RESEARCH COMMUNICATIONS

experimental findings clearly indicate that the higher age groups (yearlings) of IMC are able to resist artificial infection with *A. invadans* (whereas the same age groups of snakehead were susceptible).

- Baldoc, F. C., Blazer, V., Callinan, R., Hatai, K., Karunasagar, I., Mohan, C. V. and Bondad-Reantaso, M. G., Outcomes of a short expert consultation on epizootic ulcerative syndrome (EUS): Reexamination of causal factors, case definition and nomenclature. In *Diseases in Asian Aquaculture – V* (eds Walker, P. and Bondad-Reantaso, M. G.), Fish Health Section, Asian Fisheries Society, Manila, 2005, pp. 555–585.
- Thompson, K. D., Lilley, J. H., Chen, S. C., Adams, A. and Richards, R. H., The immune response of rainbow trout (*Oncorhynchus* mykiss) against *Aphanomyces invadans*. Fish Shellfish Immunol., 1999, 9, 195–201.
- Johnson, R. A., Zabrecky, J., Kiryu, Y. and Shelds, J. D., Infection experiments with *Aphanomyces invadans* in four species of estuarine fish. J. Fish Dis., 2004, 27, 287–295.
- Lilley, J. H., Callinan, R. B., Chinabut, S., Kanchanakhan, S., MacRae, I. H. and Phillips, M. J., *Epizootic Ulcerative Syndrome* (EUS) Technical Handbook, AAHRI, Bangkok, Thailand, 1998, p. 88.
- 5. Jayaraman, R., UDS strikes fish in Tamil Nadu. *Fish. Chim.*, 1991, **11**, 30–31.
- Vishwanath, T. S., Mohan, C. V. and Shankar, K. M., Mycotic granulomatosis and seasonality are the consistent features of Epizootic Ulcerative Syndrome (EUS) of fresh and brackishwater fishes of Karnataka, India. *Asian Fish. Sci.*, 1997, 10, 155–160.
- Vishwanath, T. S., Mohan, C. V. and Shankar, K. M., Epizootic ulcerative syndrome (EUS) associated with a fungal pathogen, in Indian fishes: Histopathology – 'a cause for invasiveness'. *Aquaculture*, 1998, **165**, 1–9.
- Das, M. K., Epizootic Ulcerative Syndrome (EUS) in fishes its present status in India. CICFRI (ICAR), 1997, Bull. No. 69, p. 22.
- Kumar, D., Dey, R. K. and Sinha, A., Outbreak of Epizootic Ulcerative Syndrome (EUS) of fish in India. In *Aquaculture Productivity* (eds Sinha, V. R. P. and Srivastava, H. C.), Oxford and IBH, New Delhi, 1991, pp. 345–356.
- Roberts, R. J., Campbell, B. and MacRae, I. H., In Proceedings of the Regional Seminar on Epizootic Ulcerative Syndrome (EUS). The Aquatic Animal Health Research Institute, Bangkok, Thailand, 25–27 January 1994, p. 282.
- Chinabut, S. and Roberts, R. J., Pathology and Histopathology of Epizootic Ulcerative Syndrome (EUS), AAHRI, Bangkok, Thailand, 1999, p. 33.
- Chinabut, S., Roberts, R. J., Willoughby, L. G. and Pearson, M. D., Histopathology of snakehead, *Channa striatus* (Bloch), experimentally infected with the specific *Aphanomyces* fungus associated with Epizootic Ulcerative Syndrome (EUS) at different temperatures. J. Fish Dis., 1995, 18, 41–47.
- 13. Kurata, O., Kanai, K. and Hatai, K., Fungicidal activity of carp serum against *Aphanomyces piscicida*. *Fish Pathol.*, 2000, **35**, 49–50.
- 14. Papadimitriou, J. M. and Spector, W. G., The origin, properties and fate of epithelioid cells. *J. Pathol.*, 1971, **105**, 187–203.
- Spector, W. G. and Mariano, M., Macrophage behavior in experimental granulomas. In *Mononuclear Phagocytes in Immunity*, *Infection and Pathology* (ed. Vanfurth, R.), Blackwell Scientific Publications, Oxford, 1975, pp. 927–942.
- 16. Mohan, C. V., Inflammatory response of Indian major carps to *A. invadans*, fungal pathogen of EUS. IFS report, 2003, p. 36.

