



# Photosynthetic characteristics of peanut genotypes under excess and deficit irrigation during summer

Kuldeepsingh A. Kalariya<sup>1</sup> · Amrit Lal Singh<sup>2</sup> · Nisha Goswami<sup>2</sup> · Deepti Mehta<sup>2</sup> · Mahesh Kumar Mahatma<sup>2</sup> · B. C. Ajay<sup>2</sup> · Koushik Chakraborty<sup>2</sup> · P. V. Zala<sup>2</sup> · Vidya Chaudhary<sup>2</sup> · C. B. Patel<sup>2</sup>

Received: 23 December 2014 / Revised: 3 April 2015 / Accepted: 19 April 2015 / Published online: 15 May 2015  
© Prof. H.S. Srivastava Foundation for Science and Society 2015

**Abstract** In a field experiment three irrigation treatments were given to twelve peanut genotypes through drip. At 80 days after sowing (DAS) the amount of irrigation applied was 20 % higher than the evaporative demand (ET) in T<sub>1</sub>, 25 % less than ET in T<sub>2</sub> and 48 % less than ET in T<sub>3</sub> against the cumulative evaporative demand of 412 mm. The relative water content (RWC) of peanut leaves reduced by cutting irrigation from 93.5 % in T<sub>1</sub> to 91.1 % in T<sub>2</sub> and 77.2 % in T<sub>3</sub> but, net photosynthetic rate ( $P_N$ ) was higher in T<sub>2</sub> ( $29.6 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) than T<sub>1</sub> ( $28.6 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) and T<sub>3</sub> ( $24.3 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) at 75–80 DAS. Peanut genotype ICGV 91114 showed the highest  $P_N$  ( $30.9 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) which was statistically at par with GG 20, ICGV 86590, TAG 24, SB XI, TMV 2 and TPG 41. The non-photochemical quenching ( $NPQ$ ) varied with different irrigation treatment with lowest in T<sub>2</sub> and highest in T<sub>3</sub>. The de-epoxidation state ( $DeS$ ) was 38 % in T<sub>1</sub> and T<sub>2</sub> but, increased to 47 % in T<sub>3</sub> due to the severe water deficit stress. Applying 20 % higher irrigation than the ET demand (T<sub>1</sub>) does not warrant any extra benefits in terms of higher photosynthesis in peanut at 75–80 DAS. Further, a reduction of 25 % of the ET (T<sub>2</sub>) in peanut seems to be the ideal condition for photosynthesis and desirable chlorophyll fluorescence parameters at 80 DAS. Gimar 3 and ICGV 91114 showed  $NPQ$  value above 2.2 and higher de-epoxidation state, maintained least deviation in  $F_v/F_m$  and  $F_v'/F_m'$  under severe water deficit condition are promising peanut genotypes.

**Keywords** Chlorophyll fluorescence · Irrigation · Peanut · Photosynthesis · Xanthophylls

## Introduction

Peanut (*Arachis hypogaea* L.) is an important food legume as well as an oilseed crop being grown in 112 countries of the world on about 25 million ha of land. The production of peanut is about 41.1 m ton and is grown mostly in tropics and subtropics of arid and semi-arid regions where the availability of water is a major constraint. Frequent drought of various spells and intensities in these areas results in the productivity of peanut being less than 1000 kg ha<sup>-1</sup> in more than 35 % of peanut growing countries. About 5.7 million ha in India is under the cultivation of peanut with productivity of about 1300 kg ha<sup>-1</sup>. Due to suitable environment and photoperiod matching to the growing season, the commercial peanut cultivation is possible mainly in Asia (47 % of the world peanut cultivation area contributing 60 % of the total world production), Africa (47 % area, 27 % production) and America (4.4 % area and 8 % production) (FAO 2012).

Photosynthesis is the most important process influencing crop production and the high  $P_N$  is one of the most important breeding strategies for crop improvement (Richards 2000). The simultaneous occurrence of water deficit stress coupled with heat and high irradiance leads to severe photo-oxidative damage to the photosynthetic apparatus which often aggravates the amount of excess excitation energy and this excess excitation energy, when not dissipated harmlessly, would be transformed to O<sub>2</sub> to form reactive oxygen species (ROS) which could damage the photosynthetic apparatus. Plants have developed many protective mechanisms to balance absorbed light energy with photosynthesis; the most important one is the xanthophyll cycle-dependent thermal energy dissipation, measured as the non-photochemical quenching ( $NPQ$ ) of chlorophyll fluorescence. Chlorophyll *a*

✉ Kuldeepsingh A. Kalariya  
kuldeep\_ka@yahoo.co.in

<sup>1</sup> Indian Council of Agricultural Research, Directorate of Medicinal and Aromatic Plants Research, Boriavi, Anand Gujarat, India

<sup>2</sup> Indian Council of Agricultural Research, Directorate of Groundnut Research ICAR-DGR, PB 5, Junagadh, Gujarat 362 001, India

fluorescence is a very sensitive probe of physiological status of leaves and plant performance in a wide range of situations (Daniele et al. 2006). Recently, mini-core germplasm of peanuts has been evaluated and grouped for variability in various physiological traits including  $P_N$ ,  $g_s$ ,  $E$  and  $F_v/F_m$  in field during dry season (Singh et al. 2014).

For plant improvement, information on photosynthetic performance cannot just be obtained by measurements of gaseous exchange (Dwyer et al. 1992). Therefore, evaluation of physiological state based on the time dependent changes of the chlorophyll  $a$  fluorescence along with photosynthetic activities is required to be studied under stress environments. Such studies in peanuts are in limited numbers with special focus on heat and light stress. Further, there are no reports on association of de-epoxidation state with the surrogates of WUE (SCMR and SLA) and  $WUE_{int}$  in peanuts under water deficit stress. Thus, a study was aimed to unravel the changes in chlorophyll fluorescence parameters, net photosynthetic rate and relative de-epoxidation state and its association with  $WUE_{int}$  at 75–80 DAS under limited water supply in 12 peanut genotypes during dry season in open field condition.

## Materials and methods

A field experiment taking 12 genotypes of peanut in split plot design giving first two flood irrigations of 60 mm followed by three irrigation treatments *viz.*  $T_1$ : well watered in which the amount of irrigation water supplied replenish the cumulative PAN evaporation,  $T_2$ - 50 % of  $T_1$  and  $T_3$ -25 % of  $T_1$  through drip during dry season (January- June) in year 2013 was conducted at the research farm of ICAR-Directorate of Groundnut Research, Junagadh, Gujarat (lat.  $21^{\circ} 31'N$ , Long  $70^{\circ} 36'E$ ) in the Vertic Ustochrept soil with pH of 8.5 and electrical conductivity of  $0.16 \text{ dSm}^{-1}$ . A total of 12 rows of peanut were sown in plot of  $5 \times 4 \text{ m}$  size in three replications with four rows of each genotype in each replication. Before sowing, fertilisers (40 N: 50 P: 50 K) were applied in furrows. A set of 12 peanut genotypes, including three from Spanish bunch (AK 265, GG 20, Gimar 2) and nine from Virginia bunch (Gimar 3, ICGV 86590, ICGV 91114, JL 24, SB XI, SG 99, TAG 24, TMV 2 and TPG 41) were sown during last week of January in year 2013 maintaining a population density of  $22 \text{ plants m}^{-2}$ . Recommended agronomic and plant protection measures were followed, except for the irrigation treatments. The total quantity of irrigation water calculated from total hours of irrigation multiplied by the discharge rate of the drippers in addition to first two flood irrigations of 60 mm each showed that at 80 DAS, treatments  $T_1$ ,  $T_2$  and  $T_3$  received 498, 309 and  $215 \text{ Lm}^{-2}$ , respectively (Table 1). Thus, at 80 DAS, the actual quantity of water given in  $T_1$  was 20 % higher than the evaporative demand (ET), in  $T_2$  it was 25 % less than ET and in  $T_3$  the supplied irrigation quantity was 48 % less than

ET against the cumulative evaporative demand of 412 mm. The observations on RWC were recorded during 75–80 DAS.

