RESEARCH ARTICLE

Cytotaxonomic Studies in Four Species of Genus *Puntius* (Hamilton, 1822) from Central India

Rabindra Kumar Saroniya · Naresh Sahebrao Nagpure · Devendra Nath Saksena · Basdeo Kushwaha · Ravindra Kumar

Received: 1 September 2012/Revised: 14 February 2013/Accepted: 1 April 2013/Published online: 30 July 2013 © The National Academy of Sciences, India 2013

Abstract The cytotaxonomic analyses of four species of genus Puntius, viz. P. chola, P. conchonius, P. sophore and P. ticto from central India was carried out for the first time. The metaphase chromosomes were prepared from kidney and gill tissues. The diploid chromosome number and karyotype formula (KF) were found to be 50 & 2m + 4sm + 2st + 42t in P. chola; 50 & 14m + 28sm + 8st in P. conchonius; 48 & 4m + 2st + 42t in P. sophore and 50 & 14m + 24sm +8st + 4t in P. ticto. Based on KF, the fundamental arm number were determined as 56, 92, 52 and 88, respectively, for these species. The karyomorphological features indicated that P. conchonius and P. ticto are closely related and the same holds true for P. chola and P. sophore. The chromosomes of all the four species exhibited constitutive heterochromatic blocks at their centromeric position, as detected by Cbanding technique. Variations in number of NORs were observed with presence of single pair of NORs in P. chola and P. conchonius,

R. K. Saroniya

N. S. Nagpure (⊠) · B. Kushwaha · R. Kumar Molecular Biology and Biotechnology Division, National Bureau of Fish Genetic Resources, Canal Ring Road, P.O. Dilkusha, Lucknow, UP 226 002, India e-mail: nsnagpure@gmail.com; nagpurens@yahoo.co.in

B. Kushwaha e-mail: basdeo.scientist@gmail.com

R. Kumar e-mail: ravindra.scientist@gmail.com

D. N. Saksena School of Studies in Zoology, Jiwaji University, Gwalior, MP 474 011, India e-mail: dnsaksena@gmail.com whereas in *P. sophore* and *P. ticto* multiple NORs were observed. Thus, based on the karyological features it can be hypothesized that *P. conchonius* and *P. ticto* may be in advanced stage of karyo-evolution.

Keywords Cytogenetics · Fish · Karyomorphology · *Puntius*

Introduction

The cyprinid fish genus *Puntius* comprises of more than 60 species found in India and new species are being discovered, especially from the North–Eastern and Southern parts of India [1, 2]. Many species of this genus are considered as weed fishes, while some of them are of ornamental value. Taxonomic ambiguities exist between many closely related *Puntius* species [2–4]. Further, these fish inhabits and breeds in common water-bodies; therefore, chances of inter-breeding and hybridization are higher. Cytotaxonomic studies are, therefore, required to document inter- as well as intra-specific variations and to resolve taxonomic ambiguities among the species.

The study on fish chromosome has received considerable attention because of their importance in classification, evolution and heredity [5, 6]. The cytogenetic techniques are considered as authentic tools for species characterization and have extensively been used to resolve taxonomic ambiguities in closely related species, identification of strains/cytotypes, genetic polymorphisms, sex determination, polyploidy etc. [7–9]. Comparison of chromosome number and structure among different species reveals phylogenetic relationship and throws light on their karyoevolution and can also be helpful in planning conservation strategies for threatened fish species [10].

Veerbhumi Govt. P.G.College, Mahoba, UP 210 427, India e-mail: rks_rk10@yahoo.com

Four Puntius species, viz. P. chola (known as 'swamp barb'), P. conchonius (known as 'rosy barb'), P. sophore (known as 'pool barb') and P. ticto (known as 'two spot barb') collected from central India, were investigated to find out karyotypic variations and cytotaxonomic relationship among them. The nucleolar organizer regions (NORs) staining is considered to be one of the most commonly used technique for cytogenetic characterization of fish. The silver nitrate (AgNO₃) stained NORs detects only transcriptionally active sites [11], while GC specific fluorochrome chromomycin A₃ (CMA₃) stains both active and inactive NORs probably due to their high GC content [12]. C-banding is another useful technique used to study the localization of constitutive heterochromatic (CH) bands and its staining with fluorescent dye may further increase the resolution of bands.

As far as authors are aware, there is no information available on the CH bands and NORs in these four species of genus *Puntius*. Therefore, the present study was aimed to analyse the karyotypic characteristics, with particular reference to the variation in CH and NORs, and establish cytotaxonomic relationship among these species.

