

# QTL mapping for heat tolerance related traits using backcross inbred lines in wheat (*Triticum aestivum* L.)

Sunil, Harikrishna\*, Deepali Upadhyay, Rahul Gajghate, P. Shashikumara, Divya Chouhan, Swetha Singh, V. P. Sunilkumar, B. Manu, Nivedita Sinha, Sanjay Singh, Neelu Jain, G. P. Singh and P. K. Singh

Division of Genetics, ICAR-Indian Agricultural Research Institute, New Delhi 110 012; <sup>1</sup>ICAR-Indian Institute of Wheat and Barley Research, Karnal

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### Abstract

Heat stress is one of the most limiting factors for the production of wheat. Global warming and consequent changes in climate adversely affect wheat plant growth and yield. To elucidate genetic basis and map heat tolerance traits, a set of 134 backcross inbred lines (BILs) derived from the cross between WH730/\*2 HD2733 was used. The population was evaluated under late sown (LS) and very late sown (VLS) conditions, by exposing to heat stress during rabi season. Positive association of normalized difference vegetation index (NDVI), thousand grain weight (TGW), grain weight per spike (GWS), biomass and grain yield (GY) under both production conditions was observed. However, canopy temperature (CT) and days to heading (DH) showed negative correlation with GY under heat stress. A total of 9 Quantitative trait loci (QTL) were discovered on 7 chromosomes, which includes 4 QTLs in LS and 5 QTLs under VLS condition. Combining the results of these QTLs revealed a major stable QTL for DH (qDH\_iari\_5A) on chromosome 5A with 23% and 26% explaining phenotypic variance under both sowing conditions. QTL for NDVI was detected on chromosome 1B while QTL for SL and GY on chromosome 2A. The identified QTLs in the genomic regions could be targeted for genetic improvement and marker assisted selection for heat tolerance in wheat.

Key words: Wheat, QTL Mapping, Heat tolerance

### Introduction

Wheat (*Triticum aestivum* L.) is one of the most important and widely adapted strategic food crops. Heat stress is a major environmental constraint to crop production. Sensitivity to heat stress is a major limitation to growth and productivity of wheat as especially in sub-tropical and dry regions. Episodes of heatwaves in combination with drought are serious during the anthesis and grain-filling period, which are the most vulnerable stage affecting the final yield (Ortiz et al. 2008). Potential expression of important traits determining yield formation is essential for productive selection in any breeding plan (Sagar et al. 2016). The decline in yield is the outcome of decreased grain weight, early senescence, shriveled grains, reduced starch accumulation, altered starch-lipid composition in grains, lower seed germination and loss of vigor (Balla et al. 2012). To withstand heat stress and yield normal, plants have evolved with special mechanisms to adapt. The dissection of genomic regions associated with heat tolerance through the mapping of QTLs is essential to know the genetic basis of heat tolerance. It helps to understand the association of traits related to heat stress and pave the way for augmenting the wheat productivity under heat stress conditions. Therefore, breeding for tolerance to chronic as well as short term heat stress is a major objective worldwide (Ayeneh et al. 2002; Yang et al. 2002; Nachit et al. 2004; Reynolds et al. 2007; Jain et al. 2014; Reynolds et al. 2016). In view of the facts mentioned above, the present study was undertaken for characterization of morpho-physiological traits in BILs and mapping QTLs associated with heat tolerance related traits for improving heat stress tolerance in wheat.

### Materials and methods

### Experimental design and materials

In the present study, a set of 134 BILs derived from the cross between HD2733 and WH730 (IC546937, an elite wheat genotype having highly stable yield and

<sup>\*</sup>Corresponding author's e-mail: harikrishna.agri@gmail.com

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better adaptability to terminal heat stress) along with parents were sown in  $\alpha$ -lattice design under late and very late sown conditions with two replications constituting eight blocks (each block contains 17 test genotypes) in rabi 2017. WH730, Experiment was carried out at an experimental farm, Division of Genetics, ICAR-Indian Agricultural Research Institute, New Delhi. Dhanda et al. (2012) reported earlier that WH730 is a heat stress tolerant genotype which recorded high grain yield and with its low heat susceptibility index, high thermo-tolerance of membrane, high kernel weight and grain number are the important fetures of this variet. The F1 seeds were harvested and sown in rabi 2012. The F1 were backcrossed using as male parent to HD2733, and total 134 individuals of BC<sub>1</sub>F<sub>1</sub> plants were allowed to self-pollinate and then harvested seeds from each individual plant were allowed to advance up to BC<sub>1</sub>F<sub>5</sub>.

