

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/280492635>

Infradian rhythmicity in milk leukocyte activity together with plasma cortisol and prolactin level throughout the lactation period in high yielding crossbred cows

Article in *Biological Rhythm Research* · July 2015

DOI: 10.1080/09291016.2015.1066544

CITATIONS

2

READS

105

5 authors, including:



Joydip Mukherjee

West Bengal University of Animal and Fishery Sciences

70 PUBLICATIONS 272 CITATIONS

[SEE PROFILE](#)



Smrutirekha Mallick

Indian Council of Agricultural Research

11 PUBLICATIONS 41 CITATIONS

[SEE PROFILE](#)



Ajay Kumar Dang

ICAR-National Dairy Research Institute

167 PUBLICATIONS 1,259 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



assessment of early pregnancy by analysis of neutrophils dynamics & gene expression in dairy cattle [View project](#)



New Project [View project](#)

This article was downloaded by: [116.193.130.255]

On: 27 July 2015, At: 09:04

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: 5 Howick Place, London, SW1P 1WG



[Click for updates](#)

Biological Rhythm Research

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/nbrr20>

Infradian rhythmicity in milk leukocyte activity together with plasma cortisol and prolactin levels throughout the lactation period in high-yielding crossbred cows

Joydip Mukherjee^a, Smrutirekha Mallick^b, Mandira Chaudhury^c, B.S. Prakash^d & A.K. Dang^b

^a Department of Veterinary Physiology, WBUAFS, Kolkata 700037, India

^b Dairy Cattle Physiology Division, National Dairy Research Institute, Karnal, Haryana, India

^c A.R.D. Department, Government of West Bengal, Malda District, India

^d ADG (Animal Nutrition and Physiology), Division of Animal Sciences ICAR, Krishi Bhavan, New Delhi, India

Accepted author version posted online: 26 Jun 2015. Published online: 24 Jul 2015.

To cite this article: Joydip Mukherjee, Smrutirekha Mallick, Mandira Chaudhury, B.S. Prakash & A.K. Dang (2015): Infradian rhythmicity in milk leukocyte activity together with plasma cortisol and prolactin levels throughout the lactation period in high-yielding crossbred cows, *Biological Rhythm Research*, DOI: [10.1080/09291016.2015.1066544](https://doi.org/10.1080/09291016.2015.1066544)

To link to this article: <http://dx.doi.org/10.1080/09291016.2015.1066544>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims,

proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

Infradian rhythmicity in milk leukocyte activity together with plasma cortisol and prolactin levels throughout the lactation period in high-yielding crossbred cows

Joydip Mukherjee^{a*}, Smrutirekha Mallick^b, Mandira Chaudhury^c, B.S. Prakash^d and A.K. Dang^b

^aDepartment of Veterinary Physiology, WBUAFS, Kolkata 700037, India; ^bDairy Cattle Physiology Division, National Dairy Research Institute, Karnal, Haryana, India; ^cA.R.D. Department, Government of West Bengal, Malda District, India; ^dADG (Animal Nutrition and Physiology), Division of Animal Sciences ICAR, Krishi Bhavan, New Delhi, India

(Received 15 June 2015; accepted 21 June 2015)