Materials and Methods

Live fish specimens of P. chola (n = 6), P. conchonius (n = 10) and P. ticto (n = 5) were collected from Pahuj river, Jhansi, Uttar Pradesh, while specimens of P. sophore (n = 10) were obtained from Ramsagar reservoir at Barauni, Datia, Madhya Pradesh with the help of local fishermen. Sharp pointed needle, like divider, and stainless steel ruler were used for recording body measurements [13]. Specimens were identified up to species level following taxonomic keys described by Jayaram [14], Talwar and Jhingran [15] and Srivastava [16]. The average total length and wet weight of P. chola specimens were 8.25 cm (range 7.3-9.0) and 6.35 g (range 5.4-7.2). In P. conconius, P. sophore and P. ticto, the total length and wet weight of specimens were 7.57 cm (range 6.3–10.0) & 6.16 g (range 4.4-13); 8.04 cm (range 6.9-9.0) & 6.58 g (range 5.0-10.2), and 6.45 cm (range 5.9-6.8) & 3.20 g (range 2.0-4.2), respectively. The specimens were at juvenile stage and the sex was unidentifiable by visual examination.

The chromosomes were obtained from kidney and gill cells following hypotonic (KCl) treatment, fixation (methanol-acetic acid) and air drying technique as described by Bertollo et al. [17] and the dried slides were stained with Giemsa. For karyotyping, the chromosomes were classified as per the method described by Levan et al. [18]. A total of 50 slides were prepared from each species and 6 good spreads from each slide were used for karyo-morphological analyses. Chromosome lengths were measured using 'MicroMeasure' (version 3.2) [19] computer software. C-banding was carried out according to Sumner [20], but the slides were stained with fluorescent propidium iodide according to Fontana et al. [21]. The method of Howell and Black [11] was used for silver staining of NORs, while fluorescent CMA₃ staining of NORs was done according to Sola et al. [22]. All the photographs were taken at $100 \times$ magnification using fluorescent microscope and a total of 80 random spreads from each species were considered for determining a particular banding/staining pattern.

Results and Discussion

Metaphase Spreads and Karyotype

The metaphase complements and karyotype of P. chola, P. conchonius, P. sophore and P. ticto are shown in Fig. 1 and the chromosomal morphometric data are described in Table 1. In P. chola, P. conchonius, and P. ticto, the diploid chromosome number (2n) was found to be 50, whereas in P. sophore the 2n was recorded as 48. Further, variations in karyo-morphology were observed among the species. The specimens of P. chola possessed one metacentric pair (m), two submetacentric (sm) pairs, one subtelocentric (st) pair, and 21 telocentric (t) pairs of chromosomes and the karyotype formula (KF) was derived as 2m + 4sm + 2st + 42twith fundamental arm number (FN) of 56. In P. conchonius, the KF was derived as 14m + 28sm + 8st and FN as 92. In P. sophore, the karyotype composed of two pairs of metacentric, one pair of subtelocentric and 21 pairs of telocentric chromosomes with FN as 52. The karyotype of P. ticto specimens composed of seven pairs of metacentric, 12 pairs of submetacentric, four pairs of subtelocentric and two pairs of telocentric chromosomes with FN as 88.

Chromosome Morphometry and Index

The total length of the chromosomes (TL) in *P. chola* varied from 0.97 to 2.73 μ m, whereas the centromere index (CI) and relative length (RL) ranged from 14.46 to 47.15 μ m and from 2.55 to 7.17 %, respectively. In *P. conchonius* specimens, the TL varied from 1.63 to 2.97 μ m, and the CI and RL from 17.08 to 44.61 μ m and from 3.0 to 5.46 %, respectively. In *P. sophore*, the TL ranged from 0.84 to 2.68 μ m, CI from 13.73 to 48.88 μ m and RL from 2.15 to 6.87 %. In *P. ticto*, the TL varied from 1.55 to 3.64 μ m and CI from 8.98 to 48.04 μ m and RL from 2.70 to 6.35 %.

NOR Staining

The chromosome complements showing AgNO₃ and CMA₃ stained NORs in the *Puntius* species are presented

Fig. 1 Metaphase spread and

karyotype of: P. chola (a and a1), P. conchonius (b and b1),

P. sophore (c and c1) and



in Fig. 2. In P. chola and P. conchonius, single NOR pair was observed at the end of short arms of subtelocentric chromosome using both AgNO₃ and CMA₃ staining. In P. sophore specimens, however, NOR signals were found terminally on three pairs of telocentric chromosome. Similarly in P. ticto, NOR signals were detected at terminal/sub-terminal positions of two pairs of metacentric and one pair of submetacentric chromosomes.