### Phenotyping and genotyping of BILs population

The population was phenotyped for nine morphophysiologial traits namely, days to heading (DH), grain weight per 10 spikes (GWS), spike length (SL), number of spikelet's per spike (NSPS), plant biomass (PB), thousand kernels weight (TKW), grain yield (GY), canopy temperature (CT) and normalized difference vegetation index (NDVI) under late sown (LS) and very late sown (VLS) conditions as follows; Parental means described here were the best linear unbiased predictions (BLUP) which were calculated using R package. The physiological traits CT and NDVI were recorded at different stages of the crop growth.

Genomic DNA of parental genotypes and BILs was isolated using CTAB method of Saghai-Maroof et al. (1984). Quantity and quality of genomic DNA were tested on 0.8% agarose gel by using standard DNA ladder. A total of 1350 Simple sequence repeats (SSRs) were screened and 95 of which were identified as polymorphic SSRs, distributed over different chromosomes were used to genotype the entire population. Parents were evaluated for polymorphism using different series of markers (gwm/wms, wmc, barc, cfd and cfa) selected from published SSR maps (Somers et al. 2004; Roder et al. 1998; Kadam et al. 2012; Pestsova et al. 2000; Gupta et al. 2002). Amplification of genomic DNA of parental genotypes and BILs was done with 'BIO RAD T100 Thermal Cycler' using following conditions: Initial denaturation -94°C for 5 min, denaturation -94°C for 1 min, annealing  $-52^{\circ}$ C to  $60^{\circ}$ C (vary with SSR primers used) for 1 min, extension -72°C for 1 min, 35 cycles from denaturation to extension step, final extension -72°C

for 10 min. PCR products were resolved by using 2.5% horizontal agarose gel electrophoresis. Profiles of polymorphic SSR markers were scored visually by coding HD2733 alleles as "A" whereas WH730 alleles were scored as "B". While heterozygote individuals are scored as "H". Missing bands were scored as 'NA'. GGT2 software was used for graphical genotyping of 134 BILs. The construction of linkage map and QTL mapping was performed using software ICIM v4.0 following the default functions available with software. The LOD threshold value of 2.5 with 1000 permutations was used for QTL.

### Results

### Phenotypic performance of BILs

Analysis of variance indicated significant genotypic variance present in the BILs for heat tolerance (Table 1). The phenotypic values for traits under study showed normal distribution pattern. For the nine morphophysiological traits, values of correlation coefficients based on data pooled over LS and VLS conditions are presented in Table 2. Both in the LS and VLS conditions, NDVI was significantly correlated with grain yield and all other traits, except DH, NSPS and SL. The magnitudes of correlation of NDVI with other traits were higher under VLS condition relative to those in the LS condition. NDVI is strongly correlated with TKW under LS (0.61) and VLS (0.67). The regression analysis is performed using yield as an independent variable and all other traits as dependent. Results of regression analysis are presented in (Tables 3a and 3b) for LS and VLS, respectively.

# Molecular characterization of BILs population and genetic linkage map construction

A set of 1350 SSR primers were used to detect polymorphism between parental genotypes, out of which 95 SSRs were polymorphic. Per cent polymorphism between the parents is 9.18. The maximum number of polymorphic markers were found on 2A with nine markers while lowest being two on 6D chromosome. The average numbers of markers per chromosome were 4.52. The linkage map developed using 95 polymorphic markers and 134 BILs population was further utilized to analyze the composition of individual BIL using GGT2 software. The genome of BILs is composed of 63.3 per cent of HD2733 whereas 26.2 per cent of WH730 and remaining is 10.4 per cent, constitute the heterozygous and missing values. The linkage map with identified QTLs in WH730/\*2 HD2733 BILs were presented in Fig. 1.

