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### Multiplex PCR for Differentiation of Mycoplasma mycoides Cluster from other Mycoplasma

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## Multiplex PCR for Differentiation of *Mycoplasma mycoides* Cluster from other Mycoplasma

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### Abstract

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*A multiplex PCR was standardized for detection of various members of Mycoplasma mycoides cluster and their differentiation from other mycoplasma as well as some other bacteria. Using mycoplasma group specific primers GPO-1/MGSO and Mycoplasma mycoides cluster specific primers MC 323 / MC 358 simultaneously, two amplified products of 1.5 kbp and 715 bp were found only in members of M. mycoides cluster whereas, a single band of 715 bp was detected in all other non cluster mycoplasma and acholeplasma while no amplified product was detected in any other bacterial strain tested.*

### Introduction

A number of important diseases viz. contagious bovine pleuropneumonia (CBPP), contagious caprine pleuropneumonia (CCPP), arthritis, mastitis, genital disorders of livestock are

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produced by members of *Mycoplasma mycoides* cluster (Cottew *et al.*, 1987; Thiaucourt and Bolske, 1996). Amongst them, the most important disease is CBPP, a disease of OIE list A and is also included in six priority diseases under FAO emergency prevention scheme (Rweyemamu and Benkirane, 1996). The accurate detection of the causative agent of these infections has not been possible with the help of routine conventional techniques like isolation and serodiagnosis. Isolation is laborious and often hampered by contamination with other bacteria while extreme serological cross-reactivity among different species of mycoplasma may obfuscate the serodiagnosis (Cottew *et al.*, 1987). Moreover, serological tests do not detect the positive cases in early and later stages of infection. Lately, the use of polymerase chain reaction has been found helpful in detection of *Mycoplasma mycoides* cluster as it has both specificity and sensitivity (Bashiruddin *et al.* 1994; Kumar *et al.*, 2001). Recently, the introduction of multiplex PCR has proved to be very useful in detection and differentiation of different organisms in a single PCR reaction, which is saving both time and money (Caron *et al.*, 2000).

The present study was intended to standardize a multiplex PCR assay for detection of mycoplasma as well as members of *Mycoplasma mycoides* cluster in a single PCR reaction.

### **Materials and Methods**

Ten strains of mycoplasma (*M.bovis*, *M.bovirhinis*, *M: bovigentalium*, *M.mycoides* subsp. *mycoides* SC, *M. sp. bovine* group 7, *M. alkalescens*, *M. agalactiae*) and one strain each of *Acholeplasma laidlawii*, *Pasteurella* sp. and *Salmonella* sp. obtained from National Referral Laboratory on Mycoplasma, Division of Bacteriology and Mycology, IVRI, Izatnagar were used in this study. The strains of *M.mycoides* subsp. *mycoides* SC, *M. sp. bovine* group 7 and *M.agalactiae* were grown in MBHS-L liquid medium (Carmichael *et al.*, 1972) whereas, specially enriched B medium (Ernø and Stipkovits, 1973) was used for the growth of *M.bovis*, *M.bovirhinis*, *M.bovigentalium* and *M. alkalescens* strains.

### *Isolation of genomic DNA*

The genomic DNA of all the strains were isolated as per the protocol of Wilson (1987) and the concentration of genomic DNA was determined by measuring the optical density at 260 nm in a UV spectrophotometer. Finally, these were diluted in sterile distilled water to make the concentration of 10 ng/ $\mu$ l of DNA.

### *Multiplex PCR assay*

Mycoplasma group specific primers GPO-1 [5'-ACT CCT ACG GGA GGC AGC AGT A-3'] and MGSO [5'-TGC ACC ATC TGT CAC TCT GTT AAC CTC -3'] (Kuppeveld *et al.*, 1992) as well as *Mycoplasma mycoides* cluster specific primers viz. MC 323 [5'-TAG AGG TAC TTT AGA TAC TCA AGG -3'] and MC 358 [5'-GAT ATC TAA AGG TGA TGG T - 3'] (Bashiruddin *et al.*, 1994) were used simultaneously. A 20 ng of genomic DNA of each strain was used separately as a template in 50  $\mu$ l reaction mixture consisting of 10mM Tris-HCl, pH 8.8; 50 mM KCl, 1.5 mM MgCl<sub>2</sub>; 0.08% nonidet P40; 200  $\mu$ M dNTP mixture (MBI Fermentas). The programme consisted of initial denaturation at 94C for 5 min followed by 30 cycles of denaturation at 94C for 45 sec, annealing at variable temperatures (50, 53, 55 and 57C) for 1 min and extension of 1 min 20 sec at 72C. The final extension was carried out at 72C for 5 min.

A 10  $\mu$ l of each PCR product was analysed on 1.4% agarose gel electrophoresis at 7 volts/cm for 1 h 30 min in 0.5x TBE buffer (45 mM Tris, 45 mM boric acid and 2 mM EDTA, pH 8.0) alongwith the DNA maker (pBR 322 DNA digested with AluI enzyme). The gel was stained with ethidium bromide and finally, it was visualized and photographed under UV gel documentation system.

