



First Complete Genome Sequence of a Distinct *Papaya Ringspot Virus* Isolate from the Northeastern Region of India

Ritesh Mishra,^a  Basavaprabhu L. Patil^a

^aICAR-National Research Centre on Plant Biotechnology, IARI, New Delhi, India

ABSTRACT This is the first report of a *Papaya ringspot virus* (PRSV) isolate from the northeastern region of India. The nucleotide sequence identity of PRSV-Meghalaya was in the range of 72.6 to 82.5% with other Indian PRSV isolates, and the highest identity of 84.4% was with a French isolate. Population genetic analysis indicated positive selection.

Papaya ringspot virus (PRSV) belongs to the family *Potyviridae* and the genus *Potyvirus*, with a positive-sense single-stranded RNA genome of ~10,349 nucleotides (nt) in size, encompassing a single open reading frame (ORF) (1–5). PRSV is classified as either PRSV-P or PRSV-W type, depending on whether it infects papaya or cucurbits, respectively (6–8). PRSV is hypothesized to have originated in India ~2,250 years ago, spread to East Asia ~600 years ago, and reached the Americas ~200 years ago (9). Although ~5 complete PRSV sequences have been published from India (10, 11), none of them are from the northeastern region (NER) of India.

In 2015, leaf samples from a PRSV-infected papaya plant were collected from Umiam (erstwhile Barapani) in the state of Meghalaya, located in the NER of India. Total RNA extraction and cDNA synthesis were done as described by Patil et al. (12). Later, the cDNA was used for PCR amplification with potyvirus universal primers (13) and degenerate primers from this study. Cloning and sequencing of the coat protein (CP) and helper component proteinase (HcPro) regions indicated it to be a distinct sequence. Later, degenerate primers were designed at conserved regions of PRSV, and the entire PRSV-Meghalaya genome was real-time PCR (RT-PCR) amplified as 5 segments of ~1.8 to 2.5 kb in size. These amplicons were cloned and sequenced as described by Patil et al. (12). The assembly of the contigs resulted in a complete genome size of 10,343 nt for the PRSV-Meghalaya isolate.

PRSV-Meghalaya was subjected to phylogenetic analysis, along with 21 full-length sequences of PRSV isolates from all over the world (12, 14, 15). PRSV-Meghalaya clustered with isolates from France, Mexico, Brazil, and the United States, in addition to all the other Indian PRSV isolates. It showed the highest nucleotide sequence identity of 84.4% with PRSV-France-E2, while the lowest sequence identity (70.6%) was with PRSV-South Korea (16), whereas the corresponding amino acid sequence identities were 89.5% with PRSV-France-E2 and 86.7% with PRSV-South Korea. PRSV-Meghalaya showed 82.5%, 81.7%, 77%, and 72.6% homology with nucleotide sequences of the four Indian isolates of PRSV (10, 11), namely, Pune, Delhi, Hyderabad, and Rajasthan, respectively.

To identify the most variable region of PRSV-Meghalaya, SimPlot was drawn for the sequences of 21 PRSV isolates (17). The genomic region of nt 241 to 1821 corresponding to P1 and nt 8841 to 9501 corresponding to the overlapping region of N1b-CP were the most variable. Recombination breakpoints were identified at 3 sites of PRSV-Meghalaya, namely nt 905 to 1134, nt 8844 to 9996, and nt 9205 to 10182, when analyzed using the RDP4 program (18). The degree of selective constraints at the amino acid level was estimated with MEGA 5.05 by analyzing separately the rate of nonsyn-

Received 13 April 2018 Accepted 17 April 2018 Published 31 May 2018

Citation Mishra R, Patil BL. 2018. First complete genome sequence of a distinct *Papaya ringspot virus* isolate from the northeastern region of India. *Genome Announc* 6:e00437-18. <https://doi.org/10.1128/genomeA.00437-18>.

Copyright © 2018 Mishra and Patil. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Basavaprabhu L. Patil, blpatil2046@gmail.com.

onymous (*dN*) and synonymous (*dS*) substitutions with the Pamilo-Bianchi-Li method (19), and DnaSP 5.10 was used to evaluate the importance of natural selection (20). The proportion of nucleotide substitution between the nonsynonymous and synonymous sites (*dN/dS*) was 1.8, indicating a positive selection for the amino acid sequence conservation in PRSV-Meghalaya (21). This is the first report of a full-length sequence of a PRSV isolate from the NER of India, indicating its independent evolution.

Accession number(s). The complete genome sequence of PRSV isolate Meghalaya has been deposited in NCBI GenBank with the accession number [MF356497](https://doi.org/10.1093/jgv/0.000740).

ACKNOWLEDGMENTS

We acknowledge the financial support from ICAR's Network Project on Transgenics in Crops (NPTC) and ICAR-NRCPB. R.M. acknowledges a postdoctoral fellowship from DST-SERB.

B.L.P. conceived the study idea, guided the entire work, analyzed the data, and wrote the manuscript. R.M. helped in cloning the PRSV sequences and in bioinformatics analysis.