Infradian rhythmicity in milk leukocyte activity together with plasma cortisol and prolactin levels throughout the lactation period in high-yielding crossbred cows has been studied in 10 high-yielding (milk production: 5000 l per lactation) Karan Fries crossbred (Holstein Friesian × Tharparkar) cows. Milk and blood samples were collected from all the experimental animals. Isolation of milk phagocytes (neutrophils and macrophages) and lymphocytes were done by density gradient centrifugation. *In vitro* phagocytic index of milk neutrophils and macrophages was performed by colorimetric NBT reductive assay. Mitogen-induced milk lymphocyte blastogenic response was estimated by colorimetric MTT (tetrazolium) assay. Total plasma cortisol and prolactin were estimated by enzyme immune assay. Highest value of plasma cortisol and prolactin was observed at calving which decreased significantly ($p < 0.01$) on 15th day postpartum for both prolactin and cortisol. Immune activity of milk leukocytes was highest on day 0 colostrum and decreased significantly ($p < 0.01$) on 7th day postpartum. A significant ($p < 0.01$) rise of plasma prolactin was observed around 135th and 225th days postpartum, whereas a peak level of plasma cortisol was observed at 105th, 180th, and 270th days postpartum. Phagocytic index of milk neutrophils and macrophages remains almost in a steady state during mid-lactation period (between 100 and 200 days postpartum). A decline in increasing trend of milk phagocytic activity was observed during late lactation. Mitogen-induced milk lymphocyte blastogenic response was highest on day 0 colostrum which decreased significantly ($p < 0.01$) on 15th day postpartum. Con A-induced milk lymphocyte blastogenic response showed an increasing trend from 120th to 210th days postpartum. Upon correlation study, it showed that the plasma cortisol has a negative effect on milk leukocyte activity, while prolactin has a positive effect, though the effect is lactation stage specific.

Keywords: infradian rhythm; milk leukocytes; cortisol; prolactin; crossbred cows; lactation cycle

1. Introduction

An infradian rhythm refers to the rhythm with a period longer than the period of a circadian rhythm (Lewy et al. 1999). Throughout the production cycle, mammary glands

*Corresponding author. Email: joyphy@gmail.com

of dairy cows undergo marked biochemical, cellular, and immunomodulatory changes to withstand the stress occurred due to partitioning of nutrients between the fetus and the mammary gland during pregnancy, colostrum production, attainment of peak yield, and maintenance of lactation persistency which makes the mammary gland more prone to intra-mammary infections. Circadian rhythmicity in immune system has been reviewed by many workers as lymphocyte proliferation (Ana et al. 1996), activity of natural killer cells (Alvaro & Sarkar 2005) and humoral immune response (Fernandes et al. 1976). Circadian rhythm in mammary tissue carcinogenesis has also been reported (Zhang & Zarbl 2008). Mukherjee et al. (2015) reported seasonal variation in milk leukocyte activity. But, the literature is very scanty on the rhythmic alteration of mammary leukocyte activity throughout the lactation cycle. Therefore, the present investigation has been formulated to find out the infradian rhythmicity in milk leukocyte activity throughout the lactation cycle in high-yielding crossbred cows. An attempt has also been made to correlate the change in milk leukocyte activity with plasma cortisol and prolactin levels throughout the lactation cycle with the assumption of their role in mammary immunity.

2. Materials and methods

2.1. Selection of experimental animals and sampling

Ten high-yielding (milk production: 5000 l per lactation) Karan Fries (KF) crossbred (Holstein Fresian × Tharparkar) cows were selected from the herd of National Dairy Research Institute, Karnal, Haryana, India. All the cows selected were kept in a loose housing system with brick flooring and managed as per the practices followed in the institute. They were offered ad lib green fodder and calculated amount of concentrate mixture based on milk production only at the time of milking. Fresh tap water was available ad lib at all time of the day.

Composite milk samples (250 ml/cow) from all four quarters were collected into sterile tubes fortnightly from the day of calving throughout the lactation cycle (up to 300 days postpartum). Blood samples (10 ml/animal) were drawn in sterile heparinized vacutainer tubes by the jugular vein puncture, posing minimum disturbance to the animal during collection on the same day of milk sampling. Immediately after collecting, the samples were transported to the laboratory in ice for further processing.

Blood samples were subjected to centrifugation (3000 rpm for 30 min) to separate plasma and stored at -20°C for further use.

2.2. Isolation of milk leukocytes

Isolation of milk phagocytes (neutrophils and macrophages) and lymphocytes was done by density gradient centrifugation (Mukherjee et al. 2013).

2.3. In vitro activity of milk leukocytes

In vitro phagocytic index of milk neutrophils and macrophages was performed by colorimetric NBT reductive assay (Choi et al. 2005). Mitogen-induced milk lymphocyte blastogenic response was estimated by colorimetric MTT (tetrazolium) assay (Mosmann 1983).

