Fishery Technology 2006, Vol. 43 (2) pp : 180 - 185

Effect of Chlorine on the Survival of Vibrio cholerae on Shrimp

Nirmala Thampuran, K.Sreeganga*, and P.K.Surendran**

Microbiology, Fermentation and Biotechnology Division, Central Institute of Fisheries Technology Cochin –29. India

Shrimp is a an important commodity in international trade and stringent quality parameters stipulates zero tolerance of *Vibrio choleare* in the imported commodity. Sanitary requirements for production of good quality shrimp for processing and export stipulates that shrimp tissue is to decontaminated from pathogens like *Vibrio cholerae* by washing with potable water carrying chlorine. Hence effective dose of chlorine to be used in the process water is very critical. The present study was aimed at assessing the microbicidal efficacy of chlorine against *Vibrio cholerae* cells in water and shrimp with and without shell. The study indicated that in potable water with 2ppm residual chlorine level, a population of 10 ⁷ cfu of *Vibrio cholerae* could be destroyed after 30 min exposure. Studies on shrimp meat contaminated with varying levels of *Vibrio cholerae* exposed to chlorine showed that 4ppm chlorine could effect complete destruction of a population of 10³ cfu /ml in 10min. On headless shrimp with shell-on, 7ppm chlorine was required to destroy 10³ cells /g of *V.cholerae* within 10 min. The data indicate that chlorine level has to be adjusted depending on nature of material and *Vibrio cholerae* present in shell-on shrimp need special attention.

Key words: Vibrio cholerae, shrimp, chlorine, survival

Toxigenic *Vibrio cholerae* O 1 and 139 are the causative agents of cholera, a water-borne and food borne disease with epidemic and pandemic potential. Various foods have been frequently implicated in the outbreak of cholera which includes milk and milk products, meat, vegetables, fruits and seafood (Tauxe *et al.*,1992). Outbreaks of cholera due to consumption of seafood, oyster, crab and shrimp have also been reported. (Oliver & Kaper ,1997).

V.cholerae may be considered as an indigenous member of estuarine / brackish water bacterial community with capability to

survive long durations in adverse conditions (Singleton et al.,1982). It is more prevalent in aquatic systems during warm climates (Isalm et al 1992) but the pathogen can also survive in frozen shrimp for long duration (Wong et al., 1995). Vibrio cholerae occur not only in coastal waters and shrimp from marine environments. (Depaola, 1981), but also in cultured fish (Fernandes et al.,1997), cultured freshwater shrimp, Macrobrachium rosenbergii (Jayasekaran & Ayyappan, 2002) and brackish water prawns (Reilly & Twiddy,1992). Fish and fishery products meant for export in India are found to carry both Vibrio cholerae O1 and non O1 (Varma

^{*} Aiswarya, P.O Nirmalagiri, Kannur 670 701, ** Poothuvallil, Dr Surendran lane, Perumpadappu, Palluruthy P.O, Cochin 682006

The study was designed with following steps. To determine the effect of chlorine on *V*. cholerae suspended in potable water, one ml of Vibrio cholerae cell suspension of known cell densities were added to 100ml potable water containing varying levels of available chlorine in beaker under sterile condition. Immediately after the addition and at intervals of 5 min, 1ml aliquot was withdrawn and the excess chlorine was inactivated by adding to 10 ml of 2% Sodium thiosulphate. The viable count was determined immediately by spread plate method in tryptone glucose yeast extract agar TSYA (Oxoid) plates. Colonies of surviving bacteria were counted after incubation of the plates at 37°C for 24h. Trials were carried out separately for various cell densities (103 to 107 cells /ml) and chlorine levels (3-10 ppm).

In the case of shrimp, PD and HL, the experiment was repeated in the same way with following modifications. 1ml of V.cholerae cell suspension of varying cell densities as given above were added to 250 g each of HL and PD shrimp suspended in 1000ml chlorinated potable water in sterile condition as before. Shrimp (25kg) immersed in 100ml chlorine-free distilled water served as control. Immediately after the addition and at 5 min. intervals, 25 g lot of shrimp was taken out and the chlorine was neutralized by dipping in 100ml of 2% Sodium thiosulphate. Shrimp was then blended in 225 ml of saline, serially diluted and viable count was estimated by spread plating onto TSYA plates. Typical Colonies of surviving population were counted after incubation of the plates at 37° caviae for 24h. Since the background flora is destroyed by heat treatment, the survivors are considered to be V cholerae only. Occasionally about 5% of colonies from a plate were isolated and confirmed as Vibrio cholerae by standard protocol (Anon,2003)