To determine soil moisture content (SMC) by gravimetric method, soil samples were drawn from the upper layer (0–15 cm) and lower layer (15–30 cm) soil depths. The RWC was measured by the formula  $RWC (\%) = [(FW - DW) / (TW - DW)] * 100$ , where, FW fresh weight, DW dry weight and TW is turgid weight.

Maximum efficiency of PSII ( $F_v/F_m$ ) of the dark adapted leaves were recorded after 30 min dark adaptation by leaf clips and actual quantum yield of PSII ( $F_v'/F_m'$ ) and net photosynthetic rate ( $P_N$ ), stomatal conductance ( $g_s$ ) and transpiration rate ( $E$ ) were recorded between 08:00 and 10:00 h by LI-COR 6400, portable photosynthesis system (LI-COR Inc. Lincoln, Nebraska, USA) with modulated fluorescence measurement as described by Maxwell and Johnson (2000). The third leaf from the main axis was kept in the chamber by ensuring the thermocouple touching it from the underside. Temperature was set at ambient and giving a stable  $T_{leaf}$  reading. The artificial light was given at 1650 PAR including 10 % of blue light. The ambient  $CO_2$  was supplied to the chamber at the flow rate of  $300 \text{ m mol s}^{-1}$  and reading was considered when all 3 factors *viz.* flow,  $CO_2$  and  $H_2O$  were stable.

To investigate the relative de-epoxidation state, the xanthophyll cycle pigments were extracted from leaf samples by crushing with liquid nitrogen in chilled mortar and pestle. The sample was homogenised in 1 mL of acetone and collected in a 2 mL Eppendorf tube. One milliliter of acetone was used to clean the mortar and pool with the extract, the volume in the Eppendorf was adjusted to 2 mL. The extracts in the tubes were centrifuge for 5 min at a speed of 5000 rpm and the supernatant was collected. The extract was filtered through a  $0.22 \mu\text{m}$  PTFE filter. The first drop that passed through the filter was discarded to avoid contamination. The HPLC vial was filled and closed with a cap and these extracts were injected in the HPLC. The separation of the xanthophyll pools was carried out as per the modified method of García-Plazaola and Becerril (1999) used for Shimadzu, HPLC system. Detection was performed with a PAD 996 detector in the range 250–700 nm.

Chromatographic conditions included HPLC solvents with the mobile phase consisting of two components: solvent A; acetonitrile: methanol: Tris buffer (0.1 M; pH 8) (84:2:14) and solvent B; methanol: ethyl acetate (68:32) (Polle et al. 2001). The injection volume was  $15 \mu\text{l}$  and the solvent flow rate was  $0.8 \text{ ml min}^{-1}$  with working pressures below 1000 psi. After pigments separation, identification and quantification was done on the basis of the absorption spectra and the standards with known concentration and peaks were detected and integrated at 445 nm for xanthophylls content. The relative de-epoxidation state was calculated as  $(A+Z)/(V+A+Z) (\%)$ .

All the data recorded were the mean values of at least three independent observations with three repetitions in each replication. The data was subjected to analysis of variance

**Table 1** Temperature (°C), relative humidity (RH %), solar radiation (SR W/m<sup>2</sup>), evaporative demand (ET mm), quantity of irrigation water applied (L m<sup>2</sup>) and soil moisture content (%) at different soil depth during peanut crop growth period

|                        | Weather parameters |         |         |                     |        | Irrigation quantity applied |                     |                     | Soil moisture content (%) |          |                |          |                |          |
|------------------------|--------------------|---------|---------|---------------------|--------|-----------------------------|---------------------|---------------------|---------------------------|----------|----------------|----------|----------------|----------|
|                        | Temperature        |         | RH (%)  | SR                  | ET     | T <sub>1</sub>              | T <sub>2</sub>      | T <sub>3</sub>      | T <sub>1</sub>            |          | T <sub>2</sub> |          | T <sub>3</sub> |          |
|                        | T. Max.            | T. Min. | T. Mean | (W/m <sup>2</sup> ) | (mm)   | (L m <sup>2</sup> )         | (L m <sup>2</sup> ) | (L m <sup>2</sup> ) | 0–15 cm                   | 15–30 cm | 0–15 cm        | 15–30 cm | 0–15 cm        | 15–30 cm |
| 0–10 DAS               | 30.9               | 15.2    | 23.1    | 45.3                | 2346   | 35.9                        | 120 <sup>a</sup>    | 120 <sup>a</sup>    | 17.3                      | 18       | 12.7           | 15.6     | 9.1            | 10.3     |
| 11–20 DAS              | 34.2               | 15.5    | 24.9    | 39.3                | 2489   | 35.1                        | 36                  | 18                  | 18.7                      | 19.9     | 18.5           | 19.3     | 18.9           | 20.1     |
| 21–40 DAS              | 36.8               | 16.3    | 26.6    | 27.9                | 5649   | 88.8                        | 90                  | 45                  | 16.5                      | 19.3     | 12.3           | 15.7     | 8.7            | 11.2     |
| 41–80 DAS              | 37.3               | 21.8    | 29.6    | 47.2                | 12,285 | 253.3                       | 252                 | 126                 | 18.1                      | 18.6     | 13.2           | 14.4     | 7.2            | 9.8      |
| Mean/total till 80 DAS | 34.8               | 17.2    | 26.1    | 39.9                | 22,769 | 413.1                       | 498                 | 309                 | 17.7                      | 19.0     | 14.2           | 16.3     | 11.0           | 12.9     |

<sup>a</sup> indicates flood irrigation

appropriate to the experimental design using DSAASTAT (Onofri 2007) and the least significant differences were calculated to assess the significance of treatment means where the “F” test was found significant at 5 %. Principal component analysis was carried out PAST (ver. 2.17c) statistical software

## Results and discussion

### Soil moisture status

The soil moisture content (SMC) of the control field at 0–15 cm and 15–30 cm soil depth remained 17.7 % and 21.0 % respectively till 80 days. The corresponding SMC in 0–15 cm soil layer was 14.2 and 11.0 in T<sub>2</sub> and T<sub>3</sub> respectively, whereas in 15–30 cm soil layer it was 16.3 in T<sub>2</sub> and 12.9 in T<sub>3</sub> (Table 1).

### Relative Water Content (RWC), Specific leaf area (SLA), soil plant analytical development chlorophyll meter reading (SCMR)

The RWC of peanut leaves decreased with decreasing irrigation quantity. It was 93.5 % in T<sub>1</sub>, 91.1 % in T<sub>2</sub> but had drastically decreased to 77.2 % in T<sub>3</sub>. Genotypes also differed significantly with the highest RWC in Girnar 2 (92.5 %) and the lowest in Girnar 3 (84 %). The interaction was significant with the highest RWC in AK 265 (96.7 %) in T<sub>1</sub> and the lowest in SG 99 (72 %) in T<sub>3</sub>. The SLA had decreased whereas the SCMR had increased due limiting irrigation quantity (Table 2). The mean SLA was 184, 171 and 162 cm<sup>2</sup> g<sup>-1</sup> dw in T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> respectively. Genotype SG 99 had the lowest SLA and the highest SCMR whereas the highest SLA was in SB XI (Table 2).