2.4. Estimation of plasma cortisol and prolactin

Total plasma cortisol was estimated by enzyme immune assay reported by Sarkar et al. (2007). Plasma prolactin was estimated by the methods of Roy and Prakash (2007).

2.5. Statistical analysis

All analyses were done using SYSTAT software package. Data from different experiments are presented as mean \pm SE. Significance was tested by employing one-way ANOVA, considering days as a factor. The correlation was tested by the Spearman rank-order correlation. To avoid the confounding effects of stage of lactation and yield, correlation was calculated during individual stages of lactation cycle i.e. early, mid, and late lactation.

3. Results

Phagocytic activity of milk neutrophils and macrophages with plasma cortisol and prolactin concentration in high-yielding crossbred cows has been presented in Figure 1. Highest value of plasma cortisol and prolactin was observed at calving which decreased significantly ($p < 0.01$) on 15th day postpartum for both prolactin and cortisol. Phagocytic index of milk neutrophils and macrophages was also highest on day 0 colostrum and decreased significantly ($p < 0.01$) on the 7th day postpartum. A significant ($p < 0.01$) rise of plasma prolactin was observed around 135th to 225th days postpartum, whereas a peak level of plasma cortisol was observed on 105th, 180th, and 270th days postpartum. Phagocytic index of milk neutrophils and macrophages remains almost in a steady state during mid-lactation period (between 100 and 200 days postpartum). A decline in the increasing trend of milk phagocytic activity was observed during late lactation.

Mitogen-induced milk lymphocyte proliferation response with plasma cortisol and prolactin concentration in high-yielding crossbred cows has been presented in Figure 2. Plasma cortisol and prolactin levels during different days postpartum have been discussed earlier. Mitogen-induced milk lymphocyte blastogenic response was highest on day 0 colostrum which decreased significantly ($p < 0.01$) on 15th day postpartum. During the rest of the peripartum period, mitogen-induced milk lymphocyte blastogenic response remained unaltered; however Con A-induced milk lymphocyte blastogenic response showed an increasing trend from 120th to 210th days postpartum.

Correlation between immune activity of milk leukocytes and plasma concentration of cortisol and prolactin in high-yielding crossbred cows during different stages of lactation cycle has been presented in Tables 1 and 2, respectively. Phagocytic index of milk

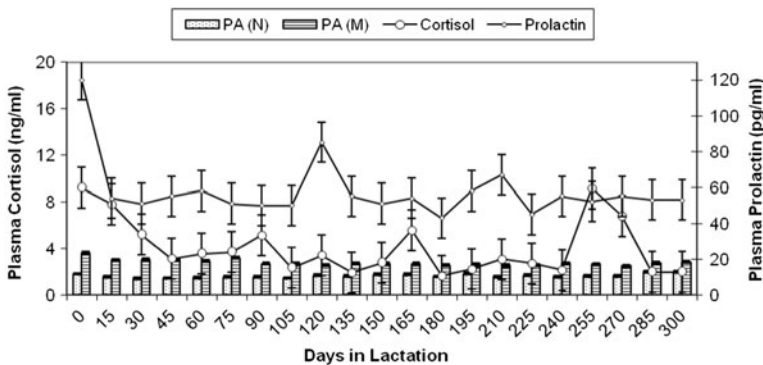


Figure 1. Phagocytic activity of milk neutrophils and macrophages with plasma cortisol and prolactin concentration in high-yielding crossbred cows.

Note: Values are expressed as mean \pm SE.

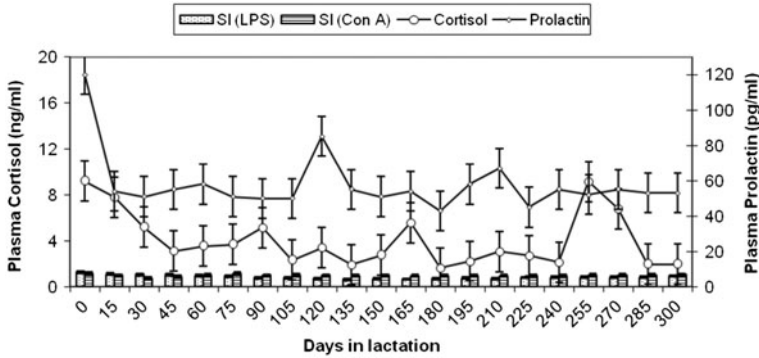


Figure 2. Mitogen-induced milk lymphocyte proliferation response with plasma cortisol and prolactin concentration in high-yielding crossbred cows.