ACKNOWLEDGEMENT. We thank Dr J. H. Lilley for kindly providing the *Aphanomyces invadans* (B99C) isolate.

Received 22 February 2006; revised accepted 21 January 2007

Weed floristic composition in palm gardens in Plains of Eastern Himalayan region of West Bengal

Arun Kumar Sit^{1,*}, Malay Bhattacharya¹, Biswanath Sarkar¹ and V. Arunachalam²

 ¹Central Plantation Crops Research Institute, Research Centre, Mohitnagar, Jalpaiguri 735 102, India
 ²Central Plantation Crops Research Institute, Kasargod 671 124, India

Weeds are unwanted plants in the crop land that compete for nutrients, water and space. Proper knowledge about weed flora is important for their management. Weather conditions in sub-Himalayan West Bengal favour weed growth. Therefore, a study was conducted to find out the weed floristic composition of different palm gardens in this region. The results showed that dicots were predominant in the palm gardens. Maximum number of weeds was found in the oil palm gardens and the least in the fruiting arecanut gardens. A total of 20 angiosperm families were found in the study area. Among them, 17 belonged to dicots and three to monocots. A total of five pteridophytes were found. Members of Poaceae, Asteraceae, Oxalidaceae and Urticaceae were found in all the plots studied. Three species, viz. Ageratum conyzoides, Oxalis corniculata and Vandelia were found to be more widely distributed in all four palms as well as in fallow land, showing Shannon's index value >0.75. Prevalence of some weeds in all the study areas revealed that they can grow under any conditions.

Keywords: Control, plains, Eastern Himalayas, palm gardens, weeds.

WEEDS compete with other crops for water, nutrients and space. Weeds also act as alternate hosts for pests and diseases. The abundance or distribution of weed species in a cropped field varies due to the nature of the crop, cultural practices and cropping pattern/system, soil type, moisture availability, location and season. Knowledge of weed flora enables one to use the required herbicide and formulate other suitable management strategies. It is also useful in exploiting abundant weeds as a cover crop or pasture and for other economic uses. Extensive literature is available on weed flora dynamics in field $crops^{1-3}$, as well as on weed flora in palms grown as plantations^{4,5}, date palm^{6,7}, arecanut⁸ and peach palm^{9,10}. Control of weeds by handweeding or application of herbicides is essential for better crop yield. However, one should have a clear idea about the existence of different weed flora under the shade of different plantation crops like arecanut, coconut and oil palm. According to Derksen et al.¹, the study of weed dynamics is essential to formulate a management strategy for

^{*}For correspondence. (e-mail: sitarunkumar@yahoo.com)

the cropping system in the Northern Great plains of Canada. The present investigation was carried out to find out weed floristic composition in palm gardens in the plains of the eastern Himalayan region, West Bengal.

The study was conducted at the CPCRI Research Centre, Mohitnagar located at 26°N lat., 88°E long., with a mean altitude of 91.3 m asl. Agro-ecologically, it is called the sub-Himalayan Terai region. It receives an annual rainfall of more than 4000 mm and maximum rainfall occurs between April (995 mm) and September (996.5 mm). The intensity of rainfall then gradually decreases up to October and there is only occasional rainfall during the rest of the year. The maximum and minimum temperature varies between 5.5 and 37°C. The soil is predominantly sandy loam of Teesta alluvial with a pH of 5.5–6.5. The area under study is cropped with arecanut, coconut and oil palms.

