



First Draft Genome Sequence of *Salmonella enterica* Serovar Gallinarum Strain VTCCBAA614, Isolated from Chicken in India

R. K. Vaid,^a N. Jindal,^b T. Anand,^a B. C. Bera,^a T. Riyesh,^a N. Virmani,^c S. Barua,^a Renu Gupta,^b N. K. Mahajan,^b C. G. Joshi,^d R. K. Singh^{a,c*}

Veterinary Type Culture Collection, ICAR-National Research Centre on Equines, Hisar, Haryana, India^a; Department of Veterinary Public Health and Epidemiology, College of Veterinary Sciences, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana, India^b; ICAR-National Research Centre on Equines, Hisar, Haryana, India^c; Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, Gujarat, India^d

* Present address: R. K. Singh, Indian Veterinary Research Institute, Izatnagar, Bareilly, Uttar Pradesh, India

Salmonella enterica subsp. enterica serovar Gallinarum biovar Gallinarum causes fowl typhoid (FT), which results in huge economic losses to poultry farmers in India. We report the draft genome sequence of Salmonella biovar Gallinarum strain VTC-CBAA614, isolated from a chicken in an FT affected broiler flock.

Received 7 September 2015 Accepted 11 September 2015 Published 22 October 2015

Citation Vaid RK, Jindal N, Anand T, Bera BC, Riyesh T, Virmani N, Barua S, Gupta R, Mahajan NK, Joshi CG, Singh RK. 2015. First draft genome sequence of Salmonella enterica serovar Gallinarum strain VTCCBAA614, isolated from chicken in India. Genome Announc 3(5):e01221-15. doi:10.1128/genomeA.01221-15.

Copyright © 2015 Vaid et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to R. K. Vaid, rk_vaid@yahoo.com.

F owl typhoid (FT) is an acute septicemic disease of pullets and adult poultry caused by a nonmotile invasive avian pathogen, *Salmonella enterica* subsp. *enterica* serovar Gallinarum biovar Gallinarum (1). The disease is responsible for considerable economic losses in breeding and commercial laying flocks causing high morbidity and acute mortality in Asia, although disease incidence may be underreported elsewhere (2–4). Disease transmission occurs horizontally through the spread of infectious agents from carrier birds via contaminated feed and water, although transovarian transmission also takes place (5). The pathological changes in acute FT are primarily observed in the liver (1).

The serogroup D nonmotile *Salmonella* biovar Gallinarum is a chicken-adapted biotype capable of causing systemic infection rather than enteritis in birds, in contrast to its motile ancestor *S. enteritidis* (6). The analysis of the close phylogenomic relationship between *Salmonella* biovar Gallinarum and *S. enteritidis* has elucidated a common ancestry (7); however, strains with intermediate biochemical markers among nonmotile salmonellae have been observed (8). The disease may have been eradicated in North America and Europe, but it is endemic throughout India (2, 3, 9). This is the first whole-genome sequence of an FT outbreak *Salmonella* biovar Gallinarum isolate from India. Strain VTCCBAA614 was isolated from the liver of a broiler chicken (*Gallus gallus domesticus*) (2).

Sequencing was achieved by 454 pyrosequencing of a shotgun library and assembled *de novo* using Newbler version 2.6. A total of 2,23,420 reads of 427 bp were generated using the GS FLX Titanium system, giving ~20× coverage. The data generated 92 contigs with an average contig size of 61,644 bp and a largest contig size of 3,02,540 bp. The total size of the genome was 46,86,634 bp, with an N_{50} of 1,25,446 bp and a Q40 of 99.75%. The annotation was carried out against strain Ty2 using the RAST server, which showed GC content of 52.20% with 4,581 predicted genes (10) The PGAP analysis data contains 4,186 coding sequences (CDSs),

314 pseudogenes, 4 rRNAs, 66 tRNAs, 11 ncRNAs, and 204 frameshifted genes. The Pathosystems Resource Integration Center (PATRIC) annotation reported 173 pseudogenes and 675 hypothetical proteins (11).

SEED subsystem analysis revealed genes involved in multifarious roles, including virulence, adhesion, bacteriocins, and resistance to antimicrobials and toxic compounds (12). Antimicrobial resistance in *Salmonella* biovar Gallinarum is an emerging concern (13). Strain VTCCBAA614 revealed 55 genes for antimicrobial resistance, including a lactam utilization protein, penicillinbinding protein, polymyxin resistance protein, macrolide-specific efflux protein, and proteins responsible for flouroquinolone and tetracycline resistance.