Note: Values are expressed as mean \pm SE.

Table 1. Correlation between immune activity of milk leukocytes and plasma concentration of cortisol in elite crossbred cows during different stages of lactation cycle.

Immune activity of milk leukocytes	Stages of lactation		
	Early	Mid	Late
PI (N)	0.40*	-0.57*	-0.60*
PI (M)	0.31	0.31	-0.46*
SI (Con A)	-0.14	0.38	-0.92*
SI (LPS)	0.35	0.21	-0.52*

*Indicates level of significance ($p < 0.05$).

Table 2. Correlation between immune activity of milk leukocytes and plasma concentration of prolactin in elite crossbred cows during different stages of lactation cycle.

Immune activity of milk leukocytes	Stages of lactation		
	Early (0–100 days)	Mid (100–200 days)	Late (200–300 days)
PI (N)	0.12	-0.57*	0.60*
PI (M)	0.23	0.07	-0.03
SI (Con A)	0.61*	0.09	0.75*
SI (LPS)	-0.14	0.05	0.28

*Indicates level of significance ($p < 0.05$).

phagocytes (neutrophils and macrophages) and lipopolysaccharide (LPS)-induced milk lymphocyte blastogenic response were positively correlated with plasma cortisol concentration during the early lactation period. During mid-lactation, PI of milk neutrophils was negatively correlated ($p < 0.05$) with plasma cortisol concentration. But, PI of milk macrophages and mitogen-induced milk lymphocyte blastogenesis was positively correlated with plasma cortisol concentration. Immune activity of all milk leukocytes was negatively correlated ($p < 0.05$) with plasma cortisol concentration during the late lactation stage.

Phagocytic index of milk neutrophils and macrophages was positively correlated with plasma prolactin during early lactation. However, PI of milk neutrophils was negatively correlated with prolactin concentration, but PI of milk macrophages and mitogen-induced milk lymphocyte blastogenesis was positively correlated during mid-lactation. During late lactation, only PI of milk macrophages was found to be negatively correlated with prolactin concentration, whereas PI of milk neutrophils and mitogen-induced milk lymphocyte blastogenesis was positively correlated with prolactin concentration.