Results and Discussion

Table 1 shows survival of V.cholerae when suspended in potable water with varying chlorine concentrations. At 2ppm residual level, no survivors could be noticed at any of cell densities after 30 min exposure. At 10 ppm level 5 min exposure was sufficient to completely destroy the peak level of 107 cells /ml. An earlier study on the effect of chlorine on V.cholerae has recommended chlorine level of 10 ppm and contact time of 5 min for total eradication of 1.80 x 106 cells ml. of V. cholerae in water (Iyer & Varma., 1991). Sousa et al (2001) reported that an initial density of 107cfu/ml. V cholerae in water suspension resulted in 100% reduction after 5 min exposure to 7 ppm chlorine.

Table 1 Viability of *V cholerae* in potable water after exposure to varying chlorine levels and exposure times

levels and exposure times					
Cell	ell Survival(%) of Vibrio cholerae at			at	
density	different residual chlorine levels			ls	
	2ppm	3ppm	4ppm	5ppm	10ppm
1.68x10 ⁷	0.08	0.05	0.002	0.001	ND
1.68x10 ⁷	0.001	0.01	0.001	ND	ND
1.68x10 ⁷	0.001	0.001	ND	ND	ND
1.68x10 ⁷	ND	ND	ND	ND	ND
	Cell density 1.68x10 ⁷ 1.68x10 ⁷ 1.68x10 ⁷	Cell density 2ppm 1.68x107 0.08 1.68x107 0.001 1.68x107 0.001	Cell density Surviva different 2ppm 3ppm 1.68x107 0.08 0.05 1.68x107 0.001 0.01 1.68x107 0.001 0.001	Cell density Survival(%) of Vil different residual cl 2ppm 3ppm 4ppm 1.68x107 0.08 0.05 0.002 1.68x107 0.001 0.01 0.001 1.68x107 0.001 0.001 ND	Cell density Survival(%) of Vibrio cholerae different residual chlorine leve 2ppm 3ppm 4ppm 5ppm 1.68x107 0.08 0.05 0.002 0.001 1.68x107 0.001 0.01 0.001 ND 1.68x107 0.001 0.001 ND ND

ND - Not Detected

Table 2. Survival of *V cholerae* on shrimp (PD) after various exposure times and chlorine levels

chlorine ppm	Number of cells	Survival Rate % Range		Survival Rate % Mean*	
	Exposedto Chlorine/ml.	5 min.	10 min.	5 min.	10 min.
2	1.28x10 ³	2.88-3.38	1.36-1.80	2.98	1.52
3	1.28x10 ³	2.11-2.40	1.48-1.77	2.29	1.62
4	1.28x10 ⁵	0.42 - 0.98	0	0.62	0
5	1.28x10 ³	0 - 0.20	0	.13	0
6	1.28x10 ³	0	0	0	0
4	1.28x10 ⁶	.11-0.17	0.2-0.4	0.14	0.03

* Mean of 3 experiments

The survival of V. cholerae associated with shrimp meat without shell is presented in Table2. When shrimp meat contaminated to a level of 10^3 to 10^6 cells /ml were exposed to chlorine, 4ppm chlorine could cause complete reduction of a population of 10^3 cfu /ml in

10min.When 10° cfu/ml were treated with 4ppm chlorine for 10 mints survival rate was 0.03%. This indicates that potable water for washing the shrimp meat carrying 10° cfu/ml *V cholerae* should carry 4 ppm of residual chlorine with contact time 10 min. Thus survival rate varies with number of cells exposed and the activity of chlorine.

Table 3. Survival of $\ V\ cholerae$ on shrimp (HL) after various exposure times and chlorine levels

chlorine Cell		Survi	Survival Rate % Mean*		
ppm density		%R			
	cfu/ml.	5 min.	10 min.	5 min.	10 min.
2	1.28x10 ³	100	42.44-49.50	100	46.22
3	1.28x10 ⁵	100	22.37-36.62	100	28.40
4	1.21x10 ³	22.1-27.2	8.06-9.11	24.20	8.70
5	1.28x10 ⁵	19.40-23.50	5.33-6.12	21.70	5.70
6	1.21x10 ³	6.80-18.20	1.63-1.83	17.40	1.70
7	1.28x10 ⁵	1.40-2.00	0	1.70	0
8	1.28x10 ³	1.60-1.90	0	1.70	0
9	1.28x10 ³	0	0	0	0
7	1.28x10 ⁵	8.10-8.30	3.62-4.12	8.23	3.89