Trait Source		Trait	Source	s Mean of squa	sum are	Trait	Sources	Mean of squ	sum are	Trait	Sources	Mean of sq	sum uare
	-	LS	VLS			LS	VLS			LS	VLS		
DH	Modal	39.41**	0.0036*	NDVI6	Modal	0.0087**	4.1622	CT6	Modal	3.78**	3.666		
	Rep	11.52	0.0014		Rep	0.0189**	3.7202		Rep	293.4**	3.855		
	Block	7.85	0.0016		Block	0.0035*	4.179		Block	2.896**	2.176		
	treatment	40.10**	0.0044*		treatment	0.0088**	7.5270*		treatment	1.414*	4.041		
NDVI2	Modal	0.0023**	0.0098	CT2	Modal	8.58**	7.428	GWS	Modal	17.0**	55065		
	Rep	0.0006	0.0045		Rep	639.9**	3.985		Rep	76.8**	66735		
	Block	0.0023**	0.0074		Block	14.93**	8.242		Block	4.58*	57447		
	treatment	0.002**	0.0143*		treatment	1.568**	12.390*		treatment	16.62**	55489		
NDVI3	Modal	0.0042**	0.0103*	* СТЗ	Modal	0.0087**	4.1622	TGW	Modal	56.15**	0.0004*		
	Rep	0.0024	0.0080*		Rep	0.0189**	3.7202		Rep	168**	0.0007**		
	Block	0.004**	0.0052		Block	0.0035*	4.179		Block	10.2	0.0003		
	treatment	0.0037**	0.0106*	*	treatment	0.0088**	7.5270*		treatment	54.32**	0.0002		
NDVI4	Modal	0.0078**	0.0104*	* CT4	Modal	8.58**	7.428	GY	Modal	16083**	0.0009**		
	Rep	0.0073*	0.0125*	*	Rep	639.9**	3.985		Rep	55481.**	0.0015**		
	Block	0.0044**	0.0037		Block	14.93**	8.242		Block	7955	0.0005		
	treatment	0.0076**	0.0063*		treatment	1.568**	12.390*		treatment	14922**	0.0003		
NDVI5	Modal	0.0143**	2.682	CT5	Modal	3.060**	5.130*	Biomass	Modal	68409**	26.54		
	Rep	0.0056	3.078		Rep	85.50**	2.863		Rep	413220**	29.38		
	Block	0.0032	2.807		Block	5.946**	6.215*		Block	42028	30.064		
	treatment	0.0151**	4.3060*		treatment	1.531	10.081**		treatment	65213**	10.66		
baratti 1			wmc728_1		barc230	0.00	wmp473.	0.00	aum535	1			
			=				efd23-	-17.50		1			
gym273-	59.40		wmo702		wmc169_1	48.70	wmc285-	-52.40	uma473				
				<b>•</b> • <b>• • •</b>		-81.50	wmc617 2-		and a start of				
emp631-	-118.40		gwm014 134.60 gwm512 144.10	g SL_ia	4 <u>.</u> 24	qDH_ia	ori_SA	q 82_	iari_4D	Ŧ			
umat24	179.15				baro151	-117.10	ofd84	-114.00	baro76 wmo61	2 115.70 84 126.50			
	1000		gwm312		-			183.60					
gdm33_1-	-221.50	Bio_ian_1B	Ŧ				Report of	8	ofd31	1><171.80	eNDWT iari 7D		
wmp387-	-280.80	NDV14_iavi_1B NDV13_iavi_1B	gwm311— — 267 40						wmc702	1			
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				22									

Table 1.	Analysis of varia	ance for	different	morpho-p	physiological	traits u	under l	late sow	n and	very	late	sown	conditio	ns in
	Rabi 2016													

Fig. 1. Genetic linkage map of WH730/\*2 HD2733 BILs population with identified QTLs

**QTL mapping for the heat stress tolerance traits** A total of 9 QTL were detected for five different agronomic traits using Inclusive Composite Interval Mapping (ICIM) under late sown and very late sown

LS	GY	DH	NDVI	СТ	GWS	SL	PS	Biomass	TGW
GY	1								
DH	-0.082	1							
NDVI	0.64**	0.04	1						
СТ	-0.34**	0.05	-0.58**	1					
GWS	0.64**	-0.18*	0.62**	-0.41**	1				
SL	-0.08	0.03	-0.04	-0.10	0.11	1			
NSPS	-0.02	0.05	0.14	-0.16	-0.007	0.42**	1		
Biomass	0.64**	-0.02	0.56**	-0.38**	0.46**	-0.03	-0.09	1	
TKW	0.61**	-0.21*	0.55**	-0.36**	0.86**	-0.02	-0.08	0.43**	1

Table 2(a). Correlation between physiological and yield component traits in BILs population under LS

Table 2(b). Correlation between physiological and yield component traits in BILs population under VLS

LS	GY	DH	NDVI	СТ	GWS	SL	PS	Biomass	TGW
GY	1								
DH	-0.01	1							
NDVI	0.71**	0.005	1						
СТ	-0.57**	0.02	-0.62**	1					
GWS	0.69**	-0.05	0.67**	-0.47**	1				
SL	-0.02	0.02	0.07	-0.07	0.16	1			
NSPS	-0.10	0.16	0.07	-0.007	-0.01	0.31**	1		
Biomass	0.63**	0.11	0.46**	-0.38**	0.35**	0.07	-0.06	1	
TKW	0.67**	-0.11	0.54**	-0.41**	0.82**	0.09	-0.19*	0.24**	1