4. Discussion

Neutrophils are the first line of defense of the mammary gland which migrate from blood to the mammary gland in response to inflammatory mediators, namely cytokines, complements, and prostaglandins (Zecconi & Smith 2000; Janeway et al. 2001). In the present investigation, highest phagocytic index of milk neutrophils and macrophages was observed in all the cows, which is in accordance with the earlier reports in buffaloes (Dang et al. 2010). Increased activity of colostrum phagocytes is due to the presence of a low molecular weight substance of less than 10 kDa (Sugisawa et al. 2003). Altered in functional capabilities of mammary gland, phagocytes were observed during the periparturient period which makes the udder vulnerable to infections (Paape et al. 2002) may be due to the impaired functioning of major defense-related activities, such as phagocytosis, respiratory burst activity, superoxide anion production, random cellular migration, and chemotaxis around this period (Kehrli et al. 1989). Further, there was also a decreased expression of adhesion receptor CD62L (L-selectin), which is necessary for penetration through the endothelium to the sites of infection (Lee and Kehrli 1998). Mehrzad et al. (2001) also reported a decreased chemiluminescence response and viability of milk neutrophils between 3 and 11 d postpartum. An increasing trend of milk phagocytic activity was observed during late lactation in our investigation, possibly due to the lower opsonic activity with decrease in IgM concentrations (Waller 2000). Mitogen-induced lymphocyte blastogenic response is used to measure the immune competence of mammary lymphocytes (Mukherjee et al. 2015). In the present investigation, Concanavalin A (Con A) and LPS were used for stimulating T and B lymphocytes, respectively. Lymphocyte proliferation response was higher in colostrum in accordance with the reports of Ogra et al. (1978) in humans. In the present study, Con A-induced milk lymphocyte blastogenic response showed an increasing trend around mid-lactation. Mukherjee et al. (2013) also reported similar findings in crossbred cows. However, Kehrli et al. (1989) reported diminished leukocyte functions during lactogenesis. Decreased milk lymphocyte function during late lactation was also reported by Mukherjee et al. 2013. Glucocorticoids play an important role in initiation and maintenance of lactation (Akers 1990). At the time of parturition, maternal glucocorticoids are significantly elevated in the blood and milk. During this time, cortisol secretion rate and plasma cortisol concentrations are significantly greater when compared with non-lactating cows or cows in established lactation (Paterson & Linzell 1974). Glucocorticoids are necessary for the formation of milk components (Ray et al. 1996) and to induce differentiation of the lobule-alveolar system (Tucker 1981). The value of plasma cortisol concentration observed in our study was similar to that reported by Pal (1996) in crossbred KF cows. In our study, higher level of plasma cortisol was found at calving which decreased significantly on 15th day postpartum. Similar pattern in the shift of cortisol levels immediately after parturition has been reported in exotic (Goff & Horst 1997) and crossbred cows (Prakash & Madan 1985). Khan and Ludri (2002) observed that the

plasma cortisol concentration declined from day 20 to day 2 prepartum, after which it abruptly increased on day 1 prepartum to reach a peak on the day of kidding in goats. The concentration of plasma prolactin in our study was lower when compared to the earlier report of Pal (1996) in crossbred KF cows. We observed large increases in prolactin in peripheral circulation during the day of parturition. In another study where cows were bled twice daily (Ingalls et al. 1971), the peak in prolactin preceded parturition by about 1 day and prolactin returned to prepartum values by 60 h postpartum. In the present study, prolactin peaked at parturition and returned to basal levels within 2 weeks. Whether this parturient peak in prolactin is related to the induction of lactation and/or to parturition as well should be investigated. Possibly, altered prolactin metabolism during lactation, as suggested for pigs (Anderson et al. 1972), masks any active role in reproductive events of the postpartum lactating cow; in heifers, it was observed that prolactin varied with the stages of the estrous cycle (Swanson & Hafs 1971).

Kodagali et al. (1980) reported that the prolactin concentration varied with each stage of lactation, being high, middle, and low in early, mid, and late lactations in Gir cows. Talvelkar and Raikar (1982) reported that prolactin concentrations fell sharply 19 weeks after parturition in Gir cows and 21 weeks after parturition in Gir X HF cross-breeds. They also reported that the stage of lactation significantly correlated with the prolactin concentration. Aliev et al. (1983) reported that the concentration of prolactin was significantly correlated with milk yield in the first three months of lactation in Ala-Tau cows. According to Akasha et al. (1987), serum prolactin concentration declined as lactation progressed. In this investigation, phagocytic index of milk phagocytes and LPS-induced milk blastogenic response in high-yielding cows was negatively correlated with plasma cortisol concentration which is in accordance with earlier reports, which indicated that the glucocorticoids have an inhibitory effect on lymphocyte (Clemens et al. 1979; Pavia et al. 1979; Kincl & Ciaccio 1980) and neutrophil (Nonnecke et al. 1997) functions *in vitro* when administered at pharmacological levels. Roth et al. (1982) reported that the primary anti-inflammatory mechanism of action of glucocorticoids is to prevent neutrophil margination by knocking L-selectin off of neutrophil surfaces (thus reducing neutrophil surveillance for infected tissues) and to potentially reduce the ability of neutrophils to leave the blood stream via decreased expression of CD18 molecules. Increased cortisol during prepartum period suppresses the genes involved in producing inflammatory, antibody, and cytotoxic responses in cattle (Nonnecke et al. 1997). Segei et al. (1980) found inhibition of human lymphocyte blastogenesis by cortisol only when suboptimal mitogen levels were used. The role of prolactin in the immune system has contradictory findings. A series of *in vitro* and *in vivo* studies showed the immunostimulatory effects of prolactin (Gala 1991; Kooijman et al. 1996; Yu-Lee 1997), whereas animals with a targeted disruption of either the prolactin (Horseman et al. 1997) or prolactin receptor (Bouchard et al. 1999) gene suggest that PRL is not essential for normal immune system development or function. In our study, we found lactation stage-specific effect of prolactin over the mammary immunity; this may be due to the fact that prolactin and other pituitary hormones are suggested to act as stress-adaptation molecules important in maintaining steady-state immune system homeostasis (Dorshkind & Horseman 2001). This interpretation is supported by *in vitro* studies showing PRL's protective effect in preventing glucocorticoid-induced lymphocyte cell death (apoptosis) (LaVoie & Witorsch 1995; Buckley 2001) and by *in vivo* studies showing that PRL improves macrophage and splenocyte functions following trauma-hemorrhage and infections (Zellweger et al. 1996).