Six other genomes of *Salmonella* serovar Gallinarum strains SG9, 287/92, FCAV198, CDC1983-67, 9184, and RKS5078 have been sequenced from North America, Brazil, and the United Kingdom, and VTCCBAA614 is the only strain sequenced from the Asian region. Although the genomic relatedness of *Salmonella* serovar Gallinarum strains from distinct geographies is open to comparison, a greater number of sequences will bring further clarity to the pseudogene contents of the two nonmotile strains. The genomic data may be useful in providing insight into the pathogenesis, biology, and mechanisms of host tropism, and may provide clues for smart vaccines.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at GenBank under the accession number JSWQ00000000. The version described in this paper is the first version, JSWQ01000000.

ACKNOWLEDGMENT

This work was supported by Veterinary Type Culture Collection grant IXX00269, from the Indian Council of Agricultural Research, National Research Centre on Equines, Hisar.

REFERENCES

- 1. Shivaprasad HL. 2000. Fowl typhoid and pullorum disease. Rev Sci Tech 19:405–424.
- Kumar T, Mahajan NK, Rakha NK. 2010. Epidemiology of fowl typhoid in Haryana, India. Worlds Poult Sci J 66:503–510. http://dx.doi.org/ 10.1017/S0043933910000565.
- 3. Rajagopal R, Mini M. 2013. Outbreaks of salmonellosis in three different poultry farms of Kerala, India. Asian Pac J Trop Biomed 3:496–500. http://dx.doi.org/10.1016/S2221-1691(13)60103-3.
- Barrow PA, Freitas Neto OC. 2011. Pullorum disease and fowl typhoid new thoughts on old diseases: a review. Avian Pathol 40:1–13. http:// dx.doi.org/10.1080/03079457.2010.542575.
- Hall WJ, Legenhausen DH, Macdonald AD. 1949. Studies on fowl typhoid I: nature and dissemination. Poult Sci 28:344-362. http:// dx.doi.org/10.3382/ps.0280344.
- McMeechan A, Lovell MA, Cogan TA, Marston KL, Humphrey TJ, Barrow PA. 2005. Glycogen production by different *Salmonella enterica* serotypes: contribution of functional *glgC* to virulence, intestinal colonization and environmental survival. Microbiology 151:3969–3977. http:// dx.doi.org/10.1099/mic.0.28292-0.
- 7. Thomson NR, Clayton DJ, Windhorst D, Vernikos G, Davidson S, Churcher C, Quail MA, Stevens M, Jones MA, Watson M, Barron A, Layton A, Pickard D, Kingsley RA, Bignell A, Clark L, Harris B, Ormond D, Abdellah Z, Brooks K, Cherevach I, Chillingworth T, Woodward J, Norberczak H, Lord A, Arrowsmith C, Jagels K, Moule S, Mungall K, Sanders M, Whitehead S, Chabalgoity JA, Maskell D, Humphrey T, Roberts M, Barrow PA, Dougan G, Parkhill J. 2008. Comparative genome analysis of *Salmonella* Entertidis PT4 and *Salmonella* Gallinarum 287/91 provides insights into evolutionary and host ad-

aptation pathways. Genome Res 18:1624-1637. http://dx.doi.org/ 10.1101/gr.077404.108.

- Crichton PB, Old DC. 1990. Salmonellae of serotypes Gallinarum and Pullorum grouped by biotyping and fimbrial-gene probing. J Med Microbiol 32:145–152. http://dx.doi.org/10.1099/00222615-32-3-145.
- Renu R, Yadav AS, Tripathi V, Singh RP. 2011. Salmonella occurrence in chicken eggs and environmental samples and their sero-prevalence in laying hens. Indian J Anim Sci 81:1087–1088.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- 11. Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJC, Yoo HS, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. Nucleic Acids Res 42:D581–D591. http://dx.doi.org/10.1093/nar/gkt1099.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42: D206–D214. http://dx.doi.org/10.1093/nar/gkt1226.
- 13. Lee YJ, Kim KS, Kim JH, Tak RB. 2004. *Salmonella gallinarum gyrA* mutations associated with fluoroquinolone resistance. Avian Pathol **33**: 251–257.