Constitutive Heterochromatin

The C-banding technique revealed the localization of constitutive heterochromatic blocks and the C-banded metaphase complements are presented in Fig. 3. The CH blocks were detected on the chromosomes of all the species and were centromeric in position. No significant variation in the position and size of the bands were observed among these species.

Table 1 Chromosome morphometric data and chromosome types of Puntius sp

CPN	P. chola					P. conchonius				P. sophore				P. ticto						
	AR	TL	CI	RL (%)	СТ	AR	TL	CI	RL (%)	СТ	AR	TL	CI	RL (%)	СТ	AR	TL	CI	RL (%)	СТ
1	1.12	1.93	47.15	5.07	m	1.56	2.97	39.06	5.46	m	1.05	2.68	48.88	6.87	m	1.41	3.64	41.48	6.35	m
2	2.25	2.73	30.77	7.17	sm	1.43	2.73	41.02	5.02	m	1.2	1.87	45.45	4.79	m	1.52	2.90	39.65	5.06	m
3	2.92	2.08	25.48	5.46	sm	1.46	2.42	40.49	4.45	m	6.28	2.33	13.73	5.97	st	1.08	2.81	48.04	4.90	m
4	5.91	1.66	14.46	4.36	st	1.65	2.34	37.61	4.31	m	0	2.34	0	5.99	t	1.19	2.46	45.53	4.29	m
5	0	2.33	0	6.12	t	1.33	2.15	42.79	3.96	m	0	2.25	0	5.76	t	1.20	2.42	45.45	4.22	m
6	0	2.25	0	5.91	t	1.24	1.95	44.61	3.58	m	0	2.14	0	5.48	t	1.67	2.27	37.44	3.96	m
7	0	1.76	0	4.62	t	1.40	1.95	41.54	3.59	m	0	1.96	0	5.02	t	1.18	1.64	45.73	2.86	m
8	0	1.74	0	4.57	t	2.52	2.43	28.39	4.47	sm	0	1.93	0	4.95	t	1.96	2.61	33.72	4.55	sm
9	0	1.48	0	3.88	t	2.73	2.43	26.75	4.47	sm	0	1.61	0	4.13	t	2.17	2.60	31.54	4.53	sm
10	0	1.44	0	3.78	t	2.70	2.41	26.97	4.43	sm	0	1.61	0	4.13	t	2.60	2.52	27.78	4.39	sm
11	0	1.44	0	3.78	t	2.02	2.33	33.05	4.29	sm	0	1.59	0	4.07	t	2.64	2.51	27.49	4.37	sm
12	0	1.43	0	3.75	t	2.43	2.3	29.13	4.23	sm	0	1.56	0	3.99	t	2.57	2.50	28.00	4.36	sm
13	0	1.35	0	3.55	t	2.52	2.29	28.38	4.21	sm	0	1.52	0	3.89	t	1.90	2.44	34.43	4.25	sm
14	0	1.35	0	3.55	t	2.46	2.22	28.83	4.09	sm	0	1.52	0	3.89	t	1.84	2.33	35.19	4.06	sm
15	0	1.34	0	3.52	t	2.46	2.15	28.84	3.96	sm	0	1.41	0	3.61	t	2.04	2.16	32.87	3.77	sm
16	0	1.32	0	3.46	t	2.31	2.12	30.19	3.90	sm	0	1.38	0	3.54	t	2.17	2.03	31.53	3.54	sm
17	0	1.32	0	3.46	t	2.14	1.95	31.79	3.59	sm	0	1.32	0	3.38	t	2.09	1.98	32.32	3.45	sm
18	0	1.31	0	3.44	t	2.78	1.93	26.42	3.55	sm	0	1.32	0	3.38	t	2.43	1.75	29.14	3.05	sm
19	0	1.29	0	3.39	t	2.49	1.85	28.65	3.41	sm	0	1.31	0	3.36	t	1.92	1.55	34.19	2.70	sm
20	0	1.27	0	3.33	t	2.06	1.81	32.59	3.33	sm	0	1.30	0	3.33	t	5.82	2.32	14.65	4.05	st
21	0	1.19	0	3.13	t	1.96	1.63	33.74	3.00	sm	0	1.29	0	3.30	t	6.06	2.19	14.15	3.82	st
22	0	1.05	0	2.76	t	4.63	2.03	17.73	3.74	st	0	0.98	0	2.51	t	6.34	2.13	13.62	3.72	st
23	0	1.03	0	2.71	t	4.15	2.01	19.40	3.69	st	0	0.96	0	2.46	t	4.24	1.99	19.09	3.47	st
24	0	1.01	0	2.65	t	4.85	1.99	17.08	3.66	st	0	0.84	0	2.15	t	9.55	1.90	9.47	3.31	t
25	0	0.97	0	2.55	t	4.54	1.94	18.04	3.57	st	-	-	-	-	-	10.13	1.67	8.98	2.91	t