Table 3(a). Regression analysis using yield asindependent variable and all other traits asdependent variable in LS environments in rabi2016, IARI, New Delhi

Variables	Coefficients	Standard error	t-value	Significance
DH	-0.15	0.70	-0.22	0.83
NDVI	507.32	184.08	2.76	0.01
СТ	48.57	33.53	1.45	0.15
GWS	5.72	2.71	2.11	0.04
SL	-12.88	8.59	-1.50	0.14
NSPS	7.62	10.67	0.71	0.48
Biomass	0.26	0.05	5.27	0.00
TGW	1.61	1.45	1.12	0.27

conditions (Table 4). These were located on five different chromosomes, 1B, 2A, 4D, 5A and 7D. A minimum of 1 QTL were available each for GY, NDVI, DH, SL and biomass and the maximum of 3 QTLs

Table 3(b). Regression analysis using yield asindependent variable and all other traits asdependent variable in VLS environment in rabi2016, IARI, New Delhi

Variables	Coefficients	Standard error	t-value	Significance
DH	-0.16	0.56	-0.29	0.77
NDVI	512.09	157.97	3.24	0.00
СТ	-60.20	30.91	-1.95	0.05
GWS	2.04	2.73	0.75	0.46
SL	-15.62	5.59	-2.79	0.01
NSPS	1.16	3.75	0.31	0.76
Biomass	0.35	0.05	7.71	0.00
TGW	8.02	1.75	4.60	0.00

were available for NDVI *i.e.*, *qNDVI3\_iari\_1* and *qNDVI4\_iari\_1B*, *qNDVI7\_iari\_7D* under late sown and very late sown conditions, respectively. QTLs were identified both in late sown (4 QTLs) and very late

### Sunil et al.

S.No.	Situation	Trait	Chr. No.	Position	Left marker	Right marker	LOD	PVE (%)	Add
1	LS	qDH_iari_5A	5A	116	Xbarc186	Xbarc151	7.11	23.15	2.69
2	LS	qNDVI3_iari_1B	1B	262	Xwmc367	Xwmc728	2.56	8.63	0.005
3	LS	qBio_iari_1B	1B	260	Xgdm33.1	Xwmc367	2.54	8.4	21.8
4	LS	qGY_iari_2A	2A	192	Xgwm388	Xwmc407	3.42	5.37	-38.34
6	VLS	qDH_iari_5A	5A	116	Xbarc151	Xbarc186	8.75	26.42	3.04
7	VLS	qNDVI4_iari_1B	1B	262	Xwmc367	Xwmc728	2.56	8.63	0.005
8	VLS	qNDVI7_iari_7D	7D	176	Xcfd31.1	Xwmc702.1	3.04	10.05	-0.01
9	VLS	qSL_iari_2A	2A	180	Xgwm388	Xwmc407	7.7	10.58	0.27
10	VLS	qSL_iari_4D	4D	94	Xwmc617.2	Xcfd84	5.09	13.29	0.28

Table 4. QTLs detected for various physiological and yield component traits in the WH730/\*2 HD2733 BILs population

Situation: LS (Late sown) and VLS (very late sown), Position of QTL as cM distance from the top of each map., LOD, A LOD threshold of 2.5 was used for declaration of QTL, PVE (%), Phenotypic variance explained by QTL, Additive effect, Positive "additive effect "indicates an increasing effect from HD2733; negative "additive effect" indicates an increasing effect from WH730

sown conditions (5 QTLs). LOD scores for individual QTL ranged from 2.54 to 8.75.

### Discussion

Among the several strategies to overcome the problem of climate change in agriculture, development of stress tolerant genotype/varieties utilizing the molecular resources is important and useful strategy. There are several reports on identification of QTLs responsible for stress tolerance in wheat. Majority of the QTL mapping studies in wheat under heat and drought stress conditions involved mapping populations such as RILs (Mason et al. 2010; Pinto et al. 2010; Mason et al. 2011; Paliwal et al. 2012), doubled haploids (Tiwari et al. 2013) and segregating  $F_2$  and  $F_3$ populations (Gupta et al. 2015). The BILs have rarely been used in QTL mapping experiments across crops (Zhang et al. 2015). In present study, significant parental variation was observed for different morphophysiological traits suggesting ample genetic diversity between the parents. The analysis of variance of different traits showed that most of the traits across the environments considered here were having significant genotypic differences. Phenotypic correlations were estimated using BLUP values of both the environments. DH is used as covariate in BLUP values analysis in order to reduce the variation induced by DH per se among the genotypes. The correlations obtained in present study are in agreement with the previous reports under heat stress (Ramya et al. 2015) where early heading and faster grain filling was observed as critical component traits of heat stress tolerance under changing climatic conditions (Tahmasebi et al. 2016). DH was negatively correlated

with GY and its components traits under heat stress.

