
Inositol fermentation	•	•	•					,									,		0	0	0
Sucrose fermentation	+	+	+	+	+	+	+	+	+	+	+	+	+		+	+	+	+	-	9	2
Arabinose fermentation	+	•	•		+				+	+	+	+				+	ı	+	-	-	9
Mannitol fermentation	+	+	+	+	+.	+	+	+	+	+	+	+	+	+	+	+	+	+	-	9	=
Glucose fermentation	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	9	Ξ
Lactose fermentation	•	,		٠,	,						٠						1		0	0	0
Salicin fermentation	•	•	1				+	•	+					+					0	_	7
Lysine decarboxylase	+	+	+	•	+	+	+	+		+	+		+	+	+		+	+	-	8	00
Arginine dihydrolase	+	•		+	•	+	+	+	+	, + .	+	+	+		+		+,	+	-	٣	6
Omithine decarboxylase	•	•		,	•									ı	•	ı		•	0	0	0
NO ₃ reduction	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	9	Ξ
Cephalothin sensitivity	+	•	+									+					•	•	_	-	-
CAMP-factor	+	+	+	•	+	+	+		(x)				(((((((((((((±(€)		ı			-	'n	3
Congo red uptake	,	٠		•	•	+	+	,			•			,			٠		0	7	0
Lipase	+	٠		+	•	+		•	+	•		+	+	•	+			+	_	7	2
Protease	+	+	+	+	+	+			+		+	+		+	+	+	+		-	2	7
Amylasė	+	+	+	+	+		+	+	+		+	+	+			+	+	+	_	2	∞
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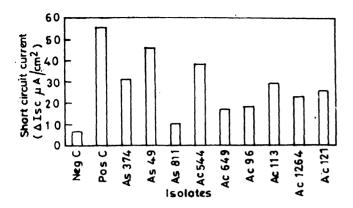


Fig. Secretory activity of Aeromonas isolates in Ussing chambers.

Nine of these isolates (3 of A. sobria and 6 of A. caviae) induced increase in the short circuit current (Δ Isc) of the ileal tissue mounted in the Ussing chambers, the average Δ Isc ranging from 10.5 μ A to 46 μ A/cm² (Fig.).

Attempts to correlate some of the phenotypic characteristics such as production of VP, LDC, ADH, CAMP factor, haemolysin etc., with the in vitro secretory response, showed no correlation between any of these parameters with secretory activity. Some earlier studies^{6,8} had reported that the enterotoxigenic strains are VP positive, arabinose negative, lysine decarboxylase positive and produce haemolysin⁵. In the present case, such a correlation was not observed either singly or in combination to be predictive of enterotoxigenicity. Singh and Sanyal¹⁸ also did not observe correlation between any of the biochemical characteristics such as VP, lysine decarboxylase, non-fermentation of arabinose or haemolysin production based on their study of 147 isolates inclusive of 54 isolates from patients of diarrhoea.

Congo red uptake has been reported as a virulence marker for several pathogenic bacteria such as Shigella¹⁹, Yersinia enterocolitica²⁰ and V. cholerae²¹. Earlier studies on the congo red uptake by Aeromonas spp showed no correlation with VP or lysine decarboxylase¹⁵; correlation between congo red uptake and enterotoxigenicity was not attempted. In the present study, only two isolates showed ability to take up congo red dye and its relationship with secretory activity could not be established.

Production of CAMP - like factor was suggested for differentiation of Aeromonas species ¹⁴. A. hydrophila produced CAMP - like factor both aerobically and anaerobically and A. sobria produced CAMP factor only aerobically, whereas A. caviae did not produce CAMP factor ¹⁴. In the former two species, which are more often reported to be associated with human disease, production of CAMP factor was considered as a virulence factor. Three of our A. caviae isolates which were α -haemolytic showed mild CAMP-like factor upon further incubation of the plates overnight at 4°C. Hence this scheme of differentiation of Aeromonas is unlikely to be useful in view of the fact that A. caviae isolates can be haemolytic ²².

Susceptibility to cephalothin was reported to be associated with A. sobria⁷, the species that is considered to be invasive among the aeromonads²³. In the present study, susceptibility to cephalothin was uncommon among A. sobria isolates and there was no relationship with secretory response of the isolates.

Production of several extracellular virulence factors such as lipase, protease and haemolysin was reported to be variable in Aeromonas isolates obtained from faecal samples of patients suffering from gastroenteritis and, it has been suggested that high protease and low lipase activity along with the autoagglutinating character be considered as virulence marker²⁴. In the present study, although 13 isolates showed protease activity and 8 isolates lipase activity, both these parameters also did not show any correlation with the secretory activity. Production of enterotoxin in vitro varies with the conditions of culture and the method of assay25 and probably many other factors. Cahill²⁶ suggested that the cytotoxins, haemolysins, enterotoxins etc., are manifestations of a single factor. In view of our observations and the lack of correlation of the phenotypic parameters with the in vitro secretory activity it is difficult to suggest any of the phenotypic characters as markers of virulence or indicators of diarrhoeagenic potential in Aeromonas species.