[·] Mean of 3 experiments

Table 3 shows the efficiency of chlorine on V cholerae present on headless shrimp with shell on. As evident from the data, 7ppm chlorine completely destroy 103 cells /g of V.cholerae within 10 mints. The chlorine level to bring out a similar effect on shrimp meat was only 4ppm. This clearly indicate that a higher chlorine concentration is required to treat shrimp with shell. At low chlorine levels of 2 and 3 ppm, the survival pattern for shrimp with and without shell was almost similar both in PD and HL shrimp and had very little bactericidal effect. After 4ppm chlorine ,there was striking decline in cell populations and more than one log reduction from the initial count was noted

Sousa (2001) reported a very slow decrease in *V cholerae* cell population with increase in chlorine concentration and attributed this to the capacity of bacterium to adhere tightly to chitin

exoskeleton and escape from chlorine. Presence of excess organic matter(Alcamo,1994) protection afforded by chitin exoskeleton (Liu and Xie., 1994), the decomposition products from chitin (Amako et al., 1987) and greater adherence to shell (Pruzzo et al., 1996) have been suggested as reason for the prolonged survival of V cholerae on shrimp with shell-on. The physical protection offered by hollow spaces in the carapace (Nascimento et al., 1998) can also be an added reason. Apart from these, activity of any disinfectant may be greatly affected by the factors such as dilution, temperature, pH or the presence of organic matter. Hence the combination of all these factors could be responsible for the survival of V cholerae on whole shrimp.

In normal conditions, we do not expect any V cholerae cells in raw shrimp. Assuming that even in extreme situations, the maximum V cholerae count on shrimp may be 100 to 1000 cfu / g , the chlorine levels recommended in this study can ensure protection against survival or growth of V cholerae.. The study clearly shows that 2ppm chlorination levels used at present is insufficient to completely eradicate viable V cholerae present on shrimp although it may successfully lessen the external bacterial contamination. If safety of the consumer from the pathogen is the prime concern of the producer or buyer, disinfecting shrimp with appropriate dosage has to be strictly followed. By using low chlorine level, the surface contamination can be reduced, but pathogens may persist and pose health risk in later stages.

Development of chlorine resistant form of *V cholerae* has been reported previously (Morris *et al.*, 1996; Rice *et al.*, 1993) and such forms were noticed during this study also. Longer contact time of *V cholerae* with carapace

results in greater resistance and development

of rugose forms (Sousa et al., 2001). These

chlorine resistant forms can also be a problem

in future years if adequate chlorine levels are

provided and also for the permission to publish this

manuscript. This work has been carried out as part of

Authors are thankful to Director, Central Institute of Fisheries Technology, Cochin 29 for the facilities

not maintained.

prawn

NATP programme and the financial assistance provided by them is gratefully acknowledged. References Alcamo I.E.(1994) Chemical control of micro organism, In Fundamental Microbiology, 4th edn, p.665-687, The Benjamin/ Cummings Publishing, New York. Anon.(2001) Document No EIC / F& FP / Ex /

Anon (2002) Risk assessment of Campylobacter in broiler chickens and Vibrios in seafood, 59p,Report of the Joint FAO/WHO Expert Consultation, Bangkok, Thailand, August 2002. Anon (2003) Laboratory Techniques for Microbiological Examination of Seafood (Surendran P.K., Nirmala Thampuran., Narayanan Nambiar, V., & Lalitha, K.V Eds.), Central Institute of

Inst. / July 2001/ Issue 2 Export

Inspection Council of India, New Delhi.