The NDVI values tend to decrease as the stress and stage of the plant advances whereas CT increased along the stage and stress on the parents. NDVI is robust trait to be used as criteria for evaluation of genotypes for drought and heat tolerance (Harikrishna et al. 2016; Monostori et al. 2016; Manu et al. 2020). Together, ground cover (leaf area) and the leaf greenness of the canopy can be measured by NDVI (Ramya et al. 2016). It provides an indirect measure of leaf health, which is reflected by leaf nitrogen content associated with photosynthetic efficiency. Few researchers used this trait as proxy trait in drought and heat stress breeding programmes (Araus et al. 2008). NDVI has been found to have positive correlation with yield under both late sown and very late sown conditions in the present study. The magnitude of association increased under VLS condition. In the BILs, differences for NDVI became clearer in later stages of crop growth; this signifies the role of NDVI in heat tolerance. CT is negatively associated with yield and its component traits under stress (Sinha et al. 2018). The magnitude of association was strong in VLS. To characterize the mapping populations and genotypes under heat stress, CT has been used earlier (Tiwari et al. 2012; Reynolds et al. 1997). Cooler canopy temperatures associated with optimal root distribution by which roots proliferate at depth under drought or near to surface under hot, irrigated condition thus maintaining cooler canopies (Pinto et al. 2015). The present study also supports this observation as indicated from the correlation and the regression analysis.

The TKW is well known as proxy trait under heat and drought tolerance and serve as component of heat tolerance (Zarei et al. 2013; Mohammadi et al. 2004; Pinto et al. 2015). Correlations between TKW and yield under heat stress is strong and positive. The positive and strong correlation of GWS and TKW with GY implies its significance in heat stress breeding.

Graphical genotyping clearly reflected the genomic composition of individual BIL and population as whole. The results perfectly matched with the hypothesis in which nearly 75% of the genome of individual line was contributed by HD2733. Mapping QTLs in bi-parental populations allowed the identification of chromosome segments controlling traits of agronomic importance with the opportunity to dissect complex traits into component loci (Marza et al. 2006). During the present study, QTL analysis allowed mapping of as many as 9 QTL (VL and VLS conditions) for physiological and yield related traits suggesting that the genetic control of these traits in wheat is polygenic. This was also supported by the continuous distribution in the component traits of heat tolerance. The findings of earlier studies for identification of QTL also suggested complex genetic control of yield and related traits (McIntyre et al. 2010; Pinto et al. 2010; Bennett et al. 2012a; Bennett et al. 2012b; Kadam et al. 2012; Lopes et al. 2012). Use of physiological traits in combination with QTL mapping provides basic understanding and can serve as useful criterion in selection of heat tolerant genotypes. A Major QTL for DH from the chromosome 5A (*qDH\_iari\_5A*) will be important for wheat improvement for heat stress using MAS, through earliness per se. This QTL was detected both under LS and VLS conditions, suggesting this QTL is more stable and can be used for selection of early genotypes for testing under both conditions. This QTL explained 23-26 per cent of phenotypic variance under LS and VLS conditions however, earlier report on QTL, governing the expression of days to anthesis was associated with Xbarc186, in Seri M82/Babax derived RILs population under heat and drought conditions, which governs early anthesis thereby early maturity explained 6.4 % phenotypic variance (Pinto et al. 2010) in 5A. Zhang et al. (2010) also reported a meta-QTL MQTL4 for yield and related traits in the proximity of Vrn-A1 suggesting the importance of this particular genomic region for traits like days to maturity, GY and other related traits.

The NDVI is an integrative measure of chlorophyll

and total plant biomass, confirmed by a significant positive correlation between NDVIg and Chlg and plant height (Pinto et al. 2016). The QTL qNDVI3\_iari\_1B flanked by markers Xwmc367 and Xwmc728 detected for NDVI3 under LS and NDVI4 under VLS environments was in agreement with a QTL on 1B that explained around 10 % of the phenotypic variance for both rate of senescence and greenness decay reported by Pinto et al. (2016). A QTL qNDVI7\_iari\_7D flanked by cfd31.1 and wmc702.1 on chromosome 7D associated with NDVI at grain filling duration under VLS. Previous studies detected QTL for NDVI on chromosome 7D associated with permanence of greenness under high temperatures (Pinto et al. 2016; Vijavalakshmi et al. 2010; Kumar et al. 2010). Previous studies have documented few QTL for spike length, but none of the studies reported QTLs for spike length on 2A chromosome, suggesting the QTL qSL\_iari\_2A, detected in 2A chromosome flanked by markers Xgwm388-Xwmc407 and with the LOD value 7.7 explaining a phenotypic variance 10.58 %, is putative novel region for spike length under heat stress. Another QTL for SL linked to Xwmc617 in 4D chromosome, the phenotypic variance explained is 13.29 per cent, having the LOD value of 5.09 was also novel to our knowledge. Some previously reported QTL were also confirmed in the present study. The gGY\_iari\_2A is detected in chromosome 2A with a LOD value of 3.4 and phenotypic variance of 5.37. In 2A chromosome, McCartney et al. (2005) reported a QTL for grain yield in DH mapping population derived from RL4452 × 'AC Domain.