Five different areas, viz. those with coconut (Cocos nucifera L), oil palm (Elaeis guineensis, Jacq.), adult arecanut (Areca catechu L.) (more than 30-yr-old), prebearing arecanut garden (three-yr-old), and fallow land were selected for weed study. Oil palm, coconut and arecanut were planted at a spacing of $10 \text{ m} \times 10 \text{ m}$, $6 \text{ m} \times 10 \text{ m}$ 6 m and 2.7 m \times 2.7 m respectively. Adult oil palm was maintained at 20-25 leaf/palm. The study was conducted during September 2004, just after the rainy season. Quadrates of 10 sq. m area were laid down in different locations in a random manner. For each type of field, ten quadrates were included for weed study. Various species and the total number of individuals of each species were noted. Average data of ten replications/quadrate were considered for statistical analysis. Frequency, abundance, density (per sq. m) of the species noted were calculated using the formula by Sharma¹¹.

On the basis of per cent frequency values for each type of field, the various species were distributed into five frequency classes. Representation of each of the five frequency classes in the total number of species was calculated using the formula by Sharma¹¹.

The proportion (P_i) of occurrence of each species or each plot was used in each trait (*i*) and the Shannon– Weaver estimates¹² were worked out using the formula:

$$H' = -\sum_{i=1}^n P_i \log_2 P_i,$$

where *n* is the total number of species/plot, P_i is the proportion of individuals in the *i*th species/plot. The index value calculated is divided by $\log_2 n$ to get equivalence.

A similarity index has been worked out using the formula $SI = 2C/(A + B)^{13}$, where C is the number of similar weed species occurring in palm garden/fallow land compared, A is the number of weed species occurring in the first plot and B is the number of weed species occurring in the second plot being compared.

Frequency, abundance and density of different weed species in different cropped areas as well as fallow land

are given in Table 1. Forty-four weed species were found in different cropped areas as well as fallow land. The number and type of weed varied in different locations of the area under study. Maximum number of weeds was present in the fallow land (24), followed by oil palm (23), young arecanut – 3-yr-old (21), coconut field (15), and adult arecanut – fruiting stage (14). The number of weed species studied in this experiment revealed that the growth of different weed species depends on the canopy spread of plantation.

Among the 24 weed species in fallow land, maximum frequency of weed population (100%) was recorded for Andropogon aciculate, Boreria alata, Brachiaria sp., Centella asiatica, Clerodendron infortunetum, Cyperus spp., Leucus aspera, Melastoma sp. and Oxalis corniculata. About 90% frequency was recorded for Cynodon dactylon, Desmodium trifoliatum, Ageratum conyzoides, Boraria sp., Cynodon dactylon and Imperata cylindrica. In coconut field, A. conyzoides, Borreria sp., Centella asiatica, Gnaphalium sp., O. corniculata, Solanum nigrum and Vandelia sp. occurred with 100% frequency. Ninety per cent frequency was observed for C. infortunetum, Dryopteris sp., I. cylindrica and Melastoma sp. Weed cover of the adult arecanut garden was different from that of the other fields. Weeds like Colocasia sp., Drymeria sp. and Stelaria media were present only in the adult arecanut field. In the case of the oil palm field, maximum frequency (100%) of weeds like A. conyzoides, Brachiaria sp., C. dactylon, D. trifoliatum, D. sanginalis, Rungia sp. and Spermacocci latifolia was recorded. I. cylindrica and Melastoma sp. were found to have 90% frequency. Among oil palms, rubber and coconut plantations, pasture establishment and livestock establishment seem to be the best in coconut gardens due to the lack of competition for light between pasture grass/legume and coconut¹⁴. A. conyzoides and O. corniculata occurred in all the fields with different frequencies, revealing that they can grow in any situation irrespective of shade. The wild Colocasia sp. was found only in the arecanut field. Drosera sp., Mimosa pudica, Pteridium sp. and Pteris sp. were found only in fallow land, which indicates that they require full sunlight for their growth and development.

Figure 1 shows that about 37.93, 80 and 64.28% weeds were recorded at 81–100% frequency range in fallow, co

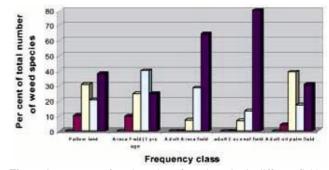


Figure 1. Per cent of total number of weed species in different fields.