### *Results and Discussion*

Out of the four annealing temperatures, only 53C was found optimum in the present multiplex PCR assay as evidenced by the presence of intense bands of both the amplified products. On using both sets of primers GPO-1/MGSO and MC 323/MC 358, a single band of 715 bp was found in all the non-cluster mycoplasma and acholeplasma strains whereas, two amplified products of 1.5 kbp

and 715 bp were detected only in the strains of *Mycoplasma mycoides* cluster viz. *M. mycoides* subsp. *mycoides* SC (PG1 and NCVP-1/86) and *M. sp. bovine* group 7 (PG50). No amplified product was detected in other bacteria viz. *Pasteurella* sp. and *Salmonella* sp. (Fig.1). Similar types of findings were also reported by Kuppeveld *et al.* (1992) using GPO-1/MGSO primers, which amplify a 715 bp region of 16S rRNA gene of mycoplasma, acholeplasma, spiroplasma and ureaplasma. While Bashiruddin *et al.* (1994) observed 1.5 kbp product in the region of CAP-21 sequence of all the members of *Mycoplasma mycoides* cluster group (*M. mycoides* subsp. *mycoides*

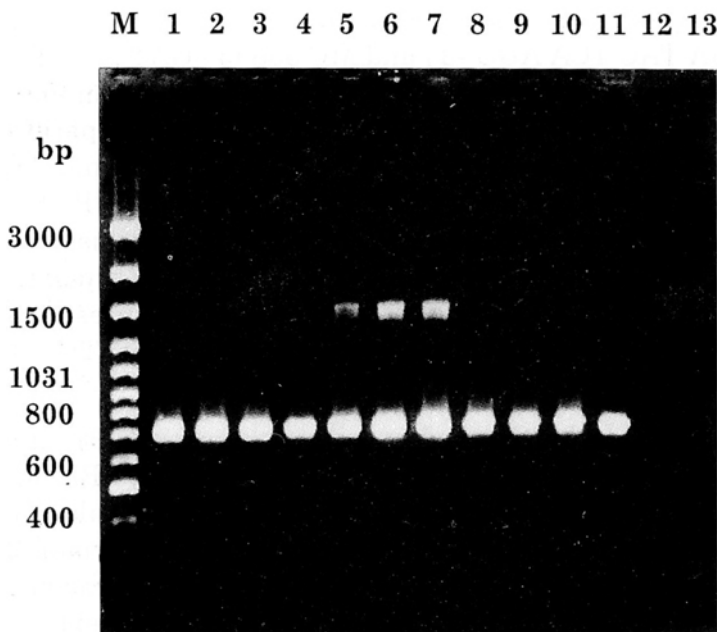


Fig. 1 : Multiplex PCR using primers GPO-1, MGSO, MC323 and MC358: Lane M: Marker (pBR322DNA digested with AluI), Lane1: *M. bovis* (NC317), Lane2: *M. bovis* (23K), Lane3: *M. bovirhinis* (NC448), Lane4: *M. bovigenitalium* (NC58), Lane 5: *M. mycoides* subsp. *mycoides* SC (PG1), Lane 6: *M. mycoides* subsp. *mycoides* SC (NCVP-1/86), Lane7: *M. sp. bovine* group7 (PG50), Lane8: *M. alkalescens* (NC242), Lane9: *M. agalactiae* (RPNS216), Lane 10: *M. agalactiae* (RPNS 200), Lane 11 : *A. laidlawii* (NC 313), Lane 12 : *Pasteurella* sp., Lane 13 : *Salmonella* sp.

type SC, *M. mycoides* subsp. *mycoides* type LC, *M. mycoides* subsp. *capri*, *M. capricolum* subsp. *capricolum*, *M. capricolum* subsp. *capripneumoniae* and *M. sp.* bovine group 7) with MC 323 / MC358 primers.

The results of present study revealed that the multiplex PCR can be used for the detection of members of *Mycoplasma mycoides* cluster and it also differentiates them from other non cluster mycoplasma and bacteria simultaneously in a single PCR assay. This ultimately saved the time and money for detection of mycoplasma upto the species level in a single PCR reaction.

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संजय दास, वी.पी. सिंहा अन्य माइकोप्लाज्मा से माइकोप्लाज्मा माइकायडिस समूह का बहुघटकी पीसीआर द्वारा विभेदीकरण

अन्य माइकोप्लाज्मा और सूक्ष्माणुओं से माइकोप्लाज्मा माइकायडिस का पता लगाकर विभेदीकरण के लिए एक बहुघटकी पीसीआर का मानकीकरण किया गया। माइकोप्लाज्मा वर्ग विशिष्ट प्राइमर, जीपीओ-1/एमजीएसओ और माइकोप्लाज्मा माइकायडिस वर्ग विशिष्ट प्राइमर एमसी 323/एमसी 358 को साथ-साथ प्रयोग करके 1.5 केवीपी और 715 बीपी के दो दीर्घकृत उत्पाद केवल माइकोप्लाज्मा माइकायडिस वर्ग से मिले, जबकि अन्य असमूही माइकोप्लाज्मा और एकोलिप्लाज्मा में केवल 715 बीपी की एक पट्टी और अन्य परीक्षित सूक्ष्माणुओं में कुछ भी नहीं मिला।