### Total chlorophyll content, Net photosynthetic rate (P<sub>N</sub>) and stomatal conductance (g<sub>s</sub>)

The total chlorophyll content increased due to water deficit treatment. It was 6.84 mg g<sup>-1</sup>dw in T<sub>1</sub>, 7.21 mg g<sup>-1</sup>dw in T<sub>2</sub> and 8.35 mg g<sup>-1</sup>dw in T<sub>3</sub>. Among the genotypes, GG 20 topped for the highest total chlorophyll whereas it was lowest in SG 99 (Table 3). The photosynthetic rate was 28.6 μmol m<sup>-2</sup> s<sup>-1</sup> in T<sub>1</sub> and 29.6 μmol m<sup>-2</sup> s<sup>-1</sup> in T<sub>2</sub> which decreased to 24.3 μmol m<sup>-2</sup> s<sup>-1</sup> in T<sub>3</sub> during 75 to 80 days after sowing (DAS). The peanut genotype ICGV 91114 showed the highest P<sub>N</sub> (30.9 μmol m<sup>-2</sup> s<sup>-1</sup>) which was statistically at par with GG 20, ICGV 86590, TAG 24, SB XI, TMV 2 and TPG 41. The interaction was significant with the highest P<sub>N</sub> in ICGV 91114 (34.5 μmol m<sup>-2</sup> s<sup>-1</sup>) in T<sub>1</sub>. The significant decrease in stomatal conductance was due to irrigation treatments (Table 3). The g<sub>s</sub> was higher in few genotypes in T<sub>1</sub> and in some of the genotypes in T<sub>2</sub>. The g<sub>s</sub> decreased in all the

**Table 2** Relative water content (RWC), specific leaf Area (SLA) and SPAD chlorophyll meter reading (SCMR) in peanut genotypes grown at 75–80 DAS, grown at various soil moisture regimes

| Genotype/Treatment  | RWC (%)        |                |                |       | SLA (cm <sup>2</sup> g <sup>-1</sup> dw) |                |                |         | SCMR           |                |                |          |
|---------------------|----------------|----------------|----------------|-------|--|----------------|----------------|---------|----------------|----------------|----------------|----------|
|                     | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean  | T <sub>1</sub>                           | T <sub>2</sub> | T <sub>3</sub> | Mean    | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean     |
| AK 265              | 96.7a          | 92.9a          | 79.5abc        | 89.7a | 180cd                                    | 175ab          | 188a           | 181ab   | 29.5d          | 38.5b          | 41.1bc         | 36.4bcd  |
| GG 20               | 93.9a          | 90.9a          | 72.2d          | 85.7a | 180cd                                    | 175ab          | 162cde         | 172abcd | 37.9a          | 38.2b          | 42.6           | 39.6ab   |
| Girnar 3            | 90.4a          | 88.1a          | 73.6cd         | 84a   | 200ab                                    | 175ab          | 158cde         | 178ab   | 31.9bcd        | 32.8de         | 43b            | 35.9bcde |
| Girnar 2            | 95.1a          | 92.5a          | 82.9a          | 90.1a | 178cde                                   | 169ab          | 135f           | 161cd   | 30.6cd         | 38.7b          | 41.4bc         | 36.9bc   |
| ICGV 86590          | 91.2a          | 91.2a          | 82.2ab         | 88.2a | 171de                                    | 160bc          | 151def         | 161cd   | 31bcd          | 37bc           | 44.2ab         | 37.4bc   |
| ICGV 9114           | 92.6a          | 90.6a          | 78.9abcd       | 87.4a | 203a                                     | 172ab          | 147ef          | 174abc  | 29.5d          | 33.4cde        | 34.1ef         | 32.3ef   |
| JL 24               | 93.3a          | 89.1a          | 76.9abcd       | 86.4a | 185bcd                                   | 181a           | 166bcd         | 177abc  | 34abc          | 31.7e          | 31.4ef         | 32.4def  |
| SB XI               | 95.7a          | 93a            | 74.2cd         | 87.6a | 200ab                                    | 184a           | 179ab          | 188a    | 35ab           | 36bcd          | 35.5e          | 35.5cdef |
| SG 99               | 93.2a          | 90.6a          | 72.0d          | 85.3a | 163e                                     | 162bc          | 147ef          | 157d    | 36.7a          | 45a            | 47.1a          | 42.9a    |
| TAG 24              | 94.0a          | 90.1a          | 75.6bcd        | 86.6a | 183cd                                    | 181a           | 160cde         | 175abc  | 31.4bcd        | 30.9e          | 32.4ef         | 31.6f    |
| TMV 2               | 93.1a          | 91.3a          | 75.5bcd        | 86.6a | 182cd                                    | 152c           | 174abc         | 169bcd  | 28.6d          | 32.9de         | 34.6ef         | 32ef     |
| TPG 41              | 92.4a          | 93.1a          | 83.2a          | 89.6a | 188abc                                   | 163bc          | 179ab          | 177abc  | 36.5a          | 37.6b          | 38.9cd         | 37.7bc   |
| Mean                | 93.5           | 91.1           | 77.2           | 87.3  | 184                                      | 171            | 162            | 172     | 32.7           | 36.1           | 38.9           | 35.9     |
| LSD <sub>0.05</sub> |                |                |                |       |  |                |                |         |                |                |                |          |
| Treatments          | 7.4            |                |                |       | 1.9                                      |                |                |         | 0.9            |                |                |          |
| Genotypes           | 3.3            |                |                |       | 9.6                                      |                |                |         | 2.36           |                |                |          |
| Interaction T × G   | 5.6            |                |                |       | 16.7                                     |                |                |         | 4.09           |                |                |          |

genotypes except SB XI in T<sub>3</sub>. There was a 15 % and 39 % reduction in stomatal conductance is reported in T<sub>2</sub> and T<sub>3</sub> respectively as compared to the T<sub>1</sub>. The  $g_s$  in T<sub>1</sub> was 0.456 m sec<sup>-1</sup> which decreased to 0.383 msec<sup>-1</sup> in T<sub>2</sub> and further to 0.278 msec<sup>-1</sup> in T<sub>3</sub> at 60 DAS. Genotypes differed significantly for  $g_s$  with the highest in GG20 (0.465 msec<sup>-1</sup>) and the lowest in Girnar 2 (0.286 msec<sup>-1</sup>).

#### Intra cellular CO<sub>2</sub> (C<sub>i</sub>), transpiration rate (E) and intrinsic water use efficiency (WUE<sub>int</sub>)

The interaction being significant, the highest  $g_s$  is reported in GG 20 (0.607 msec<sup>-1</sup>) under T<sub>1</sub>. The intra cellular CO<sub>2</sub> concentration (C<sub>i</sub>) increased due to irrigation treatment. It was highest at the severe water deficit condition. As per the reported genotypic difference lowest C<sub>i</sub> was in Girnar 2 (Table 4). The irrigation treatment affected the transpiration rate (E) which reduced by 15 % and 43 % in T<sub>2</sub> and T<sub>3</sub> over T<sub>1</sub>. The E was highest in genotype GG 20 (10.5 mmol m<sup>-2</sup> s<sup>-1</sup>) followed by genotype AK 265 (10.4 mmol m<sup>-2</sup> s<sup>-1</sup>). The interaction was significant with the highest E in TMV 2 (14.6 mmol m<sup>-2</sup> s<sup>-1</sup>) under T<sub>1</sub> and the lowest in Girnar 2 (5.4 mmol m<sup>-2</sup> s<sup>-1</sup>) under T<sub>3</sub>. The intrinsic water use efficiency (WUE<sub>int</sub>) had increased due to limited water supply with the highest in T<sub>3</sub>. It ranged between 2.6 and 3.5 in among the genotypes and the highest WUE<sub>int</sub> in T<sub>3</sub> was in genotype SG 99 (Table 4).

#### Maximum quantum yield of PSII ( $F_v/F_m$ ), actual efficiency of photosynthesis ( $F_v'/F_m'$ ) and quantum yield of PS II ( $\Phi_{PSII}$ )

The maximum quantum yield of PSII ( $F_v/F_m$ ) reduced to 0.831 in T<sub>3</sub> from 0.839 in T<sub>1</sub>. Among genotypes, highest  $F_v/F_m$  was in TMV 2 (0.844) and the lowest in TAG 24 (0.822). The interaction for  $F_v/F_m$  was significant with the highest value in TMV 2 (0.847) in T<sub>1</sub> and the lowest in TAG 24 (0.799) in T<sub>3</sub>. Interestingly, the actual efficiency of photosynthesis ( $F_v'/F_m'$ ) was significantly highest in T<sub>2</sub>. Among the genotypes, the highest  $F_v'/F_m'$  was in ICGV 86590, which was statistically at par with JL 24, SG 99 and TAG 24. The quantum yield of PSII or the proportion of absorbed energy utilised for photochemistry ( $\Phi_{PSII}$ ) was not significantly affected due to irrigation quantity. Genotypic difference were observed for proportion of absorbed energy utilised for photochemistry and the highest  $\Phi_{PSII}$  was in GG 20 (0.283) and the lowest in JL 24 (0.191). The interaction was significant for  $\Phi_{PSII}$  with the highest in GG 20 (0.320) in T<sub>1</sub> and the lowest in JL 24 (0.181) in T<sub>3</sub> (Table 5).