CPN Chromosome pair number, AR Arm ratio, TL Total length of chromosome, CI Centromeric index, RL (%) Relative length in percent; CT Chromosome type

The most commonly occurring 2n in fish family cyprinidae is 50 with the range from 34 to 446 [23, 24]. According to the studies performed by various workers on *Puntius* species in India, the 2n = 50 seemed to be the modal number for the genus [25], with the range from 48 to 52 (www.fishbase.org/ version 06/2012). In the present study, the 2n = 50 was found in *P. chola*, *P. conchonius* and *P. ticto*, whereas in *P. sophore* the same was found to be 48. The comparison of results of the present study with earlier reports has been presented in Table 2. The karyomorphology of all the species showed variation from the earlier reports, except of *P. sophore* in which the KF (4m + 2st + 42t) and FN (52) was in confirmation with the finding of Rishi and Rishi [26].

The presence of different populations, races and/or sub species arising from mutation, race improvement and hybridization with other indigenous species could be the possible explanation for differences in number and type of chromosomes reported in a species that is distributed in different aquatic ecosystems [27, 28]. Intra-specific variation in karyo-morphology have also been ascribed to ambiguities in classification due to border-line centromere positions caused by cell to cell variation in the extent of chromosome contraction, which is a general problem in the description of the relatively small chromosomes of cyprinids [29-31]. Differences in FN among closely related species corroborated the importance of pericentric inversions as the main mechanism of karyotypic evolution in several modern fish orders [32-36]. Different FN reported in Labeo rohita from China (i.e. 76), Thailand (80) and India (70) reflected local differentiation in the karyotype [37, 38]. The chromosomal morphometric data revealed that *P. ticto* possessed the longest chromosome $(3.64 \ \mu m)$, while the smallest (0.84 µm) was observed in P. sophore. The maximum numbers of metacentric chromosomes (14) were found in P. conchonius and P. ticto, whereas the maximum numbers of telocentric chromosomes (42) were found in P. chola and P. sophore. No telocentric and



Fig. 2 Metaphase plates and chromosome pair(s) showing NOR regions detected by AgNO₃ and CMA₃, respectively, in: *P. chola* (a1, a2), *P. conchonius* (b1, b2), *P. sophore* (c1, c2) and *P. ticto* (d1, d2). *Bar* = 5 μ m



Fig. 3 Metaphase spreads showing constitutive heterochromatin regions, as detected by C-banding, in: *P. chola* (a), *P. conchonius* (b), *P. sophore* (c) and *P. ticto* (d). $Bar = 5 \mu m$

submetacentric chromosomes were observed in *P. conchonius* and *P. sophore*, respectively. The karyo-morphological features indicated *P. conchonius* and *P. ticto* to be closely related and the same holds true for *P. chola* and *P. sophore*. Similar closeness of *P. chola* with *P. sophore* and *P. conchonius* with *P. ticto* were also recorded on the basis of mitochondrial cytochrome b gene [39]. A phylogenetic study based on restriction fragment polymorphism analysis among different species of genus *Puntius* also indicated closeness of *P. chola* with *P. sophore* [4].