5. Conclusions

In conclusion, the present study depicts the infradian rhythm of mammary gland immunity together with plasma cortisol and prolactin. Highest activity of milk leukocytes was observed in colostrum together with highest concentration of plasma prolactin and cortisol. A steady-state immune response of mammary gland leukocytes was observed during mid-lactation period which further declined around lactation. Lactation stage-specific effect of plasma cortisol and prolactin was also established.

Acknowledgement

The authors are highly thankful to the Department of Biotechnology, government of India, to provide financial support to carry out the research project.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- Akasha MA, Anderson RR, Eilersieck M, Nixon DA. 1987. Concentrations of thyroid hormone and prolactin in dairy cattle serum and milk at three stages of lactation. *J Dairy Sci.* 70: 271–276.
- Akerv RM. 1990. Lactation physiology: a ruminant animal perspective. *Protoplasma.* 159:96–111.
- Aliev MG, Emelyanova VA, Dyusenbin KD, Kazhmuratova MM. 1983. Relationship between the dynamics of the concentration of hormone in the blood and milk secretion in cows in the course of lactation. *Dokl Vee Aka Sel Nauk.* 2:24–26.
- Alvaro A, Sarkar DK. 2005. Circadian oscillations of clock genes, cytolytic factors, and cytokines in rat NK cells. *J Immunol.* 174:7618–7624.
- Ana IE, Laura S, Agustín A, Valeria DM, Daniel PC. 1996. Twenty-four-hour rhythms in immune responses in rat submaxillary lymph nodes and spleen: effect of cyclosporine. *Brain Behav Immun.* 10:92–102.
- Anderson LL, Peters JB, Melampy RM, Cox DF. 1972. Changes in adenohipophysial cells and levels of somatotrophin and prolactin at different reproductive stages in the pig. *Reproduction.* 28:55–65.
- Bouchard B, Ormandy CJ, Di Santo JP, Kelly PA. 1999. Immune system development and function in prolactin receptor-deficient mice. *J Immunol.* 163:576–582.
- Buckley AR. 2001. Prolactin, a lymphocyte growth and survival factor. *Lupus.* 10:684–690.
- Choi EM, Kim A-J, Kim Y-O, Hwang J. 2005. Immunomodulating activity of arabinogalactan and fucoidan *in vitro*. *J Med Food.* 8:446–453.
- Clemens LE, Siiteri PK, Stites DP. 1979. Mechanism of immunosuppression of progesterone on maternal lymphocyte activation during pregnancy. *J Immunol.* 122:1978–1985.
- Dang AK, Mukherjee J, Kapila S, Mohanty AK, Kapila R, Prasad S. 2010. *In vitro* phagocytic activity of milk neutrophils during lactation cycle in Murrah buffaloes of different parity. *J Anim Physiol Anim Nutr.* 94:706–711.
- Dorshkind K, Horseman ND. 2001. Anterior pituitary hormones, stress, and immune system homeostasis. *BioEssays.* 23:2881793–2941803.
- Fernandes G, Halberg F, Yunis EJ, Good RA. 1976. Circadian rhythmic plaque-forming cell response of spleens from mice immunized with SRBC. *J Immunol.* 117:962–966.
- Gala RR. 1991. Prolactin and growth hormone in the regulation of the immune system. *Exp Biol Med.* 198:513–527.
- Goff JP, Horst RL. 1997. Physiological changes at parturition and their relationship to metabolic disorders. *J Dairy Sci.* 80:1260–1268.
- Horseman ND, Zhao W, Montecino-Rodriguez E, Tanaka M, Nakashima K, Eagle SJ, Smith F, Markoff E, Dorshkind K. 1997. Defective mammopoiesis, but normal hematopoiesis, in mice with a targeted disruption of the prolactin gene. *EMBO J.* 16:6926–6935.