#### Non-photochemical quenching (NPQ) and relative de-epoxidation state (DeS)

The NPQ increased due to water deficit stress and was lowest in T<sub>2</sub> and the highest in T<sub>3</sub> (Table 6). Among the genotypes, the capacity to disseminate the extra absorbed energy in terms

**Table 3** Total chlorophyll content ( $\text{mg g}^{-1}$  dw), net photosynthetic rate ( $P_N$  - $\mu\text{ mol m}^{-2} \text{s}^{-1}$ ) and stomatal conductance ( $g_s$   $\text{m sec}^{-1}$ ) in peanut genotypes at 75–80 DAS grown at various soil moisture regimes

| Genotype/treatment  | Total chlorophyll content ( $\text{mg g}^{-1}$ dw) |                |                |          | $P_N$ ( $\mu\text{ mol m}^{-2} \text{s}^{-1}$ ) |                |                |         | $g_s$ ( $\text{m sec}^{-1}$ ) |                |                |           |
|---------------------|--|----------------|----------------|----------|---|----------------|----------------|---------|-------------------------------|----------------|----------------|-----------|
|                     | T <sub>1</sub>                                     | T <sub>2</sub> | T <sub>3</sub> | Mean     | T <sub>1</sub>                                  | T <sub>2</sub> | T <sub>3</sub> | Mean    | T <sub>1</sub>                | T <sub>2</sub> | T <sub>3</sub> | Mean      |
| AK 265              | 5.68g  | 6.31e          | 9.37ab         | 7.12def  | 26.1bcd   | 27.9bc         | 26.1ab         | 26.7bc  | 0.478bcd                      | 0.335cde       | 0.286abc       | 0.366abcd |
| GG 20               | 6.48ef   | 9.25c          | 8.91b          | 8.21a    | 31.2abc   | 32.6a          | 23.5bc         | 29.1ab  | 0.607a                        | 0.292de        | 0.276abc       | 0.465a    |
| Gimar 3             | 7.79ab   | 6.52e          | 7.28de         | 7.19cdef | 29abcd  | 25.9c          | 23bcd          | 26bc    | 0.412cd                       | 0.234e         | 0.253bc        | 0.319bcd  |
| Gimar 2             | 8.03a  | 5.1f           | 9.21ab         | 7.45bcde | 31.4ab  | 26.1c          | 19.4d          | 25.6bc  | 0.437cd                       | 0.336cde       | 0.188c         | 0.286d    |
| ICGV 86590          | 7.26bcd  | 5.92e          | 9.69a          | 7.63abcd | 29.6abcd  | 30.4ab         | 25.5abc        | 28.5abc | 0.515abc                      | 0.449abc       | 0.306abc       | 0.386abcd |
| ICGV 9114           | 6.5ef  | 7.94d          | 8.99b          | 7.81abc  | 34.5a   | 34.1a          | 24.1bc         | 30.9a   | 0.574ab                       | 0.338cde       | 0.266bc        | 0.43ab    |
| JL 24               | 4.93h  | 10.23b         | 9.42ab         | 8.19a    | 24.3cd  | 27.3bc         | 22.0cd         | 24.5c   | 0.435cd                       | 0.341cde       | 0.275abc       | 0.349abcd |
| SB XI               | 6.2fg  | 11.56a         | 6.84e          | 8.2a     | 28.8abcd  | 26.0c          | 29.0a          | 27.9abc | 0.471bcd                      | 0.437abc       | 0.39a          | 0.4abcd   |
| SG 99               | 7.59abc  | 4.32g          | 7.86cd         | 6.59fg   | 23.2d   | 32.4a          | 23.4bcd        | 26.3bc  | 0.244e                        | 0.369bcd       | 0.238bc        | 0.306cd   |
| TAG 24              | 6.8def   | 6.07e          | 8.07c          | 6.98efg  | 30.2abcd  | 31.3ab         | 25abc          | 28.8ab  | 0.432cd                       | 0.479ab        | 0.286abc       | 0.362abcd |
| TMV 2               | 7.9ab  | 8.8c           | 6.86e          | 7.85ab   | 29abcd  | 30.3ab         | 22.8bcd        | 27.3abc | 0.486bcd                      | 0.474ab        | 0.259bc        | 0.408abc  |
| TPG 41              | 6.97cde  | 4.47fg         | 7.65cd         | 6.36g    | 26.5bcd   | 30.7ab         | 28.2a          | 28.5abc | 0.383d                        | 0.513a         | 0.319ab        | 0.392abcd |
| Mean                | 6.84   | 7.21           | 8.35           | 7.47     | 28.6  | 29.6           | 24.3           | 27.5    | 0.456                         | 0.383          | 0.278          | 0.373     |
| LSD <sub>0.05</sub> |  |                |                |          |   |                |                |         |                               |                |                |           |
| Treatments          | 0.97   |                |                |          | 1   |                |                |         | 0.07                          |                |                |           |
| Genotypes           | 0.18   |                |                |          | 2.4   |                |                |         | 0.07                          |                |                |           |
| Interaction TxG     | 0.3  |                |                |          | 4.2   |                |                |         | 0.12                          |                |                |           |

**Table 4** Intra cellular CO<sub>2</sub> (C<sub>i</sub>) transpiration rate (E mmol m<sup>-2</sup> s<sup>-1</sup>) and intrinsic water use efficiency (WUE<sub>int</sub>) in peanut genotypes at 75–80 DAS, grown at various soil moisture regimes

| Genotype/treatment  | C <sub>i</sub> |                |                |       | E (mmol m <sup>-2</sup> s <sup>-1</sup> ) |                |                |       | WUE <sub>int</sub> |                |                |        |
|---------------------|----------------|----------------|----------------|-------|---|----------------|----------------|-------|--------------------|----------------|----------------|--------|
|                     | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean  | T <sub>1</sub>                            | T <sub>2</sub> | T <sub>3</sub> | Mean  | T <sub>1</sub>     | T <sub>2</sub> | T <sub>3</sub> | Mean   |
| AK 265              | 220ab          | 209abc         | 247ab          | 225ab | 13.7ab                                    | 10.1bc         | 7.4abc         | 10.4a | 1.9c               | 2.8b           | 3.6bc          | 2.8bc  |
| GG 20               | 216ab          | 225abc         | 215bc          | 219ab | 13.1ab                                    | 11.9a          | 6.5bcde        | 10.5a | 2.4abc             | 2.7b           | 3.6bc          | 2.9abc |
| Girnar 3            | 197ab          | 203bc          | 234abc         | 211ab | 9.7f                                      | 9.9bc          | 5.6e           | 8.4b  | 3.0a               | 2.7b           | 4.1ab          | 3.3ab  |
| Girnar 2            | 152c           | 204bc          | 210bc          | 189b  | 13.2ab                                    | 9.6c           | 5.4e           | 9.4ab | 2.4abc             | 2.8b           | 3.6bc          | 2.9abc |
| ICGV 86590          | 220ab          | 220abc         | 257a           | 232a  | 12.7bc                                    | 9.5c           | 8.3a           | 10.2a | 2.4abc             | 3.2ab          | 3.1cd          | 2.9abc |
| ICGV 9114           | 227a           | 198c           | 213bc          | 213ab | 13.5ab                                    | 9.1c           | 6cde           | 9.6ab | 2.6abc             | 3.7a           | 4.0b           | 3.4ab  |
| JL 24               | 180bc          | 210abc         | 258a           | 216ab | 10.0ef                                    | 10.4bc         | 8.4a           | 9.6ab | 2.5abc             | 2.7b           | 2.6d           | 2.6c   |
| SB XI               | 215ab          | 228abc         | 244ab          | 229ab | 9.7f                                      | 9.5c           | 6.3bcde        | 8.5b  | 3a                 | 2.7b           | 4.7a           | 3.5a   |
| SG 99               | 205ab          | 219abc         | 195c           | 206ab | 11.2def                                   | 10.5bc         | 7.5ab          | 9.7ab | 2.1bc              | 3.1ab          | 3.1cd          | 2.8bc  |
| TAG 24              | 187abc         | 228abc         | 249ab          | 221ab | 11.3cde                                   | 10.4bc         | 8.4a           | 10a   | 2.7ab              | 3.0b           | 3.0cd          | 2.9abc |
| TMV 2               | 221ab          | 240ab          | 238ab          | 233a  | 14.6a                                     | 10.5abc        | 5.9de          | 10.3a | 2c                 | 2.9b           | 4ab            | 3.0abc |
| TPG 41              | 222a           | 246a           | 259a           | 242a  | 11.6cd                                    | 11.2ab         | 7.1abce        | 10a   | 2.3bc              | 2.7b           | 4.2ab          | 3.1abc |
| Mean                | 205            | 219            | 235            | 220   | 12  | 10.2           | 6.9            | 9.7   | 2.4                | 2.9            | 3.6            | 3.0    |
| LSD <sub>0.05</sub> |                |                |                |       |   |                |                |       |                    |                |                |        |
| Treatments          | 11.9           |                |                |       | 0.58                                      |                |                |       | 0.24               |                |                |        |
| Genotypes           | 23.4           |                |                |       | 0.84                                      |                |                |       | 0.39               |                |                |        |
| Interaction T × G   | NS             |                |                |       | 1.46                                      |                |                |       | 0.68               |                |                |        |