The banding studies can help in the precise mapping of genes onto the chromosomes and the evolutionary relationship between species can be inferred at a gross level by comparing banding patterns. The C-banding is very often species-specific and its distribution may vary considerably from species to species [40]. The characteristics of the C-bands, which aids to the identification of species, are their size, location on the chromosome and the position of the C-banded chromosome in the karyotype [41]. In the present study; however, the CH bands were found at the centromeric position of the chromosomes in all the species and no variation were observed with regards to the position and size/intensity of bands. Similar distribution pattern of heterochromatin throughout the chromosomes have been reported by several workers in many closely related species belonging to the same genus: *Notropis lutrensis* and *N. venustus* [42], *Schizodon borelli* and *S. isognathum* [43] and *Vimba vimba* and *V. elongata* [44]. Nuclear satellite

Species	2n	KF	FN	Region	References		
P. chola	50	2m + 2sm + 46t	54	Arunachal Pradesh	Sahoo et al. [55]		
	50	2m + 4sm + 2st + 42t	56	Uttar Pradesh	Present study		
P. conchonius	50	22m + 16sm + 12t	88	Kashmir	Ganai and Yousuf [25]		
	50	16m + 24sm + 2st + 8t	90	Orissa	Khuda Bukhsh et al. [56].		
	48	10m + 20sm + 10st + 8t	78	Jammu & Kashmir	Sharma and Agrawal [57]		
	50	14m + 28sm + 8st	92	Uttar Pradesh	Present study		
P. sophore	48	2m + 46t	50	Orissa	Biswal et al. [58].		
	50	2m + 4sm + 44t	56	Tamil Nadu	Khuda Bukhsh et al. [52].		
	48	4m + 2st + 42t	52	Haryana	Rishi and Rishi [26]		
	48	4m + 4st + 40t	52	West Bengal	Manna and Prasad [59]		
	48	4m + 2st + 42t	52	Madhya Pradesh	Present study		
P. ticto	50	28m + 16sm + 6t	94	Arunachal Pradesh	Sahoo et al. [55].		
	50	14m + 22sm + 6st + 8t	86	West Bengal	Manna and Prasad [60]		
	50	14m + 24sm + 8st + 4t	88	Uttar Pradesh	Present study		

Table 2 Comparison of chromosomal morphology of Puntius species reported by different workers

DNA have one property in common, namely heterochromatinization, despite of their species specificity, and the apparent species specificity may be the result of natural selection for duplicated short polynucleotide segments. The centromeric heterochromatin is believed to confer protection and strength to the centromeric chromatin [45]. This condition may arise due to the Robertsonian fusions [46] or could have been formed by tandem duplication/pericentric inversion of heterochromatic DNA [47]. It is opined that karyotypic stability might reached after canalization to an optimal karyotypic configuration [48], which could be a reason for similarity in distribution of heterochromatin in these *Puntius* species.

The NORs are the chromosomal sites of genes that were presumably transcribed at preceding interphase and are important in view of their intimate relationship with protein synthesis [11, 49]. An important characteristic of NORs in fish is related to its inter- and/or intra-species polymorphism. NOR characteristics can be utilized as a marker for cytotaxonomic studies and can even aid in constructing phylogenetic hypotheses (cyto-systematics) for several fish groups [8]. In fish, presence of NORs on single pair of chromosome was considered to be plesiomorphic or primitive condition [50]. Single pair of NOR was observed at the end of the short arms of subtelocentric chromosomes (4th pair) in P. chola and at the end of the short arms of subtelocentric chromosome in P. conchonius with both CMA₃ and AgNO₃ staining, whereas multiple NORs were found in P. sophore and P. ticto that could be speciesspecific character. The information on size, position and number of NORs are suitable for tracing intra- and interspecific differences and may serve to demarcate and derive the taxonomic status of species in terms of karyo-evolution [49, 51–53].

Comparative phylogenetic analyses have been employed to examine the evolutionary history of fish chromosome. The most parsimonious ancestral state for major actinopterygiian clades has been observed as 48 chromosomes [54]. Moreover, the presence of more number of telocentric chromosomes is also an ancestral condition. Based on the karyo-morphological features, P. chola and P. sophore may be considered as primitive species in the present study. On the other hand, P. conchonius and P. ticto may be considered as derived species or may be in the advanced stages of karyo-evolution due to presence of many numbers of bi-armed chromosomes. In the present study, P. chola satisfies the condition of being primitive species due to presence of single NOR. Surprisingly, P. sophore does not follow the condition of primitive species due to presence of multiple NORs, as proposed by Gold and Amemiya [50]. The species belonging to the genera Puntius have similarity in external phenotypic characters that lead to taxonomic uncertainty. Moreover, P. chola and P. sophore looks alike with similar morphometric and meristic characters, except the presence of one pair of maxillary barbells and an extra band on dorsal fin in P. chola. Similarly, P. conchonius greatly resembles with P. ticto based on morphology, except the presence of one black spot at anterior body in the later [14–16]. Further studies using molecular tools may add to reaffirm the cytotaxonomic and phylogenetic relationships in these species.