## **Authors Contribution**

Conceptualization of research (PKS, GPS NJ); Designing of the experiments (HK); Contribution of experimental materials (PKS); Execution of field/lab experiments and data collection (S, DC, SS, SVP, SK, MB); Analysis of data and interpretation (HK, NJ, SS, RG, DU); Preparation of the manuscript (DU, S, HK, NJ).

### Declaration

The authors declare no conflict of interest.

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### Reference

- Araus J. A., Slafer G. A., Royo C. and Serret M. D. 2008. Breeding for yield potential and stress adaptation in cereals. Crit. Rev. Plant Sci., **27**(6): 377-412.
- Ayeneh A., van Ginkel M., Reynolds M. P. and Ammar K. 2002. Comparison of leaf, spike, peduncle, and canopy temperature depression in wheat under heat stress. Field Crops Res., **79**: 173-184.
- Balla K., Karsai I., Bencze S. and Veisz O. 2012. Germination ability and seedling vigour in the progeny of heat-stressed wheat plants. J. Acta. Agron. Hung., **60**(4): 299-308.
- Bennett D., Izanloo A., Edwards J., Kuchel H., Chalmers K., Tester M., Reynolds M., Schnurbusch T. and Langridge P. 2012a. Identification of novel quantitative trait loci for days to ear emergence and flag leaf glaucousness in a bread wheat (*Triticum aestivum* L.) population adapted to southern Australian conditions. Theor. Appl. Genet., **124**: 697-711.
- Bennett D., Reynolds M., Mullan D., Izanloo A., Kuchel H., Langridge P. and Schnurbusch T. 2012b. Detection of two major grain yield QTL in bread wheat (*Triticum aestivum* L.) under heat, drought and high yield potential environments. Theor. Appl. Genet., **125**: 1473-1485.
- Dhanda S. S. and Munjal R. 2012. Heat tolerance in relation to acquired thermo-tolerance for membrane lipids in bread wheat, Field Crops Res., **135**: 30-37.
- Gupta P., Balyan H. S., Edwards K. J., Isaac P., Korzun V., Gautier M. F., Joudrier P., Schlatter A., Dubcovsky J., De la Pena R. and Khairallah M. 2002. Genetic mapping of 66 new microsatellite (SSR) loci in bread wheat. Theor. Appl. Genet., **105**: 413-422.
- Gupta M., Chawla V., Garg P., Yadav N. R. and Sharma B. 2015. Genetic analysis of yield and heat stress related traits in wheat (*Triticum aestivum* L. em. Thell) using microsatellite markers. J. Nat. Appl. Sci., **7**(2): 739-744.
- Harikrishna, Singh G. P., Jain N., Singh P. K., Sai Prasad S. V., Ambati D., Das T. R., Kumar A., Bhat J. A., Bellundagi A., Vijay Priyanka, Sinha N., Mishra P. C., Misra S. C. and Prabhu K. V. 2016. Physiological characterization and grain yield stability analysis of RILs under different moisture stress conditions in wheat (*Triticum aestivum* L). Indian J. Physiol., **21**: 576-582 DOI: 10.1007/s40502-016-0257-9.
- Jain N., Singh G. P., Singh P. K., Ramya P., Harikrishna, Ramya K. T., Todkar L., Bellundagi A., Kumar Prashant K. C., Vijay P., Jadon V., Dutta S., Rai N., Sinha N. and Prabhu K. V. 2014. Molecular approaches for wheat improvement under drought and heat stress, Indian J. Genet., **74**(4): 578-583.
- Kadam S., Singh K., Shukla S., Goel S., Vikram P., Pawar V., Gaikwad K., Khanna- Chopra R. and Singh N.

2012. Genomic associations for drought tolerance on the short arm of wheat chromosome 4B. Funct. Integr. Genomics, **12**: 447-464.