		Fallow land		Juven	Juvenile arecanut field (3-yr-old)	: field	Adu	Adult arecanut field	t field	Adu	Adult coconut field	field	õ	Oil palm field		10
Weed species	н	А	D	ц	Α	D	Ц	A	D	н	Α	D	Ц	А	D	index
Ageratum conyzoides (A)	70	63.0	4.41	100	165.2	16.52	70	13.6	0.95	100	135.6	13.56	100	74.8	7.48	0.820
Andropogon aciculate (P)	100	351.5	35.15		I	I	I	I	I	I	I	I	70	44.9	3.14	0.176
Borreria alata (R)	100	50.1 201 2	5.01	100	25.0 22 ī	2.50				100	29.9	2.99	60 190	5.16	0.31	0.716
Brachiaria sp. (C)	100	591.3	5.913	20	23.5	1.17	100	70.9	7.09		1 1		100	119.9	96.11	0.486
Centella asiatica (Ap)	100	60.3 22.0	60.3	60	35.83	2.15	I	I	I	001	16.9	1.69	08 6	13.0	1.04	0.721
Clerodendron infortunetum (V)	100	23.9	23.9	I	I	I	I	0 1	;	96	10.1	16.	0/	6.9	0.48	0000
<i>Colocasia</i> sp. (Ar)			1	I	I	I	60	1.8	0.11	I	I	I	I	I	I	0.000
Cymbopogon citrates (P)	50	12.0	6.0	L	l	I.	I	I	I	I		I	L	l	I :	0.000
Cynodon dactylon (P)	90	438.2	394.4	100	710.9	71.09	I :	I ;	I	I	I	I	100	143.0	14.30	0.580
Cyperus spp. (C)	100	197.6	197.6	I	I	I	90	39.7	3.57	I	I	I	I	I	I	0.266
Desmodium trifoliatum (F)	90	113.5	102.2	70	103.3	7.23	I	I	I	I	I	I	100	101.2	10.12	0.675
Digitaria sanguilalis (P)	I	I	I	80	25.4	2.03	80	13.4	1.07	I	I	I	100	797.3	79.73	0.114
Drosera sp. (D)	60	59.5	35.7	I	I	I	I	I	I	I	I	I	I	I	I	0.000
Drymeria sp. (Ca)	I	I	I	I	I	I	100	194.0	19.40	I	I	I	I	I	I	0.000
Dryopteris sp. (Pter)	60	23.66	14.2	I	I	I	I	I	I	90	38.1	3.43	50	27.0	1.35	0.444
Echinocloa colonum (P)	I	I	I	50	5.8	0.29	I	I	I	I	I	I	40	51.0	2.01	0.235
Elucine indica (P)	I	I	I	80	19.4	1.55	I	I	I	I	I	I	I	I	I	0.000
Emelia sonchifolia (A)	I	I	I	Ι	I	I	100	24.1	2.41	Ι	I	I	I	I	I	0.000
Euphorbia hirta (E)	80	27.87	22.3	80	29.8	2.38	I	I	I	I	I	I	I		I	0.430
Gnaphalium sp. (A)	I	I	I	I	I	I	100	74.5	7.45	100	32.3	3.23	I	I	I	0.38
Hyptis sp. (L)	60	2.83	1.7	I	I	I	I	I	I	I	I	I	I	I	I	0.000
Imperata cylindrica (P)	60	166.5	6.66	100	393.6	39.36	I	I	I	90	36.7	3.30	90	79.3	7.14	0.61
Leucus aspera (L)	100	71.4	71.4	30	3.3	0.10	I	I	I	I	I	I	80	10.5	0.84	0.248
Lindenbergia sp. (S)	I	I	I	60	21.33	1.28	I	I	I	I	I	I	I	I	I	0.00
Lygodium sp. (Pter)	70	6.71	4.7	I	I	I	I	I	I	I	I	I	50	2.6	0.13	0.325
<i>Melastoma</i> sp. (M)	100	28.6	28.6	40	2.3	0.09	I	I	I	90	7.9	0.71	90	8.0	0.72	0.590
Micania micrantha (A)	30	6.0	1.8	50	2.6	0.13	I	I	I	I	I	I	50	3.0	0.15	0.677
Mimosa pudica (F)	60 100	10.33	6. 2		00	1 0						(•	1	0		0.00
Dxalls corniculata (U)	100	104.9	104.9	001	95.4 51.0	40.6 22 c	001	0.767	07.67	001	40.5 7 7 1	4.03 4.05	00	0.0	0.80	101.0
I uucoigiu sp. (U) Dolvacanua currantala (Do)	- 09	10.33	و م	2	0.10	10.0 	2	0.1		00	0.01	CC.1	8	0.0		1/0.0
Ptoridium sn. (Ptor)	40	6.05	1.0 V V			1										0000
Previx sp. (1 cc.)	9 06	1 33	040	I	I	I	I	I	I	I	I	I	I	I	I	0.000
Rungia sp. (Ac)			5 I	I	I	I	I	I	I	100	74.1	7.41	100	25.6	2.56	0.354
Saccharum spontaneum (P)	I	I	I	80	14.6	1.17	I	I	I	80	21.8	1.74	I	I	I	0.419
Scoparia dulsis (S)	80	10.75	8.6	60	3.5	0.21	I	I	I	I	I	I	I	I	I	0.308
<i>Selaginella</i> sp. (pter)	I	I	I	I	I	I	I	I	I	60	234.3	14.06	I	I	I	0.000
Solanum nigrum (So)	ı	I	I	70	3.9	.27	70	7.3	0.51	100	11.6	1.16	I	I	I	0.580
Spermacocci latifolia (R)	70	13.85	9.7	I	I	I	100	15.4	1.54	I	I	I	100	19.7	1.97	0.658
Spilanthes sp. (A)	I	I	I	I	I	I	I	I	I	I	I	I	60	4.2	0.25	0.000
Stelaria media (Ca)	I	I	I	I	I	I	100	159.2	15.92	I	I	I	I	I	I	0.000
Triumfetta rhomboidea (T)	50	4.4	2.2	L	I ;	I I	L	I	1	L	I	I	50	1.8	0.09	0.374
Vandelia sp. (S)	60	39.0	23.4	80	69.3	5.54	100	25.5	2.59	100	71.3	7.13	L ;	1	1	0.795
Vernonia cinera (A)	70	15.28	10.7	I	I	1	I	I		I	I	1	60	6.5	0.39	0.36]
Shannon's index			0.645			0.488			0.515			0.618			0.468	