Acknowledgments The first author is highly thankful to UGC, New Delhi for providing financial help to the study in the form of minor research project. The authors are thankful to the Head, Department of Zoology, Jiwaji University, Gwalior (MP) and to the Director, National Bureau of Fish Genetic Resources, Lucknow (UP) to provide all necessary facilities and help to conduct this study.

References

- Devi KR, Indra TJ, Knight JDM (2010) *Puntius rohani* (Teleostei: Cyprinidae), a new species of barb in the *Puntius filamentosus* group from the southern Western Ghats of India. J Threat Taxa 2(9):1121–1129
- Kullander SO, Fang F (2005) Two new species *Puntius* from Northern Myanmar (Teleostei: Cyprinidae). Copeia 2:290–302
- Nagpure NS, Kushwaha B, Srivastava SK, Kumar Ravindra, Gopalakrishnan A, Baseer VS, Verma MS (2003) Characterization of three endemic fish species from Western Ghats using cytogenetic markers. Nucleus 46(3):110–114
- Balaraj S, Basheer M (2012) Genetic diversity among *Puntius* sophore complex using restriction fragment length polymorphism. J Med Allied Sci 2(2):49–53
- Gold JR, Li YC, Shipley NS, Powers PK (1990) Improved methods for working with fish chromosomes with a review of metaphase chromosome banding. J Fish Biol 37(4):563–575
- Barat A, Sahoo PK, Ponniah AG (2002) Karyotype and nucleolar organizer regions (NORs) in a few hill stream fishes. In: Ayyappan S, Jena JK, Joseph MM (eds) The fifth Indian fisheries forum proceedings, AFSIB, Mangalore and AoA, Bhubaneswar, p 111–114
- Manna GK, Khuda-Bukhsh AR (1977) Karyomorphology of cyprzinid fishes and cytological evaluation of the family. Nucleus 20:119–127
- Amemiya CT, Gold JR (1988) Chromosomal NORs as taxonomic and systematic characters in North American cyprinid fishes. Genetica 76(2):81–90
- Kushwaha B, Srivastava SK, Nagpure NS, Ogale SN, Ponniah AG (2001) Cytogenetic studies in two species of Mahseer, *Tor khudree* and *Tor mussullah* (Cyprinidae, Pisces) from India. Chromosome Sci 5:47–50
- Nagpure NS, Kumar Ravindra, Kushwaha B, Srivastava SK, Gopalakrishnan A, Basheer VS, Verma MS (2004) Cytogenetic studies of fish species *Horabagrus nigricollaris, Puntius denisonii* and *P. sarana subnasutus*, endemic to the Western Ghats. Nucleus 47(3):143–148
- Howell WM, Black DA (1980) Controlled silver staining of nucleolus organizer regions with a protective colloidal developer: a one-step method. Experientia 36:1014–1015
- Schmid M, Guttenbach M (1988) Evolutionary diversity of reverse (R) fluorescent chromosome bands in vertebrates. Chromosoma 97:101–114
- 13. Jayaram KC (2002) Fundamentals of fish taxonomy. Narendra Publishing House, Delhi
- Jayaram KC (1991) Revision of the genus *Puntius* Hamilton from the Indian Region (Pisces: Cypriniformes, Cyprinidae, Cyprininae) records of the zoological survey of India, Occasional paper no. 135, Zoological survey of India, Calcutta
- Talwar PK, Jhingran AG (1991) Inland fishes of India and adjacent countries, vol 1. IBH Publishing Co. Pvt. Ltd, New Delhi
- 16. Srivastava G (1980) Fishes of UP and Bihar. Vishwavidyalaya Prakashan, Varanasi
- Bertollo LAC, Takahashi CS, Filho OM (1978) Cytotaxonomic considerations on *Hoplias lacerdae* (Pisces, Erythrinidae). Rev Bras Genet 1(2):103–120
- Levan A, Fredga KY, Sandberg AA (1964) Nomenclature for centromeric position on chromosomes. Hereditas 52:201–220
- Reeves A, Tear J (2000) MicroMeasure for Windows, version 3.3. Free program distributed by the authors over the internet from http://www.colostate.edu/Depts/Biology/MicroMeasure. Accessed 10 Nov 2011
- Sumner AT (1972) A simple technique for demonstrating centromeric heterochromatin. Exp Cell Res 75:304–306