- Kumar U., Joshi A. K., Kumari M., Paliwal R., Kumar S. and Röder M. S. 2010. Identification of QTLs for stay green trait in wheat (*Triticum aestivum* L.) in the 'Chirya 3'×'Sonalika'population. Euphytica, **174**(3): 437-445.
- Lopes M. S., Reynolds M. P., Jalal-Kamali M. R., Moussa M., Feltaous Y., Tahir I. S. A., Barma N., Vargas M., Mannes Y. and Baum M. 2012. The yield correlations of selectable physiological traits in a population of advanced spring wheat lines grown in warm and drought environments. Field Crops Res., **128**: 129-136.
- Manu B., Kumara P. S., Biradar S., Chauhan D., Phuke R., Ambati D., Prasad S. S., Mishra, P. C., Mishra, K. K., Harikrishna, N. J. and Singh, P.K. 2020. Genetic gain and morpho-physiological characterisation of BILs (Backcross inbred lines) under different moisture regimes in wheat (*Triticum aestivum* L.). Indian J. Genet., 80(1): 84-93.
- Marza F., Bai G. H., Craver B. F. and Zhou W. C. 2006. Quantitative trait loci for yield and related traits the wheat population Ning7840 × Calrk. Theor. Appl. Genet., **112**: 688-698.
- Mason R. E., Mondal S., Beecher F. W., Pacheco A., Jampala B., Ibrahim A. M. and Hays D. B. 2010. QTL associated with heat susceptibility index in wheat (*Triticum aestivum* L.) under short-term reproductive stage heat stress. Euphytica, **174**(3): 423-436.
- Mason R. E., Mondal S., Beecher F. W. and Hays D. B. 2011. Genetic loci linking improved heat tolerance in wheat (*Triticum aestivum* L.) to lower leaf and spike temperatures under controlled conditions. Euphytica, 180(2): 181-194.
- McCartney C. A., Somers D. J., Humphreys D. G., Lukow O., Ames N., Noll J., Cloutier S. and McCallum B. D. 2005. Mapping quantitative trait loci controlling agronomic traits in the spring wheat cross RL4452×'AC Domain'. Genome, **48**(5): 870-883.
- McIntyre C. L., Mathews L., Ratty A., Chapman S. C., Drenth J., Ghaderi M., Reynolds M. and Shorter R. 2010. Molecular detection of genomic regions associated with grain yield and yield- related components in an elite bread wheat cross evaluated under irrigated and rainfed conditions. Theor. Appl. Genet., **120**: 527-541.
- Mohammadi V., Qannadha M. R., Zali A. A. and Yazdi-Samadi B. 2004. Effect of post anthesis heat stress on head traits of wheat. Int. J. Agr. Biol., **6**: 42-44.
- Monostori I., Árendás T., Hoffman B., Galiba G., Gierczik K., Szira F., and Vágújfalvi A. 2016. Relationship between SPAD value and grain yield can be affected by cultivar, environment and soil nitrogen content in wheat. Euphytica, **211**(1): 103-12.

- Nachit M. M. and Elouafi I. 2004. Durum adaptation in the Mediterranean dryland breeding: Breeding, stress physiology, and molecular markers. Crop Sci. Soc. Am. CSSA, **32**: 203-218.
- Ortiz R., Sayre K. D., Govaert B., Gupta R., Subbarao G. V., Ban T. and Reynolds M. 2008. Climate change: Can wheat beat the heat. Agric. Ecosyst. Environ., **126**(1): 46-58.
- Paliwal R., Röder M. S., Kumar U., Srivastava J. P. and Joshi A. K. 2012. QTL mapping of terminal heat tolerance in hexaploid wheat (*T. aestivum* L.). Theor. Appl. Genet., **125**(3): 561-575.
- Pestsova E., Salina E., Börner A., Korzun V., Maystrenko O. I. and Röder M. S. 2000. Microsatellites confirm the authenticity of inter-varietal chromosome substitution lines of wheat (*Triticum aestivum* L.). Theor. Appl. Genet., **101**: 95-99.
- Pinto R. S., Reynolds M. P., Mathews K. L., McIntyre C. L., Olivares-Villegas J. J. and Chapman S. C. 2010. Heat and drought adaptive QTL in a wheat population designed to minimize confounding agronomic effects. Theor. Appl. Genet., **121**(6): 1001-1021.
- Pinto R. S. and Reynolds M. P. 2015. Common genetic basis for canopy temperature depression under heat and drought stress associated with optimized root distribution in bread wheat. Theor. Appl. Genet., **128**(4): 575-585.
- Pinto R. S., Lopes M. S., Collins N. C. and Reynolds M. P. 2016. Modelling and genetic dissection of staygreen under heat stress. Theor. Appl. Genet., **129**(11): 2055–2074.
- Ramya P., Jain N., Singh G. P., Singh P. K. and Prabhu K. V. 2015. Population structure, molecular and physiological characterisation of elite wheat varieties used as parents in drought and heat stress breeding in India. Indian J. Genet., **75**: 250-252.
- Ramya P., Singh G. P., Singh P. K., Jain N., Pandey M. K. and Sharma K. 2016. Effect of recurrent selection on drought tolerance and related morpho-physiological traits in bread wheat. PLoS ONE, **11**(6): e0156869.
- Reynolds M. P., Nagarajan S., Razzaque M. A. and Ageeb O. A. A. 1997. Using canopy temperature depression to select for yield potential of wheat in heat-stressed environment. Wheat Program Special Report, CIMMYT, Mexico, p. 42.
- Reynolds M. P., Hobbs P. R. and Braun H.J. 2007. Challenges to international wheat improvement. J. Agric. Sci.,**145**: 223-227.
- Reynolds M. and Langridge P. 2016. Physiological breeding. Curr. Opin. Plant Biol., **31**:162-171.
- Roder M. S., Korzun V., Wendehake K., Plaschke J., Tixier M. H., Leroy P. and Ganal M. W. 1998. A microsatellite map of wheat. Genetics, **149**(4): 2007-2023.
- Sagar V., Yadav R., Gaikwad K. B. and Gupta S. 2016. Exploring indicator scoring as a selection tool in plant breeding: A study under conservation vs