of *NPQ* was highest in Girnar 3 (1.88) and the lowest in JL 24 (1.10). The *DeS* was 38 % in T<sub>1</sub> and T<sub>2</sub> but, increased to 47 % in T<sub>3</sub>. The highest de-epoxidation state was found in ICGV86590 (48.7 %) followed by SG 99 (47.2 %) whereas the lowest de-epoxidation state was found in TMV 2 (33.8 %). Genotypes performed differently for the state of de-epoxidation under different irrigation treatment. The per cent de-epoxidation was higher in genotypes GG 20, ICGV 86590, JL 24, SG 99, Girnar 3, ICGV 91114 and TMV 2 in T<sub>2</sub> as compared to the T<sub>1</sub>. But, under T<sub>3</sub> the per cent de-epoxidation had increased in all the genotypes.

#### Genotype by trait biplots and trait relationship analysis

The genotype-by-trait (GT-biplot) for each of the three treatments explained more than 85 % variation. The GT-biplot of first five PCs with eigen values more than one explained 86.8 % variation observed in T<sub>1</sub> (Fig. 1a). Among the components of the PCA of T<sub>1</sub> (Fig. 2a, b and c) the highest contribution was governed by *g<sub>s</sub>* very closely followed by *NPQ* and *F<sub>v</sub>/F<sub>m</sub>* in PC 1. The PC 2 was governed by *P<sub>N</sub>* and *DeS* whereas the PC 3 was governed by *C<sub>i</sub>* and *E*. Overall the vectors of WUE<sub>int</sub>, *E* and *SLA* explained maximum variation among genotypes. A fair contribution to the variation also comes from *DeS* and *NPQ* whereas the contribution of *RWC*, *Φ<sub>PS II</sub>*, and total chlorophyll content were relatively low in T<sub>1</sub>. Genotype SG 99 out performed for de-epoxidation state, AK 265 for *E*, ICGV 91114 for *P<sub>N</sub>* and

Girnar 3 for high WUE<sub>int</sub>. An acute angle observed between *P<sub>N</sub>*, *g<sub>s</sub>* and *Φ<sub>PS II</sub>* indicated positive associations among these traits. Genotypes ICGV 91114 and GG 20 were best for this group of traits. This further explains that increasing *g<sub>s</sub>* can improve the *P<sub>N</sub>* under well watered condition.

PCA of T<sub>2</sub> explains that the variation was derived from five PCs (Fig. 2d, e and f) reaching 87.7 % of total variation observed. PC 1 was dominated by WUE<sub>int</sub>, and *SLA*, PC 2 was dominated by *Φ<sub>PS II</sub>* and PC 3 was again dominated by WUE<sub>int</sub>. Over all, among the components of the PCA (Fig. 1b), the highest contribution was governed by *g<sub>s</sub>*, very closely followed by *NPQ* and *F<sub>v</sub>/F<sub>m</sub>* in PC 1. Higher proportion to the variation due to *Φ<sub>PS II</sub>*, *C<sub>i</sub>*, *NPQ*, WUE<sub>int</sub> and *DeS* is observed in GT-biplot of T<sub>2</sub>. This was the condition when WUE<sub>int</sub> was closely and positively associated with *DeS* and *Φ<sub>PS II</sub>* whereas *SCMR*, *F<sub>v</sub>/F<sub>m</sub>* and *DeS* were very closely associated with the *P<sub>N</sub>*. GG 20 was the best for higher stomatal conductance and low *NPQ* whereas Girnar 3 tops for high *NPQ* and low *g<sub>s</sub>*. The first PCA component in GT-Biplot of the T<sub>2</sub> explains the contribution by the *P<sub>N</sub>* very closely followed by the *DeS* (Fig. 2b).

Similarly as T<sub>1</sub> and T<sub>2</sub>, the variation in T<sub>3</sub> also from major first five PCs contributing to a total of 87.7 % variation (Fig. 2g, h and i). PC 1 of the loadings in the T<sub>3</sub> showed that the highest contribution to the variation was governed by the chlorophyll content whereas in PC 2 the variation was mainly contributed by *F<sub>v</sub>/F<sub>m</sub>* and total chlorophyll content whereas in PC 3, the major variation was governed by decreased *F<sub>v</sub>/F<sub>m</sub>*

**Table 5** Maximum efficiency of photosynthesis ( $F_v/F_m$ ), actual efficiency of photosynthesis ( $F_v'/F_m'$ ) and quantum yield of PS II ( $\Phi_{PSII}$ ) grown at various soil moisture regimes

| Genotype/treatment  | $F_v/F_m$      |                |                | $F_v'/F_m'$ |                |                | $\Phi_{PSII}$  |          |                | Mean      |                |                |
|---------------------|----------------|----------------|----------------|-------------|----------------|----------------|----------------|----------|----------------|-----------|----------------|----------------|
|                     | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean        | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean     | T <sub>1</sub> |           | T <sub>2</sub> | T <sub>3</sub> |
| AK 265              | 0.845a         | 0.831abc       | 0.825c         | 0.834ab     | 0.571abcd      | 0.615abcde     | 0.565abcd      | 0.584abc | 0.219ef        | 0.242abcd | 0.235ab        | 0.232bc        |
| GG 20               | 0.837ab        | 0.839ab        | 0.825c         | 0.833ab     | 0.538d         | 0.621abcd      | 0.513cd        | 0.557abc | 0.361a         | 0.259ab   | 0.23ab         | 0.283a         |
| Gimar 3             | 0.84a          | 0.845a         | 0.837abc       | 0.841a      | 0.54d          | 0.592bcde      | 0.572abcd      | 0.568abc | 0.32ab         | 0.281a    | 0.214abc       | 0.272ab        |
| Gimar 2             | 0.842a         | 0.823c         | 0.838abc       | 0.834ab     | 0.545d         | 0.562de        | 0.549bcd       | 0.552bc  | 0.297bc        | 0.235abcd | 0.209bc        | 0.247ab        |
| ICGV 86590          | 0.843a         | 0.838ab        | 0.841ab        | 0.841a      | 0.636a         | 0.673a         | 0.579abc       | 0.629c   | 0.224ef        | 0.234bcd  | 0.228ab        | 0.229bc        |
| ICGV 9114           | 0.846a         | 0.836abc       | 0.826bc        | 0.836ab     | 0.562bcd       | 0.579cde       | 0.506d         | 0.549bc  | 0.287bcd       | 0.259ab   | 0.227abc       | 0.258ab        |
| JL 24               | 0.844a         | 0.834abc       | 0.831abc       | 0.836ab     | 0.585abcd      | 0.665a         | 0.634a         | 0.628a   | 0.193f         | 0.199d    | 0.181c         | 0.191c         |
| SB XI               | 0.838ab        | 0.829bc        | 0.841a         | 0.836ab     | 0.590abcd      | 0.555de        | 0.58abc        | 0.575abc | 0.242de        | 0.218bcd  | 0.245ab        | 0.235bc        |
| SG 99               | 0.824b         | 0.839ab        | 0.841a         | 0.835ab     | 0.619abc       | 0.666a         | 0.579abc       | 0.621a   | 0.218ef        | 0.246abc  | 0.218abc       | 0.227bc        |
| TAG 24              | 0.834ab        | 0.833abc       | 0.799d         | 0.822b      | 0.627ab        | 0.649ab        | 0.573abcd      | 0.616ab  | 0.225ef        | 0.234abcd | 0.258a         | 0.239ab        |
| TMV 2               | 0.847a         | 0.845a         | 0.838abc       | 0.844a      | 0.553cd        | 0.546c         | 0.547bcd       | 0.548c   | 0.25cde        | 0.218bcd  | 0.21bc         | 0.226bc        |
| TPG 41              | 0.824b         | 0.834abc       | 0.834abc       | 0.831ab     | 0.603abcd      | 0.647abc       | 0.586ab        | 0.612abc | 0.237ef        | 0.21cd    | 0.239ab        | 0.228bc        |
| Mean                | 0.839          | 0.836          | 0.831          | 0.835       | 0.581          | 0.614          | 0.565          | 0.587    | 0.256          | 0.236     | 0.224          | 0.239          |
| LSD <sub>0.05</sub> |                |                |                |             |                |                |                |          |                |           |                |                |
| Treatments          | 0.004          |                |                |             | 0.024          |                |                |          | NS             |           |                |                |
| Genotypes           | 0.009          |                |                |             | 0.041          |                |                |          | 0.026          |           |                |                |
| Interaction T × G   | 0.015          |                |                |             | NS             |                |                |          | 0.045          |           |                |                |