Fisheires Technology, Cochin -682029, India. APHA (1998) Standard Methods For Examination Of Water and Waste Water, 20th edn., American Public Health Association, Washington D.C, pp 455-456. Dalgaard,P (1995) Prevalence of Vibrio cholerae and Salmonella in major shrimp production Thailand. area in International J Food Microbiol. 28, pp 101-

113

Fernandes, C.F., Flick J.R.Jr., Silva, J.L., and Mc caskey, T.A. (1997) Comparison of quality in aqua cultured fresh water cat fish II- pathogens E.coli O157 H7 Campylobacter, Vibrio Plessiomonas and Klebsiella., J. Food Prot. 60, pp 1182-1188 Huss, H.H., Ababouch, L. and Gram, L (2003) Assessment and Management of Seafood Quality, FAO Fisheries Technical Paper No 444, Rome, FAO, Food Agriculture Organization, Geneva, 230 p. Islam, M.S., Alarm, M.J. and Neogi P.K.B (1992) Seasonality and toxigenicity of Vibrio

Depaola, A.(1981) Vibrio cholerae in marine foods

and environmental waters: Literature

review. J. Food Science. 46, pp 66-70.

cholerae non o1 isolated from different components of pond eco system of Dhaka city Bangladesh. World J.Microbio. & Biotechnol. 8, pp 160-163. Iyer, T.S.G. and Varma, P.R.G. (1991) Viability of Vibrio cholerae in water subjected to different levels of chlorination Fish Tech. 28, pp 158-159. Jayasekaran, G. and Ayyappan S. (2002) Post harvest microbiology of farm reared tropical fresh water

Macrobrachium rosenbergii J.Food Sci. 67, pp 1859-61. Liu, S.G. and Xie, Z.H. (1994) Protection of Vibrio cholerae from heating and chlorination by chitin., Lancet 1, pp 16-19. Morris, J.G., Szetein, M.B., Rice E.W., Nataro J.P., Losonsky, G.A., Panigrahi, P., Tacket, C.O. and Johnson J.A. (1996) Vibrio cholerae 01 can assume chlorine resistant rugose survival form that is

pp 1368-1464.

virulent for humans J.Infect.Dis. 174,

1320.

Washington D.C, pp 228-264. Pruzzo,, C., Crippa, A., Betone, S., Pane, L.. and Carli, A. (1996) . Attachment of Vibrio alginolyticus to chitin mediated by chitin-binding proteins. Microbiol. 142,

Reilly, P.J.A. (2000) Discussion paper on the use

of chlorinated water. Code of practice for

fish and Fishery Products CX/FFP/00/

13 Food and Agriculture Organization

/ World Health Organization , Rome ,

pp 2181-2186.

Nascimento, A.R,., Viera, R.H.S.F., Almieda,

Oliver, J.D. and Kaper, J.B.(1997) Vibrio species

H.B., Patel, T.R. and Iaria, S.T. (1988)

Survival of V.cholerae 01 strains in

shrimp subjected to freezing and

boiling J.Food Protection 61, pp 1317-

in Food Microbiology; Fundementals and

Frontiers (M.P Doyle ,L.R .Beuchat &

T.J Montville Eds.) ASM Press,

Italy. Reilly, P.J.A. and Twiddy D.R. (1992) Salmonella and Vibrio cholerae in brackish water cultured tropical prawn International J.Food Microbiol. 16, pp 293-301.

Rice, E.W., Clark, R.M., Fox, K.R., Reasoner, D.J., Dunnigan, M.E., Panigrahi, P., Johnson, J.A and Moriis J.G. (1993)

Vibrio cholerae o1 can assume a rugose

Int.J.Environ.Health Res. 3, pp 89 - 98. Roberts, D. (1992) Growth and survival of Vibrio cholerae in foods PHLS Microbiology Digest, 9, pp 24-31

resistant survival form that resist

killing by chlorine yet retains virulence.

R.R. (1982) Effect of temperature and

salinity on Vibrio cholerae growth. Appl.

Environ. Microbiol. 44, pp 1047-1058.

Singleton, F.L., Atwell, R., Jangi, S. and Colwell,

Geneva, Switzerland.

Wong, H.C., Li Li Chen and Yu, C.M. (1995) J.Food Prot., 58, pp 263-267.

Sousa ,O.V., Viera, R.H.S.F., Patel, T.R., Hoefer, E. and Mesquita, V.P. (2001) Food Microbiol. 18, pp 355-359. Tauxe, R.V. and Blake, P.A. (1992) Epidemic

cholerae in Latin America. J. American Medical Association.276, pp 1388-1390. Varma, P.R.G., Iyer, T.S.G., Joseph, M.A. and Zacharia, S. (1989) Studies on the incidence of Vibrio cholerae in fishery

products J. Food Sci. & Tech. 26, pp 341-

342. WHO.(1996) Guidelines for Drinking Water Quality Health Criteria and Other Supporting Information . 2nd edn, Vol 12, World Health Organization,