REFERENCES

- Adams MJ, Antoniw JF, Beaudoin F. 2005. Overview and analysis of the polyprotein cleavage sites in the family *Potyviridae*. *Mol Plant Pathol* 6:471–487. <https://doi.org/10.1111/j.1364-3703.2005.00296.x>.
- Wylie SJ, Adams M, Chalam C, Kreuze J, López-Moya JJ, Ohshima K, Praveen S, Rabenstein F, Stenger D, Wang A, Zerbini FM, ICTV Report Consortium. 2017. ICTV virus taxonomy profile: *Potyviridae*. *J Gen Virol* 98:352–354. <https://doi.org/10.1099/jgv.0.000740>.
- Tripathi S, Suzuki JY, Ferreira SA, Gonsalves D. 2008. *Papaya ringspot virus-P*: characteristics, pathogenicity, sequence variability and control. *Mol Plant Pathol* 9:269–280. <https://doi.org/10.1111/j.1364-3703.2008.00467.x>.
- Yeh SD, Jan FJ, Chiang CH, Doong TJ, Chen MC, Chung PH, Bau HJ. 1992. Complete nucleotide sequence and genetic organization of papaya virus RNA. *J Gen Virol* 73:2531–2541. <https://doi.org/10.1099/0022-1317-73-10-2531>.
- Yeh SD, Gonsalves D. 1985. Translation of papaya ringspot virus RNA *in vitro*: detection of a possible polyprotein that is processed for capsid protein, cylindrical-inclusion protein, and amorphous-inclusion protein. *Virology* 143:260–271.
- Gonsalves D. 1998. Control of papaya ringspot virus in papaya: a case study. *Annu Rev Phytopathol* 36:415–417. <https://doi.org/10.1146/annurev.phyto.36.1.415>.
- Purcifull DE, Hiebert E. 1979. Serological distinction of watermelon mosaic virus isolates. *Phytopathology* 69:112–116. <https://doi.org/10.1094/Phyto-69-112>.
- Purcifull DE, Edwardson JR, Hiebert E, Gonsalves D. 1984. Papaya ringspot virus, p 292. *In* CMI/AAB Descriptions of Plant Viruses. Association of Applied Biologists, Warwick Enterprise Park, Wellesbourne, Warwick, UK.
- Olarte Castillo XA, Fermin G, Tabima J, Rojas Y, Tennant PF, Fuchs M, Sierra R, Bernal AJ, Restrepo S. 2011. Phylogeography and molecular epidemiology of *Papaya ringspot virus*. *Virus Res* 159:132–140. <https://doi.org/10.1016/j.virusres.2011.04.011>.
- Jain RK, Pappu HR, Pappu SS, Varma A, Ram RD. 1998. Molecular characterisation of papaya ringspot potyvirus isolates from India. *Ann Appl Biol* 132:413–425. <https://doi.org/10.1111/j.1744-7348.1998.tb05218.x>.
- Jain RK, Sharma J, Siva Kumar AS, Sharma PK, Byadgi AS, Verma AK, Varma A. 2004. Variability in the coat protein gene of *Papaya ringspot virus* isolates from multiple locations in India. *Arch Virol* 149:2435–2442. <https://doi.org/10.1007/s00705-004-0392-x>.
- Patil BL, Dangwal M, Mishra R. 2017. Variability of emaravirus species associated with sterility mosaic disease of pigeonpea in India provides evidence of segment reassortment. *Viruses* 9:183. <https://doi.org/10.3390/v9070183>.
- Gibbs A, Mackenzie A. 1997. A primer pair for amplifying part of the genome of all potyvirids by RT-PCR. *J Virol Methods* 63:9–16. [https://doi.org/10.1016/S0166-0934\(96\)02103-9](https://doi.org/10.1016/S0166-0934(96)02103-9).
- Jones DT, Taylor WR, Thornton JM. 1992. The rapid generation of mutation data matrices from protein sequences. *Comput Appl Biosci* 8:275–282. <https://doi.org/10.1093/bioinformatics/8.3.275>.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* 30:2725–2729. <https://doi.org/10.1093/molbev/mst197>.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser* 41:95–98.
- Lole KS, Bollinger RC, Paranjape RS, Gadkari D, Kulkarni SS, Novak NG, Ingersoll R, Sheppard HW, Ray SC. 1999. Full-length human immunodeficiency virus type 1 genomes from subtype C-infected seroconverters in India, with evidence of intersubtype recombination. *J Virol* 73:152–160.
- Martin DP, Murrell B, Golden M, Khoosal A, Muhire B. 2015. RDP4: detection and analysis of recombination patterns in virus genomes. *Virus Evol* 1:vev003. <https://doi.org/10.1093/ve/vev003>.
- Pamilo P, Bianchi NO. 1993. Evolution of the Zfx and Zfy genes: rates and interdependence between the genes. *Mol Biol Evol* 10:271–281. <https://doi.org/10.1093/oxfordjournals.molbev.a040003>.
- Librado P, Rozas J. 2009. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25:1451–1452. <https://doi.org/10.1093/bioinformatics/btp187>.
- Kryazhimskiy S, Plotkin JB. 2008. The population genetics of *dN/dS*. *PLoS Genet* 4:e1000304. <https://doi.org/10.1371/journal.pgen.1000304>.