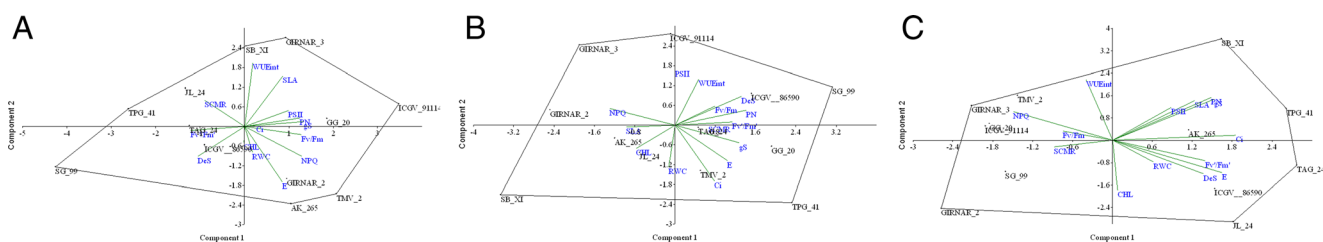
**Table 6** Non-photochemical quenching (*NPQ*) and relative depoxidation state (*DeS*) in peanut genotypes at 75–80 DAS, grown at various soil moisture regimes

| Genotype/treatment  | <i>NPQ</i>     |                |                |        | <i>DeS</i>     |                |                |        |
|---------------------|----------------|----------------|----------------|--------|----------------|----------------|----------------|--------|
|                     | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean   | T <sub>1</sub> | T <sub>2</sub> | T <sub>3</sub> | Mean   |
| AK 265              | 1.77a          | 1.3bcde        | 1.85ab         | 1.64ab | 34.8e          | 30.8f          | 48.0de         | 37.9d  |
| GG 20               | 1.61a          | 1.17bcde       | 2.19a          | 1.66ab | 35.1d          | 43.0bc         | 44.4f          | 40.8c  |
| Gimar 3             | 1.44a          | 2.06a          | 2.13a          | 1.88a  | 31.8g          | 37.7e          | 38.8g          | 36.1de |
| Gimar 2             | 1.67a          | 2.03a          | 1.58abc        | 1.76ab | 45.9a          | 40.4d          | 54.3b          | 46.8b  |
| ICGV 86590          | 1.14a          | 0.78e          | 1.78ab         | 1.23ab | 40.9c          | 44.4b          | 60.7a          | 48.7a  |
| ICGV 9114           | 1.72a          | 1.54abc        | 2.17a          | 1.81ab | 29.9h          | 38.1e          | 44.4f          | 37.5d  |
| JL 24               | 1.22a          | 0.82de         | 1.25bc         | 1.1b   | 33.1ef         | 38.1e          | 51.0c          | 40.8c  |
| SB XI               | 1.35a          | 1.79ab         | 1.73abc        | 1.63ab | 30.1h          | 25.2g          | 49.2cd         | 34.9ef |
| SG 99               | 1.30a          | 0.96cde        | 1.54abc        | 1.26ab | 45.2a          | 49.2a          | 47.2e          | 47.2ab |
| TAG 24              | 1.43a          | 0.9cde         | 1.06c          | 1.13b  | 45.9a          | 37.7e          | 54.5b          | 46.0b  |
| TMV 2               | 1.75a          | 1.53abcd       | 1.88ab         | 1.72ab | 32.3fg         | 32.6f          | 36.5h          | 33.8f  |
| TPG 41              | 1.11a          | 0.97cde        | 1.28bc         | 1.12b  | 43.2b          | 41.5cd         | 55.5b          | 46.8b  |
| Mean                | 1.46           | 1.32           | 1.7            | 1.5    | 37.4           | 38.2           | 48.7           | 41.4   |
| LSD <sub>0.05</sub> |                |                |                |        |                |                |                |        |
| Treatments          | 0.26           |                |                |        | 0.54           |                |                |        |
| Genotypes           | 0.41           |                |                |        | 1.07           |                |                |        |
| Interaction T × G   | NS             |                |                |        | 1.85           |                |                |        |

and  $\Phi_{PSII}$ . Overall, four most important traits that has contributed heavily are  $WUE_{int}$ ,  $P_N$ ,  $C_i$ ,  $E$  and total chlorophyll content in T<sub>3</sub> (Fig. 1c). Two prominent groups of traits are visualised from the GT-by plot of the T<sub>3</sub> (Fig. 1c). The first group is comprised of  $P_N$ ,  $SLA$ ,  $\Phi_{PSII}$  and  $g_s$  and the second one comprising of  $RWC$ ,  $DeS$  and  $F_v/F_m$ . Both these groups were associated with the  $C_i$  which in turn was plotted almost at right angle with the  $WUE_{int}$  indicating a weak relationship. From the GT-biplot, it is clear that genotypes behaved differently under different soil moisture regimes. Most of the traits contributed for the observed variation in T<sub>1</sub> and T<sub>2</sub> independently, but in T<sub>3</sub>, these traits were divided in group of traits making the relationship more complex.