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References

- Altwegg M, Geiss HK. Aeromonas as a human pathogen. CRC Crit Rev Microbiol 1989; 16: 253-86.
- Janda JM. Recent advances in the study of the taxonomy, pathogenicity and infectious syndromes associated with the genus Aeromonas. Clin Microbiol Rev 1991; 4: 397-410.
- Barer MR, Millership SE, Tabaqchali S. Relationship of toxin production to species in the genus Aeromonas. J Med Microbiol 1986; 22: 303-9.
- Burke V, Gracey M, Robinson J, Peck D, Beaman J, Bundell C. The microbiology of childhood gastroenteritis: Aeromonas and other infective agents. J Infect Dis 1983; 148: 68-74.
- Burke V, Robinson J, Atkinson HM, Gracey M. Biochemical characteristics of enterotoxigenic Aeromonas spp. J Clin Microbiol 1982; 15: 48-52.
- Turnbull PC, Lee JV, Miliotis MD, van de Walle S, Koornhoff HJ, Jeffery L, et al. Enterotoxin production in relation to taxonomic grouping and source of isolation of Aeromonas species. J Clin Microbiol 1984; 19: 175-80.
- Janda JM, Motyl MR. Cephalothin susceptibility as a potential marker for the Aeromonas sobria group. J Clin Microbiol 1985; 22: 854-5.
- Kirov SM, Rees B, Wellock RC, Goldsmid JM, Van Galen AD. Virulence characteristics of *Aeromonas* spp in relation to source and biotype. *J Clin Microbiol* 1986; 24: 827-34.
- Namdari H, Bottone E. Cytotoxin and enterotoxin production as factors delineating enteropathogenicity of Aeromonas caviae. J Clin Microbiol 1990; 28: 1796-8.
- Komathi AG, Ananthan S, Alavandi SV. Incidence and enteropathogenicity of *Aeromonas* spp in children suffering from acute diarrhoea in Chennai. *Indian J Med Res* 1998; 107: 252-6.
- Smibert RM, Krieg NR. Phenotypic characterisation. In: Gerhardt P, Murray RGE, Costilow RN, Nester EW, Woods WA, Krieg NR, editors. Manual of methods for general and molecular biology. Washington DC: American Society for Microbiology (ASM); 1991 p. 607-54.
- 12. Fasano A, Kay BA, Russel RG, Maneval DR Jr, Levin MM.

- Enterotoxin and cytotoxin production by enteroinvasive Escherichia coli. Infect Immun 1990; 58: 3717-23.
- Sears CL, Kaper JB. Enteric bacterial toxins: mechanisms of action and linkage to intestinal secretion. *Microbiol Rev* 1996; 60: 167-215.
- Figura N, Guglielmetti P. Differentiation of motile and mesophilic Aeromonas strains into species by testing for a CAMP-like factor. J Clin Microbiol 1987; 25: 1341-2.
- 15. Statner B, George L. Congo red uptake by motile Aeromonas species. J Clin Microbiol 1987; 25: 876-8.
- Carnahan AM, Behram S, Joseph SW. Aerokey II: A flexible key for identifying clinical *Aeromonas* species. *J Clin Microbiol* 1991; 29: 2843-9.
- San Joaquin VH, Picket DA. Aeromonas-associated gastroenteritis in children. Pediatr Infect Dis J 1988; 7: 53-7.
- Singh DV, Sanyal SC. Biochemical characteristics and enterotoxicity of Aeromonas species isolated from man and environment. J Diarrhoeal Dis Res 1992; 10: 231-4.
- Maurelli AT, Blackmon B, Curtiss R 3rd. Loss of pigmentation in *Shigella flexneri* 2a is correlated with loss of virulence and virulence-associated plasmid. *Infect Immun* 1984; 43: 397-401.
- 20. Prpic JK, Robbins-Browne RM, Davey RB. Differentiation between virulent and avirulent *Yersinia enterocolitica* isolates by using Congo red agar. *J Clin Microbiol* 1983; *18*: 486-90.
- Payne SM, Finkelstein RA. Detection and differentiation of iron-responsive avirulent mutants on Congo red agar. *Infect Immun* 1977; 18: 94-8.
- Singh DV, Sanyal SC. Haemolysin and enterotoxin production by Aeromonas caviae isolated from diarrhoeal patients, fish and environment. J Diarrhoeal Dis Res 1992; 10: 16-20.
- 23. Daily OP, Joseph SW, Coolbaugh JC, Walker RI, Merrel BR, Rollins DM, et al. Association of Aeromonas sobria with human infection. J Clin Microbiol 1981; 13: 769-77.
- Pin C, Marin ML, Selgas D, Garcia ML, Tormo J, Casas C. Differences in the production of several extracellular virulence factors in clinical and food Aeromonas spp strains. J Appl Bacteriol 1995; 78: 175-9.
- 25. Ljungh A, Wretlind B, Mollby R. Separation and characterisation of enterotoxin and two haemolysins from *Aeromonas hydrophila*. *Acta Pathol Microbiol Scand B* 1981; 89: 387-97.
- Cahill MM. Virulence factors of motile Aeromonas species. J Appl Bacteriol 1990; 69: 1-16.

Reprint requests: Dr S. Ananthan, Professor, Department of Microbiology, Dr A.L.M. Post-Graduate Institute of Basic Medical Sciences, Taramani, Chennai 600113