RESEARCH COMMUNICATIONS

	Fallow land	Juvenile arecanut garden	Adult arecanut garden	Adult coconut garden	Adult oil palm garden
Ageratum conyzoides	\checkmark	\checkmark		\checkmark	
Andropogon aciculate	\checkmark	_	_	_	\checkmark
Borreria alata	\checkmark		_		\checkmark
Brachiaria sp.	\checkmark	\checkmark	\checkmark	_	\checkmark
Centella asiatica	\checkmark		_		\checkmark
Clerodendron infortunetum	\checkmark	-	-		\checkmark
Colocasia sp.	-		\checkmark	_	-
Cymbopogon citrates	\checkmark	-	-	_	-
Cynodon dactylon	\checkmark		-	_	\checkmark
Cyperus spp.	\checkmark	-	\checkmark	_	_
Desmodium trifoliatum	\checkmark		-	_	\checkmark
Digitaria sanguilalis	_		\checkmark	-	\checkmark
Drosera sp.	\checkmark	-	-	_	-
Drymeria sp.	-	-	\checkmark	-	-
Dryopteris sp.	\checkmark	-	-		\checkmark
Echinocloa colonum	_		-	-	\checkmark
Elucine indica	_		-	-	_
Emelia sonchifolia	-	-	\checkmark	_	-
Euphorbia hirta	\checkmark		-	_	-
Gnaphalium sp.	_	-	\checkmark		_
Hyptis sp.	\checkmark	-	-	_	-
Imperata cylindrica	\checkmark		-		\checkmark
Leucus aspera	\checkmark		-	-	\checkmark
Lindenbergia sp.	-		-	-	-
Lygodium sp.	\checkmark	-	-	-	\checkmark
Melastoma sp.	\checkmark		-		
Micania micrantha	\checkmark		-	_	\checkmark
Mimosa pudica	\checkmark	-	-	-	-
Oxalis corniculata	\checkmark		\checkmark		\checkmark
Pauzolgia sp.	_		\checkmark	\checkmark	\checkmark
Polygonum aurantale		-	-	_	_
Pteridium sp.		-	-	-	-
Pteris sp.	\checkmark	-	-	_	-
Rungia sp.	-	_	-		\checkmark
Saccharum spontaneum	_		-	\checkmark	-
a	1	1			