## Discussion

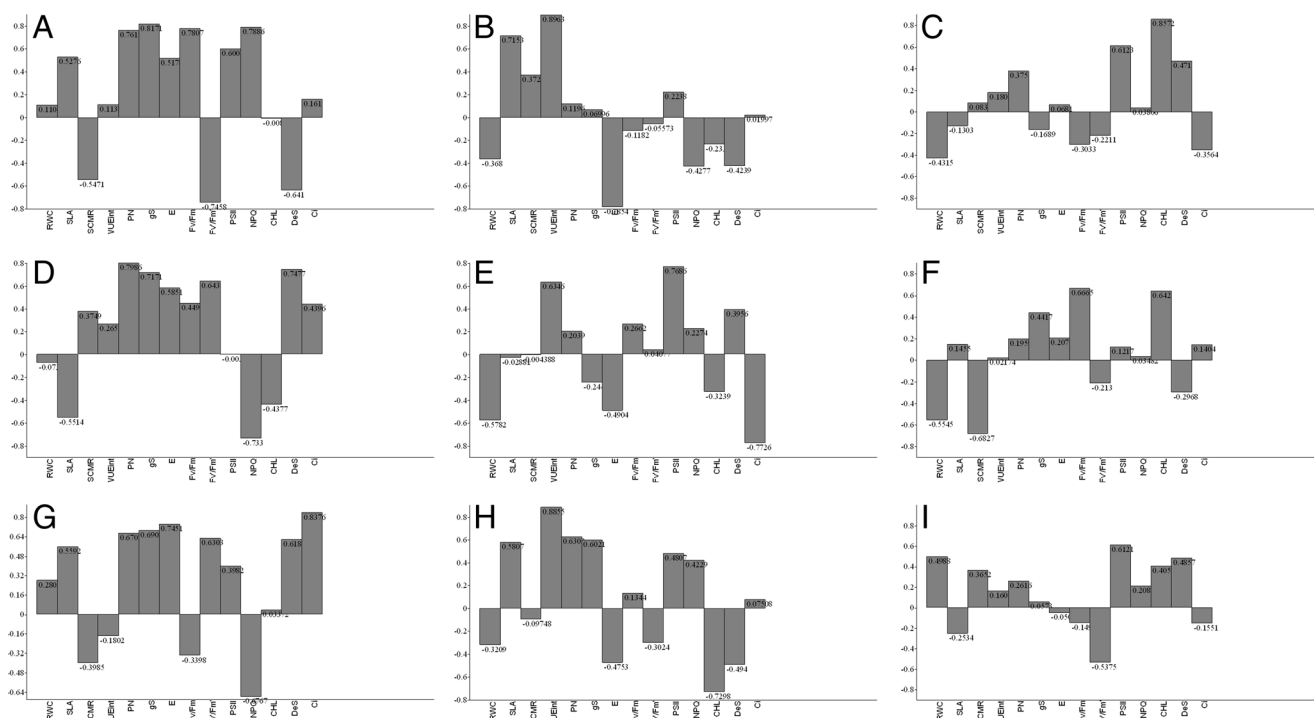
The  $RWC$ , stomatal resistance, rate of transpiration and canopy temperatures are important parameters that influence water relations in peanut (Nautiyal et al. 1995, 2012). The  $RWC$  of the leaf is more stable and sensitive than water potential in peanut limited soil moisture availability (Clavel et al. 2005). The significant reduction in  $RWC$  in this study was in close agreement with soil water availability under different irrigation treatments. Daniele et al. (2006) found that in peanut, the genotypic discrimination of  $RWC$  trait depends on the water regime and also genetic background which seems to be the reason behind genotypic difference in  $RWC$ .



**Fig. 1** Vector view of genotype × trait biplot summarising the interrelationship among the traits under various irrigation treatments (a) T<sub>1</sub> (b) T<sub>2</sub> and (c) T<sub>3</sub>.  $RWC$  relative water content,  $SLA$  specific leaf area,  $P_N$  net photosynthesis rate,  $E$  transpiration rate,  $WUE$  intrinsic  $WUE$ ,  $g_s$  stomatal conductance,  $F_v/F_m$  Maximum quantum yield of PSII  $F_v/F_m$  =

actual efficiency of photosynthesis,  $\Phi_{PSII}$  Energy utilized for photochemistry,  $NPQ$  Non-photochemical quenching,  $DeS$  relative depoxidation state,  $Chl$  total chlorophyll content,  $SCMR$  SPAD chlorophyll meter reading,  $C_i$  intracellular  $CO_2$  concentration





**Fig. 2** Principal component s of various treatments (a) PC 1 T<sub>1</sub>, (b) PC 2 T<sub>1</sub>, (c) PC 3 T<sub>1</sub>, (d) PC 1 T<sub>2</sub>, (e) PC 2 T<sub>2</sub>, (f) PC 3 T<sub>2</sub>, (g) PC 1 T<sub>3</sub>, (h) PC 2 T<sub>3</sub> and (i) PC 3 T<sub>3</sub>

Decrease in SLA or more appropriately the thickening of leaves usually have higher chlorophyll content per unit leaf area and hence had a greater photosynthetic capacity compared with thinner leaves. The photosynthetically active light-transmittance characteristics of the leaf is dependent on the unit amount of chlorophyll content per unit leaf area (chlorophyll density) (Richardson et al. 2002). The SCMR is the indirect measure of the chlorophyll density. The decrease in SLA and increase in SCMR and total chlorophyll content are closely associated in this study. Significant and positive correlations between SCMR and chlorophyll content (Akkasaeng et al. 2003) and chlorophyll density (Arunyanark et al. 2008) have been reported.

Reduction in canopy photosynthesis by imposing moisture stress is found mainly due to reduction in stomatal conductance and leaf area. As moisture stress increases, stomata start closing as a mechanism to reduce transpiration, as a consequence, the entry of carbon dioxide is also reduced. The  $P_N$  in peanut leaves decreases as relative water content (RWC) and water potential ( $\Psi$ ) decreases due to water deficit stress condition (Kalariya et al. 2013). The photosynthesis is fundamental in both biomass accumulation and productivity and it could be best utilized in identifying the efficient genotypes and to understand the physiological traits of productivity both under normal and stress conditions. Under increasing moisture deficit, the low SLA type peanut genotypes were able to maintain

higher RWC,  $P_N$  and  $g_s$  (Nautiyal et al. 2002). In general, the limitation of net photosynthetic rate in low moisture stressed plant is mainly through stomatal closure (Cornic and Massacci 1996) and/or by metabolic impairment (Flexas and Medrano 2002), however, the severity of drought decides the relative magnitude of stomatal and non-stomatal factors limiting photosynthesis. Plants transpire less under limited soil water status, as a result decreased rate of transpiration was observed in this study. Subramaniam and Maheswari (1990) reported that leaf water potential, transpiration rate and photosynthetic rate decreased progressively with increasing duration of water stress in peanut. The gas exchange variables at 60 DAS were affected by various frequency of irrigation and the best results for gaseous exchange characteristics were gained by highest irrigation frequency tested (every 2 days) in peanuts (Sousa et al. 2014). The transpiration efficiency (TE) is the  $WUE_{int}$  (the ratio of instantaneous  $CO_2$  assimilation (A) to transpiration at leaf level. A high photosynthetic efficiency consumes more  $CO_2$  and ultimately decreases  $C_i$  value. The increased value of  $C_i$  in T<sub>3</sub> seems to be because of in-efficiency of the photosynthetic incorporation of  $CO_2$ . A tight link has been found between large differences in TE in several crops and attributes of plants that make them restrict water losses under high vapour-pressure deficits (Vadez et al. 2014).

The GT-biplot shows the negative relationship between the E and  $WUE_{int}$  across the treatments and a very strong and

positive association between the  $P_N$  and the  $g_s$  under severe water deficit stress condition. The  $g_s$  was positively correlated with the  $C_i$  during  $T_2$  and  $T_3$ . Reduced  $g_s$  had resulted in restricted influx of  $CO_2$  and efflux of water however, a low  $P_N$  resulted in increased the  $C_i$ .

As per the reports of Baker and Horton (1987), decrease in  $F_v/F_m$  indicates a chronic photoinhibition due to photoinactivation of PSII centres. Reduced  $F_v/F_m$  is previously reported in durum wheat (Bogale et al. 2011) and Sapiñish peanut genotypes (Kalariya et al. 2013). Most of the plants adapt themselves to water stress by dissipating the excess excitation energy thermally with the down regulation of PSII activity to protect photosynthetic apparatus from photo-damaging effect under water deficit stress often coinciding with high leaf temperature (Bilger and Bjorkman 1990). The level non-radiative energy dissipation in the LHC II of PSII helps prevent the over reduction of the electron transfer chain and thereby provides protection against the photo-damage is indicated by  $NPQ$  (Krause and Weis 1991, Finazzi et al. 2006). This is achieved by the xanthophyll cycle through inter conversions of three carotenoid pigments: violaxanthin (V), antheraxanthin (A), and zeaxanthin (Z), which are catalysed by two enzymes: violaxanthin de-epoxidase (VDE: EC1.10.99.3) and zeaxanthineoxidase (ZE: EC1.14.13.90). The accumulation of Z and A, along with the trans-thylakoid pH gradient, mediates non radiative dissipation of light energy in the antennae (Björkman and Demmig-Adams 1994). This non radiative dissipation of light energy is an alternative energy path that diverts energy from PSII, effectively down-regulating PSII's efficiency which is dependent on the accumulation of de-epoxidation products (A+Z) of the xanthophyll cycle. Furthermore, Z may directly protect the thylakoid membrane against photo-oxidation as an antioxidant. Higher plants when exposed to photo-inhibition, the xanthophyll cycle dependent  $NPQ$  is the most useful mechanism to dissipate excess energy. Limited water supply has increased the  $NPQ$  in close accordance with the de-epoxidation state. Mishra et al. (2012) proposed that chlorophyll fluorescence emission responds to the level of water stress and thus, can be used to identify elevated drought tolerance in high-throughput screens for selection of resistant genotypes.