conventional tillage systems. Indian J. Genet., **76**(3): 266-273.

- Saghai-Maroof M. A., Soliman K. M., Jorgensen R. A. and Allard R. W. 1984. Ribosomal DNA sepacer-length polymorphism in barley: mendelian inheritance, chromosomal location, and population dynamics. Proc. Natl. Acad. Sci., 81: 8014-8019.
- Sinha N., Priyanka V., Ramya K. T., Leena T., Bhat J. A., Harikrishna Jain N., Singh P. K., Singh G. P. and Prabhu K. V. 2018. Assessment of marker-trait associations for drought and heat tolerance in bread wheat. Cereal Res. Commun., 46(4): 639-649.
- Somers D. J., Isaac P. and Edwards K. 2004. A high-density microsatellite consensus map for bread wheat (*Triticum aestivum* L.). Theor. Appl. Genet., **109**: 1105-1114.
- Tahmasebi S., Heidari B., Pakniyat H. and McIntyre C. L. 2016. Mapping QTLs associated with agronomic and physiological traits under terminal drought and heat stress conditions in wheat (*Triticum aestivum* L.). Genome, **60**(1): 26-45.
- Tiwari C., Wallwork H., Dhari R., Arun B., Mishra V. K. and Joshi A. K. 2012. Exploring the possibility of obtaining terminal heat tolerance in a doubled haploid population of spring wheat (*Triticum aestivum* L.) in the eastern Gangetic plains of India. Field Crops Res., **135**: 1-9.
- Tiwari C., Wallwork H., Kumar U., Dhari R., Arun B., Mishra V. K. and Joshi A. K. 2013. Molecular mapping of high temperature tolerance in bread wheat adapted to the Eastern Gangetic Plain region of India. Field Crops Res., **154**: 201-210.
- Vijayalakshmi K., Fritz A. K., Paulsen G. M., Bai G., Pandravada S. and Gill B. S. 2010. Modeling and mapping QTL for senescence-related traits in winter wheat under high temperature. Mol. Breed., 26: 163-175.
- Yang J., Sears R. G., Gill B. S. and Paulsen G. M. 2002. Quantitative and molecular characterization of heat tolerance in hexaploid wheat. Euphytica, **126**: 275-282.
- Zarei B., Naderi A., Jalal Kamali M. R., Lack S. and Modhej A. 2013. Determination of physiological traits related to terminal drought and heat stress tolerance in spring wheat genotypes. Int. J. Agric. Crop Sci., 21(5): 2511-2520.
- Zhang L. Y., Liu D. C., Guo X. L., Yang W. L., Sun J. Z., Wang D. W., and Zhang A. 2010. Genomic distribution of quantitative trait loci for yield and yield-related traits in common wheat. J. Integr. Plant. Biol., 52(11): 996-1007.
- Zhang J., Yu J., Pei W., Li X., Said J., Song M. and Sanogo S. 2015. Genetic analysis of Verticillium wilt resistance in a backcross inbred line population and a meta-analysis of quantitative trait loci for disease resistance in cotton. BMC genomics, **16**(1): 577.