- Ingalls W, Hafs HD, Oxender WD. 1971. Growth hormone, prolactin, and luteinizing hormone in heifers before and after parturition. *J Dairy Sci.* 54:768 (Abstr.).
- Janeway CA, Travers P, Walport M, Shlomchik M. 2001. Basic concepts in immunology. In: Janeway CA, Travers P, Walport M, Shlomchik M, editors. *Immunobiology: the immune system in health and disease*. New York, NY: Garland Publishing; p. 1–34.
- Kehrli ME, Nonnecke BJ, Roth JA. 1989. Alterations in bovine lymphocyte function during the periparturient period. *Am J Vet Res.* 50:215–220.
- Khan JR, Ludri RS. 2002. Hormone profile of crossbred goats during the periparturient period. *Trop Anim Health Prod.* 34:151–162.
- Kincl FA, Ciaccio LA. 1980. Suppression of immune responses by progesterone. *Endocrinol Exp.* 14:27–33.
- Kodagali SB, Deshpande BR, Sane CR, Sheth AR, Shah GV, Gadgil BA. 1980. Serum prolactin levels of postpartum Gir cows. *Indian J Dairy Sci.* 32:185–187.
- Kooijman R, Hooghe-Peters EL, Hooghe R. 1996. Prolactin, growth hormone, and insulin-like growth factor-1 in the immune system. *Adv Immunol.* 63:377–454.
- LaVoie HA, Witorsch RJ. 1995. Investigation of intracellular signals mediating the anti-apoptotic action of prolactin in Nb2 lymphoma cells. *Exp Biol Med.* 209:257–269.
- Lee EK, Kehrli ME Jr. 1998. Expression of adhesion molecules on neutrophils of periparturient cows and neonatal calves. *Am J Vet Res.* 59:37–43.
- Lewy H, Naor Z, Ashkenazi IE. 1999. From ultradian to infradian rhythms: LH releases patterns *in vitro*. *Chronobiol Int.* 16:441–450.
- Mehrzaad J, Dosogne H, Meyer E, Burvenich C. 2001. Local and systemic effects of endotoxin mastitis on the chemiluminescence of milk and blood neutrophils in dairy cows. *Vet Res.* 32:131–144.
- Mosmann T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods.* 65:55–63.
- Mukherjee J, De K, Chaudhury M, Dang AK. 2015. Seasonal variation in *in vitro* immune activity of milk leukocytes in elite and non-elite crossbred cows of Indian sub-tropical semi-arid climate. *Biol Rhythm Res.* 46:425–433. doi:10.1080/09291016.2015.1020200.
- Mukherjee J, Varshney N, Chaudhury M, Mohanty AK, Dang AK. 2013. Immune response of the mammary gland during different stages of lactation cycle in high versus low yielding Karan Fries crossbred cows. *Livest Sci.* 154:215–223.
- Nonnecke BJ, Burton JL, Kehrli MR Jr. 1997. Associations between function and composition of blood mononuclear leukocyte populations from holstein bulls treated with dexamethasone. *J Dairy Sci.* 80:2403–2410.
- Ogra SS, Weintraub DI, Ogra PL. 1978. Immunological aspects of human colostrum and milk: interaction with the intestinal immunity of the neonate. *Adv Exp Med Biol.* 107:95–103.
- Paape MJ, Mehrzaad J, Zhao X, Detilleux J, Burvenich C. 2002. Defense of the Bovine mammary gland by polymorphonuclear neutrophil leukocytes. *J Mamm Gl Biol Neopl.* 7:109–121.
- Pal Y. 1996. Circulatory levels of some hormones and metabolites during initiation and early lactation in crossbred cows and buffaloes. [PhD thesis]. Karnal: National Dairy Research Institute.
- Paterson JYF, Linzell JL. 1974. The secretion of cortisol and its mammary uptake in the goat. *J Endocrinol.* 50:493–499.
- Pavia C, Siiteri PK, Perlman JD, Stites DP. 1979. Suppression of murine allogeneic cell interactions by sex hormones. *J Reprod Immunol.* 1:33–38.
- Prakash BS, Madan ML. 1985. Periparturient plasma progesterone and prostaglandin F (PGF) levels in Karan Swiss cows. *Indian J Anim Sci.* 55:642–646.
- Ray DW, Davis JR, White A, Clark AJ. 1996. Glucocorticoid receptor structure and function in glucocorticoid-resistant small cell lung carcinoma cells. *Cancer Res.* 56:3276–3280.
- Roth JA, Kaerberle ML, Hsu WH. 1982. Effect of glucocorticoids on the bovine immune system. *J Am Vet Med Ass.* 180:894–901.
- Roy KS, Prakash BS. 2007. Development and validation of a simple sensitive enzyme immunoassay (EIA) for quantification of prolactin in buffalo plasma. *Theriogenology.* 67:572–579.
- Sarkar M, Das BC, Bora B, Kumar V, Mohan K, Meyer HHD, Prakash BS. 2007. Application of sensitive enzyme immunoassay for determination of cortisol in blood plasma of yaks (*Poephagus grunniens* L). *Gen Comp Endocrinol.* 154:85–90.