 $\sqrt{}$

 $\sqrt{}$

_

 $\sqrt{}$

 $\sqrt{}$

Table 2. Weed species in different crop lands

' $\sqrt{}$ ' Presence and '-' absence of weed species in respective fields.

 $\sqrt{}$

 $\sqrt{}$

conut and adult arecanut fields respectively, whereas in young arecanut gardens 40% weeds were in the 61-80% frequency range.

Scoparia dulsis Selaginella sp.

Solanum nigrum Spermacocci latifolia Spilanthes sp. Stelaria media

Vandelia sp.

Vernonia cinera

Triumfetta rhomboidea

The density of the weed species varied considerably in the different fields. Maximum density of weed in fallow land was recorded for Bracharia (59.13) and minimum for Pteris sp. (0.4). In the arecanut field (3-yr-old), maximum density was recorded for C. dactylon (71.09) and minimum for Melastoma sp. (0.09), whereas in the adult arecanut field, maximum weed density was recorded for O. corniculata (29.26) and minimum for Colocasia sp. (0.11). In the coconut field, maximum density was recorded for Selaginella sp. (14.06) and minimum for Melastoma (0.71). In oil palm gardens, maximum weed density was recorded for Digitaria sanguinalis (79.73) and the minimum for Triumfetta rhomboidea (0.09).

V

λ

_ √

Maximum abundance of different weed species in fallow land, young arecanut garden, adult arecanut, coconut and oil palm gardens was recorded as C. dactylon (438.2), C. dactylon (710.9), O. corniculata (292.6), Selaginella sp. (234.3) and C. dactylon (143.0) respectively, whereas minimum abundance was recorded as Pteris (1.33), Melastoma sp. (2.3), Colocasia sp. (1.8), Melastoma sp. (7.9) and Triumfetta rhomboidea (1.8), respectively.

RESEARCH COMMUNICATIONS

Among the 44 weed species, only two (A. conyzoides and Oxalis sp.) were present in all the fields irrespective of the crops whereas, seven weeds (Cymbopogon citrate, Drosera sp., Hyptis sp., M. pudica, Polygonum aurantale, Pteridium sp. and Pteris sp.) were present in fallow land only (Table 2). This reveals the fact that full sunlight is an essential requirement for the growth of these seven weeds. Pauzolgia sp. was present in all the palm gardens but not in fallow land. Two weeds (Elicine indica and Linderbergia sp.) are specific to young arecanut gardens, whereas Colocasia sp., Drymeria sp., Emelia sonchifolia and Stelaria medica were present only in adult arecanut plantations. Selaginella sp. was present only in the coconut field and Euphorbia hirta and Scoparia dulsis were found only in young arecanut gardens and fallow land. Cyprus spp. was present only in the adult arecanut plantations and fallow land. Spilantes sp. was present in oil palm garden, but Vandelia sp. was absent in this garden. Weed species like A. aciculate, Lygodium sp., T. rhomboidea and Vernonia cinera were present only in adult arecanut plantations. Such weeds need to be observed in many palm gardens and their specific association needs to be understood by detailed investigations. It was observed that when weed species are grown under shade conditions, the plant part is soft when compared to those in open fields where the plant parts are relatively hardy.