It is clear from the study that applying 20 % higher water than the ET ( $T_i$ ) does not warrant any extra benefit in terms of higher photosynthesis in peanut at 80 DAS. Further, a reduction of 25 % of the ET through drip (excluding first two flood irrigations) in peanut seems to be the ideal condition for photosynthesis and desirable chlorophyll fluorescence parameters at 75–80 DAS. But, a reduction of nearly 50 % of the evaporative demand through drip leads to a drastic reduction in photosynthesis, stomatal conductance and transpiration in peanut at 75–80 DAS.

As per the principal component analysis, a straight negative association of the DeS and the  $NPQ$  indicated that an efficient

de-epoxidation state is meant to photosynthetic machinery in peanuts. The  $WUE_{int}$ ,  $C_i$ ,  $P_N$ , and  $NPQ$  are major source of variation under water deficit stress condition in peanuts during dry season. Our results suggest that the genotypes Girnar 3 and ICGV 91114 which showed  $NPQ$  value above 2.2 and higher de-epoxidation state, maintained least deviation in  $F_v/F_m$  and  $F_v'/F_m'$  under severe water deficit condition are promising genotypes against water deficit stress induced photoinhibition.

**Acknowledgments** We are grateful to the Director, Directorate of Groundnut Research and the Indian Council of Agricultural Research for funding the research work.

## References

- Akkasaeng C, Vorasoot N, Jogloy S, Patanotai A (2003) Relationship between SPAD readings and chlorophyll contents in leaves of peanut (*Arachis hypogaea* L.). Thai J Agric Sci 36(3):279–284
- Arunyanark A, Jogloy S, Akkasaeng C, Vorasoot N, Kesmla T, Nageswararao RC, Wright GC, Patanothai A (2008) Chlorophyll stability is an indicator of drought tolerance in peanut. J Agron Crop Sci 194:113–125
- Baker NR, Horton P (1987) Physiological factors associated with fluorescence quenching during photoinhibition. In CJ Arntzen, DJ Kyle, CB Osmond (eds) Topics in Photosynthesis, Photoinhibition. Elsevier, Amsterdam, The Netherlands 9:145–168
- Bilger W, Bjorkman O (1990) Role of the xanthophyll cycle in photoprotection elucidated by measurements of light-induced absorbance changes, fluorescence and photosynthesis in leaves of *Hederacanariensis*. Photosynth Res 25:173–185
- Björkman O, Demmig-Adams B (1994) Regulation of photosynthetic light energy capture, conversion, and dissipation in leaves of higher plants. In: Schulze E-D, Caldwell MM (eds) Ecophysiology of photosynthesis. Springer, Berlin, pp 17–47
- Bogale K, Tesfaye T, Geleto (2011) Morphological and physiological attributes associated to drought tolerance of Ethiopian durum wheat genotypes under water deficit condition. J Biodiversity Environ Sci 1(2):22–36
- Clavel D, Drame NK, Roy Macauley H, Braconnier S, Laffray D (2005) Analysis of early responses to drought associated with field drought adaptation in four Sahelian groundnut (*Arachishypogaea* L.) cultivars. Environ Exp Bot 54:219–230
- Cornic G, Massacci A (1996) Leaf photosynthesis under stress. In: Baker RN (ed) Photosynthesis and the environment. Kluwer Academic Publishers, The Netherlands
- Daniele C, Omar D, Jean L, Khalfaoi SB (2006) Genotypes variations in fluorescence parameters among closely related groundnut (*Arachis hypogaea* L.) lines and their potential for drought screening programs. Field Crops Res 96:296–306
- Dwyer JF, Mcpherson EG, Schroeder HW, Rowntree RA (1992) Assessing the benefits and costs of the urban forest. J Arboric 18(5):227–234
- FAO (2012) <http://faostat.fao.org/site/567/DesktopDefault.aspx?PageID=567#ancor>. Accessed 15 Aug 2014
- Finazzi G, Johnson GN, Dall'osto L, Zito F, Bonente G, Bassi R (2006) Nonphotochemical quenching of chlorophyll fluorescence in *Chlamydomonas reinhardtii*. Biochemistry 45:1490–1498
- Flexas J, Medrano H (2002) Drought inhibition of photosynthesis in C3 plants: stomatal and non-stomatal limitation revisited. Ann Bot-London 89:183–189

- García-Plazaola JI, Becerril JM (1999) A rapid high-performance liquid chromatography method to measure lipophilic antioxidants in stressed plants: simultaneous determination of carotenoids and tocopherols. *Phytochem Anal* 10:307–313
- Geocleber Gomes De Sousa, Benito Moreira De Azevedo, Carlos Newdmar Vieira Fernandes, Thales Vinicius De Araújoviana, Marialilian Santos Silva (2014) Growth, gas exchange and yield of peanut in frequency of irrigation. *Rev Cienc Agron* 45 (1):27–34
- Kalariya KA, Singh AL, Chakraborty K, Zala PV, Patel CB (2013) Photosynthetic characteristics of groundnut (*Arachis hypogaea* L.) under water deficit stress. *Indian J Plant Physiol* 18(2):157–163
- Krause GH, Weis E (1991) Chlorophyll fluorescence and photosynthesis: the basics. *Annu Rev Plant Physiol Plant Mol Biol* 42:313–349
- Maxwell K, Johnson GN (2000) Chlorophyll fluorescence- a practical guide. *J Exp Bot* 51(345):659–668
- Mishra KB, Iannacone R, Petrozza A, Mishra A, Armentano N, La Vecchia G, Trtílek M, Cellini F, Nedbal L (2012) Engineered drought tolerance in tomato plants is reflected in chlorophyll fluorescence emission. *Plant Sci* 182:79–86
- Nautiyal PC, Ravindra V, Joshi YC (1995) Gas exchange and leaf water relations in two peanut cultivars of different drought tolerance. *Biol Plant* 37(3):371–374
- Nautiyal PC, Rachaputi N, Joshi YC (2002) Moisture-deficit induced changes in leaf-water content, leaf carbon exchange rate and biomass production in groundnut cultivars differing in specific leaf area. *Field Crops Res* 74:67–79
- Nautiyal PC, Ravindra V, Rathnakumar AL, Ajay BC, Zala PV (2012) Genetic variations in photosynthetic rate, pod yield and yield components in Spanish groundnut cultivars during three cropping seasons. *Field Crops Res* 125:83–91
- Onofri A (2007) Routine statistical analyses of field experiments by using an Excel extension. *Proceedings 6th National Conference Italian Biometric Society: “La statistica nell’escienze della vita e dell’ambiente”*, Pisa, p 93–96
- Polle JEW, Niyogi KK, Melis A (2001) Absence of lutein, violaxanthin and neoxanthin affects the functional chlorophyll antenna size of photosystem-II but not that of photosystem-I in the green alga *Chlamydomonas reinhardtii*. *Plant Cell Physiol* 42(5):482–491
- Richards A (2000) Selectable traits to increase crop photosynthesis and yield of grain crops. *J Exp Bot* 51(447–458):2000
- Richardson AD, Duigan SP, Berlyn GP (2002) An evaluation of non-invasive methods to estimate foliar chlorophyll content. *New Phytol* 153:185–194
- Singh AL, Nakar RN, Chakraborty K, Kalariya KA (2014) Physiological efficiencies in mini-core peanut germplasm accessions during summer season. *Photosynthetica* 52(4):627–635
- Subramaniam VB, Maheswari M (1990) Physiological responses of groundnut to water stress. *Indian J Plant Physiol* 33(2):130–135
- Vadez V, Kholova J, Medina S, Kakkera A, Anderberg H (2014) Transpiration efficiency: new insights into an old story. *J Exp Bot*. doi:10.1093/jxb/eru040