- Fontana F, Tagliavini J, Congiu L, Lanfredi M, Chicca M, Laurenti C, Rossi R (1998) Karyotypic characterization of great sturgeon, *Huso huso*, by multiple staining techniques and fluorescent in situ hybridization. Mar Biol 132:495–501
- 22. Sola L, Rossi AR, Iaselli V, Rasch EM, Monaco PJ (1992) Cytogenetics of bisexual/unisexual species of *Poecilia*. II. Analysis of heterochromatin and nucleolar organizer regions in *Poecilia mexicana* by C-banding and DAPI, quinacrine, chromomycin A₃, and silver staining. Cytogenet Cell Genet 60:229–234
- Yu X, Yu XJ (1990) A Schizothoracine fish species *Diptychus* dipogon with a very high number of chromosomes. Chromosome Info Serv 48:17–18
- 24. Mani I, Kumar R, Singh M, Kushwaha B, Nagpure NS, Srivastava PK, Murmu K, Rao DSK, Lakra WS (2009) Karyotypic diversity and evolution of seven mahseer species (Cyprinidae) from India. J Fish Biol 75:1079–1091
- 25. Ganai FA, Yousuf AR (2011) A karyological analysis of *Puntius conchonius* (Hamilton, 1822) (Pisces, Cyprinidae), a new cyto-type from Dal Lake Srinagar Kashmir, J&K, India. Int J Fish Aquacult 3(11):213–217
- 26. Rishi KK, Rishi S (1981) Giemsa banding in fish chromosome. In: Manna GK, Sinha U (eds) Proceedings 3rd all India congress cytology and genetics. Perspectives Cytol Genet, pp 3: 103–106
- Rab P, Collares-Pereira MJ (1995) Chromosomes of European cyprinid fishes (Cyprinidae, Cypriniformes): a review. Folia Zool 44:193–214
- Nazari S, Pourkazemi M, Porto JIR (2011) Chromosome description and localization of nucleolus organizing regions (NORs) by Ag-staining technique in *Alburnus filippii* (Cyprinidae, Cypriniformes) in Anzali Lagoon, North Iran. Iran J Fish Sci 10(2):352–355
- Collares-Pereira MJ (1985) Cytotaxonomic studies in Iberian cyprinids II. Karyology of Anaecypris hispanica (Steindachner, 1866), Chondrostoma lemming (Steindachner, 1866), Rutilus arcasi (Steindachner, 1866) and R. macrolepidotus (Steindachner, 1866). Cytologia 50:879–890
- Rab P, Roth P (1989) Chromosome studies of European leuciscine fishes (Pisces: Cyprinidae): karyotypes of *Rutilus pigus virgo* and *R. rutilus*. Folia Zool 38:239–245
- Collares-Pereira MJ, Rab P (1995) Chromosomes of European cyprinid fishes (Cyprinidae, Cypriniformes): a review. Folia Zool 44:193–214
- Feldberg E, Porto JIR, Santos EBP, Valentim FC (1999) Cytogenetic studies of two freshwater sciaenids of the genus *Plagioscion* (Perciformes, Sciaenidae) from the central Amazon. Genet Mol Biol 22:351–356
- Feldberg E, Porto JIR, Bertollo LAC (2003) Chromosomal changes and adaptation of chichlid fishes during evolution. In: Val AL, Kapoor BG (eds) Fish adaptation. IBH, New Delhi, pp 287–310
- Molina WF (2006) Chromosome changes and stasis in marine fish groups. In: Pisano E, Ozouf-Costaz C, Foresti F, Kapoor BG (eds) Fish cytogenetics. Science Publisher, Enfield, pp 69–110
- 35. Peres M, Adriano W, Bertollo LAC, Moreira-Filho O (2007) Karyotypic characterization of two species of the genus *Serrapinnus* (Characiformes, Characidae), with the description of a structural polymorphism in *S. heterodon*. Caryologia 60(4):319–324
- Costa GWWF, Molina WF (2009) Karyoevolution of the toadfish *Thalassophryne nattereri* (Batrachoidiformes: Batrachoididae). Genet Mol Res 8(3):1099–1106
- 37. Gui J, Li Y, Zhou Y, Zhou T (1986) Studies on the karyotypes of Chinese cyprinid fishes. VIII. Karyotypic analyses of fifteen species of Barbinae with a consideration for their phyletic evolution. Trans Chin Ichthyol Soc 5:119–127 in Chinese with English abstract