Shannon's index of weed flora diversity was observed for oil palm (0.468) and young arecanut garden (0.488), whereas it was high in fallow land (0.645). In this study, coconut gardens have recorded high weed flora diversity (0.618) next only to fallow land (0.465). Of the 44 weed species in the study, 15 are limited in distribution among the systems investigated. They are shown in Table 1, with a Shannon's index value of 0. Only three species were found to be distributed at greater frequency in all four palms, except oil palm where Vandelia sp. was not found, as well as in fallow land which shows Shannon's index value >0.75. They are A. conyzoides, O. corniculata and Vandelia sp. These need special attention as general weeds in humid tropics in the palm gardens. A. conyzoides prevailed as the dominant weed species in glyphosatetreated peach palm gardens⁹.

There are many reports on light penetration in canopies of different palm species. About 47.8% of light penetrates the canopy of adult arecanut palm, which is more than 50% of light penetration in young arecanut palm¹⁵. It is 43% in young coconut palm¹⁶, and as low as 47–50% in oil palm gardens¹⁷.

Reports on light penetration in palm canopies support our results. Oil palm gardens show a high similarity index (0.72) of weed flora (Table 3) with fallow land. As the oil palm canopy is able to use only 47-50% of light¹⁷ the weeds are able to grow luxuriantly as in fallow land utilizing excess light. Weed flora in adult arecanut gardens was dissimilar (0.29) to that in fallow land as well as in oil palm gardens. This could be due to the low light availability in adult arecanut gardens¹⁶. The similarity index of weed flora in adult and juvenile arecanut regions was 0.42. This further supports the difference in light availability¹⁶ in juvenile and adult arecanut palms which is as high as 50%.

Weed species distributed in different taxonomic groups are given in Table 4. As many as 17 families of dicots, three families of monocots and four members of pteridophytes were noticed during the study. Members of the families Droseraceae and Polygonaceae were present only in fallow land, while members of Araceae and Caryophyllaceae were found only in adult arecanut gardens. Members of Poaceae, Asteraceae, Oxalidaceae and Urticaceae were found in all the plots studied. Pteridophytes were present in fallow land, coconut and oil palm gardens. Dicots were dominant in all the plots studied. Among monocots, only Poaceae, Cyperaceae and Araceae were noticed in the study. Dicots dominated over monocots in all the study areas. High annual rainfall, fertile loamy soil of the

Table 3. Similarity index of weed species in palm gardens

Fallow land	Juvenile arecanut	Adult arecanut	Coconut	Oil palm
Fallow land	0.58	0.29	0.38	0.72
Juvenile arecanut		0.42	0.59	0.67
Adult arecanut			0.44	0.29
Coconut Oil palm				0.56

 Table 4. Weed species of different botanical families in palm gardens under study

		2			
Family	Fallow land	Juvenile arecanut	Adult arecanut	Coconut	Oil palm
Monocotyledons	5	6	5	1	5
Araceae (Ar)	0	0	1	0	0
Cyperaceae (C)	2	1	2	0	1
Poaceae (P)	3	5	1	1	4
Dicotyledons	19	14	10	12	15
Acanthaceae (Ac)	0	0	0	1	1
Apiaceae (Ap)	1	1	0	1	1
Asteraceae (A)	3	2	3	2	4
Caryophyllaceae (Ca)	0	0	2	0	0
Droseraceae (D)	1	0	0	0	0
Euphorbiaceae (E)	1	1	0	0	0
Fabaceae (F)	2	1	0	1	1
Labiatae (L)	2	1	0	0	1
Melastomaceae (M)	1	1	0	1	1
Oxalidaceae (O)	1	1	1	1	1
Polygonaceae (Po)	1	0	0	0	0
Rubiaceae (R)	2	1	0	1	2
Scrophulariaceae (S)	2	3	1	1	0
Solanaceae (So)	0	1	1	1	0
Tiliaceae (T)	1	0	0	0	1
Urticaceae (U)	0	1	1	1	1
Verbenaceae (V)	1	0	0	1	1
Pteridophytes (Pter)	4	0	0	1	2
Total	28	20	13	14	22

Teesta valley, and the wide range of summer and winter temperatures favoured luxuriant dicot vegetation in the area. Monocots were found to be few in number in cropped area. The density of the monocots was almost universally low in the palm gardens than in fallow land, which indicates that the adaptability of monocots is more in fallow land than in cropped land.