- Segei GB, Lukacher A, Gordon BR, Lichtman MA. 1980. Glucocorticoid suppression of human lymphocyte DNA synthesis: influence of phytohemagglutinin concentration. *J Lab Clin Med.* 95:624–632.
- Sugisawa H, Itou T, Saito M, Moritomo T, Miura Y, Sakai T. 2003. A low-molecular-weight fraction of bovine colostrum and milk enhances the oxidative burst activity of polymorphonuclear leukocytes. *Vet Res Comm.* 27:453–461.
- Swanson LV, Hafs HD. 1971. LH and prolactin in blood serum from estrus to ovulation in Holstein heifers. *J Animal Sci.* 33:1038–1041.
- Talvelkar BA, Raikar RK. 1982. Correlation of serum LH and serum prolactin levels during post partum period in purebred Gir and crossbred cows. *Indian J Dairy Sci.* 35:505–510.
- Tucker HA. 1981. Physiological control of mammary growth, lactogenesis, and lactation. *J Dairy Sci.* 64:1403–1421.
- Waller KP. 2000. Mammary gland immunology around parturition. Influence of stress, nutrition and genetics. *Adv Exp Med Biol.* 480:231–245.
- Yu-Lee Y-L. 1997. Molecular actions of prolactin in the immune system. *Proc Soc Exp Biol Med.* 215:35–52.
- Zecconi A, Smith KL, editors. 2000. IDF Position Paper on Ruminant Mammary Gland Immunity. Symposium on Immunology of Ruminant Mammary Gland. Stresa, Italy. pp. 1–120.
- Zellweger R, Zhu X-H, Wichmann MW, Ayala A, DeMaso CM, Chaudry IH. 1996. Prolactin administration following hemorrhagic shock improves macrophage cytokine release capacity and decreases mortality from subsequent sepsis. *J Immunol.* 157:5748–5754.
- Zhang X, Zarbl H. 2008. Chemopreventive doses of methylselenocysteine alter circadian rhythm in rat mammary tissue. *Cancer Prev Res (Phila).* 1:119–127.