- Magtoon W, Arai R (1993) Karyotyes and distribution of nucleolus organizer regions in Cyprinid fishes from Thailand. Jpn J Ichthyol 40(1):77–85
- 39. Pallavi, Goswami M, Nautiyal P, Malakar AK, Nagpure NS (2012) Genetic divergence and molecular phylogenetics of *Puntius* spp. based on the mitochondrial cytochrome b gene. Mitochondrial DNA 23(6):477–483
- Ren X, Cui J, Yu Q (1992) Chromosomal heterochromatin differentiation of cyprinid fishes. Genetica 87(1):47–51
- Clark MS, Wall WJ (1996) Chromosomes—the complex code. Chapman and Hall, London
- Gold JR, Amemiya CT, Ellison JR (1986) Chromosomal heterochromatin differentiation in North American cyprinid fishes. Cytologia 51:557–566
- 43. Martins C, Galetti PM Jr (1998) Karyotype similarity between two sympatric schizodon fish species (Anostomidae, Aharaciformes) from the Paraguay river basin. Genet Mol Biol 21(3):355–360
- 44. Rábová M, Ráb P, Ozouf-Costaz C, Ene C, Wanzeböck J (2003) Comparative cytogenetics and chromosomal characteristics of ribosomal DNA in the fish genus *Vimba* (Cyprinidae). Genetica 118(1):83–91
- 45. Yunis JJ, Yasmineh WG (1971) Heterochromatin, satellite DNA, and cell function. Structural DNA of eucaryotes may support and protect genes and aid in speciation. Science 174(4015):1200–1209
- Costa GWWF, Molina WF (2009) Karyoevolution of the toadfish *Thalassophryne nattereri* (Batrachoidiformes: Batrachoididae). Genet Mol Res 8(3):1099–1106
- 47. Karahan A, Ergene S (2010) Cytogenetic analysis of *Garra variabilis* (Heckel, 1843) (Pisces, Cyprinidae) from Savur stream (Mardin), Turkey. Turk J Fish Aquat Sci 10:483–489
- Bickham JW, Baker RJ (1979) Canalization model of chromosome evolution. Bull Carnegie Mus Nat Hist 13:70–84
- 49. Nagpure NS, Kumar Ravindra, Srivastava SK, Kushwaha B, Gopalakrishnan A, Basheer VS (2005) A report on high number of chromosomes in three endemic fish species from Western Ghats of India. Nucleus 48(3):165–170
- 50. Gold JR, Amemiya CT (1986) Cytogenetic studies in North American minnows (cyprinidae) XII. Pattern of chromosomal

nucleolus organizer region variation among 14 species. Can J Zool 65:1869-1877

- Gold JR (1984) Silver staining and heteromorphism of nucleolar organizer regions in North American cyprinid fish. Copeia 1:133–139
- Klinkhardt MB (1998) Some aspects of karyoevolution in fishes. Anim Res Dev 47:7–36
- 53. Nagpure NS, Kumar R, Srivastava SK, Kushwaha B, Gopalakrishnan A, Basheer VS (2006) Cytogenetic characterization of two marine ornamental fishes, *Chaetodon collare* and *Stegastes insularis.* J Mar Biol Assoc India 48(2):267–269
- Mank JE, Avise JC (2006) Phylogenetic conservation of chromosome numbers in Actinopterygiian fishes. Genetica 127(1–3): 321–327
- 55. Sahoo PK, Nanda P, Barat A (2007) Karyotypic analysis of *Neolissocheilus hexagonolepis* (McClelland), *Puntius ticto* (Ham.) and *P. Chola* (Ham.) (Family: Cyprinidae, Pisces). Cytologia 72(4):409–413
- 56. Khuda-Bukhsh AR, Chanda T, Barat A (1986) Karyomorphology and evolution in some Indian Hill-stream fishes with particular reference to polyploidy in some species. In: Proceedings of the 2nd International Conference on Indo-Pacific Fishes, Ichthyological Soc Japan, p 886–898
- Sharma OP, Agarwal A (1981) The somatic and meiotic chromosomes of *Puntius chonconius* (Cyprinadae). Genetica 56: 235–237
- Biswal JR, Sahoo PK, Mohanty PK (2010) Karyotypic diversity of some freshwater fish species in rural ponds of Orissa, India. Cytologia 75(3):237–242
- 59. Manna GK, Prasad R (1971) A new perspective in the mechanism on evolution of chromosome in fin fishes. In: Proceedings of first all India congress on cytology and genetics, pp 237–240
- Manna GK, Prasad R (1973) Somatic and germinal chromosomes of two species of fishes belonging to the genus *Puntius*. J Cytol Genet 7:145