Gopinathan Nair and Chami⁴ found dicotyledons to be dominant in coconut gardens. Cyperaceae, Poaceae and Commmelinaceae are the major monocot families, and Asteraceae, Fabaceae and Rubiaceae are the predominant dicot families seen in coconut gardens. They also concluded that the weeds of the families Cyperaceae and Poaceae are the most troublesome. Souza *et al.*¹⁰ found the plants of Poaceae, Euphorbiaceae, Fabaceae, Cyperaceae and Verbenaceae to be predominant weeds in Cupuacu and peach palm gardens.

The present investigation reveals that the different weed species can grow in different shade conditions and that their growth depends on the availability of sunlight along with other growth conditions. Prevalence of dicot weed species was higher in all conditions under study than that of the monocots. Restriction of some weed species to particular areas supports the fact that they require special conditions for growth, whereas the presence of some weeds in all the study areas shows that they can grow under varied light conditions.

- Derksen, A. D., Andersen, L. R., Blackshaw, E. R. and Maxwell, B., Weed dynamics and management strategies for cropping systems in the Northern Great Plains. *Agron. J.*, 2002, 3, 174–185.
- Fuente, E. B., Saurez, S. A., Ghersa, C. M. and Leon, R. J. C., Soyabean weed communities: Relationship with cultural history and crop yield. *Agron. J.*, 1999, **91**, 234–241.
- Moyer, J. R., Owenya, Z. J. and Kibuwa, S. P., Weed population and agronomic practices at wheat farms on the Hanang plains of Tanzania. *Trop. Pest Manage.*, 1989, 35, 26–29.

- Gopinathan Nair, R. and Chami, P., A survey of weeds in the fields of coconut research station, Kasaragod. *Indian Coconut J.*, 1963, XVII, 40–47.
- Thomas, C. G. and Abraham, C. T., Weeds of coconut gardens in the central zone of Kerala. *Indian Coconut J.*, 1996, 26, 8–10.
- 6. El-Halawany, E. F., Flora and vegetation of date palm orchards in the north eastern part of the Nile delta, Egypt. *Egypt. J. Bot.*, 2001, **41**, 41–64.
- Abd-El-Ghani, M. M., Weed community of date palm orchards in the Feiran Oasis (South Sinai, Egypt). *Fragm. Florist. Geobot.* (*Krakow*), 1998, 43, 257–271.
- Sahapurmath, G. B., Srivanna, H. and Girisha, H. V., Studies on weed intensity in existing areca based agro forestry models. *Karnataka J. Agric. Sci.*, 2003, 16, 260–264.
- Bogantes, A. and Aguero, R., Weed dynamics and control in peach palm (*Bactris gasileps* K.) for palm hearts. *Agron. Mesoamericana*, 2003, **149**, 41–49.
- Souza, L. S. A., Silva, J. F. and Souza, M. D. B., Floristic composition of weeds in agro-systems of Cupuacu (*Theobroma grandiflorum*) and peach palm (*Bactris gasipaes*). *Planta-Daninha*, 2003, 21, 249–255.
- 11. Sharma, P. D., *Ecology and Environment*, Rastogi Publication, Meerut, 1998.
- 12. Shannon, C. E. and Weaver, W., *The Mathematical Theory of Communication*, Univ. of Illinois Press, Urbana, 1949.
- 13. Odum, E. P., *Fundamentals of Ecology*, W.B. Saunders Company, Philadelphia, 1971, p. 574.
- 14. Thomas, D., Pastures and livestock under tree crops in humid tropics. *Trop. Agric.*, 1978, **55**, 39–44.
- 15. Muralidharan, A., Biomass productivity, plant interactions and economics of intercropping in arecanut garden. PhD thesis, University of Agriculture Science, Bangalore, 1980, p. 271.
- Bavappa, K. V. A., Coconut based cropping system. Indian Coconut J., 1995, 25, 6–8.
- Wilson, J. R. and Ludlow, M. M., The environment and potential growth of herbage under plantations. In Proceedings of a Workshop on Forages for Plantation Crops, Bali, Indonesia, 27–29 June 1990, pp. 10–24.

Received 5 August 2005; revised accepted 3 December 2006