Chapter

## Role of Natural Additives on Quality and Shelf Life Extension of Fish and Fishery Products

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#### Abstract

Fish and fishery products have drawn greater attention due to their high nutritional value owing to the presence of cheap superior quality proteins, essential fatty acids, and macro and micronutrients. But higher water content, non- protein nitrogen, and post mortem pH (6–7) in fish favor rapid spoilage by autolysis or putrefaction, and can result in health risk as well as economic loss. Moreover, the quality of fish is affected by species, harvesting season, handling and method of processing. Thus, application of food additives become necessary to maintain the shelf life, nutritional content, texture and flavor of the raw material as well as processed products. Considerable research is being done on applications of natural additives after the emergence of the concept 'Green consumerism' which resulted in decreased consumer preference for using synthetic food additives. In this background, this chapter will review the natural additives used for quality maintenance and shelf life extension of fish and fishery products.

Keywords: Spoilage, Autolysis, Putrefaction, Shelf life, Green consumerism

#### 1. Introduction

Fish and fishery products have become increasingly popular due to their high demand and nutritional value. According to Food and Agriculture Organization (FAO), worldwide production of finfish, mollusks (mainly bivalves), and crustaceans are 54.3, 17.7, and 9.4 million tons respectively [1]. Human consumption of fish is around 88% of total production and among them, 44% consist of live, fresh, or chilled products, and 35% consist of frozen products [2]. But, among animalderived products, fish is considered as the most perishable commodity as it contains a high amount of water, high post mortem pH (greater than 6), non-protein-nitrogen content, free amino acids, lower content of connective tissues, and presence of an osmoregulant, trimethylamine oxide (TMAO) [3]. Spoilage or the deterioration process refers to any change in the condition of food in terms of taste, smell, appearance, or texture and becomes undesirable or unacceptable for human consumption. Generally, the process involves 3 stages; rigor mortis, autolysis, and putrefaction. Rigor mortis or the muscle stiffening will last for hours (time may vary with temperature) after their catch. Subsequently softening occurs due to enzymatic or oxidative self-digestion, and completed by microbiological processes (putrefaction) [4].

#### Food Additives

Every year, chemical and microbial deterioration alone contributes 25% of gross primary product loss (agricultural and fishery products). Besides this, there are several other factors such as harvesting season, type of species, capturing method, handling, the time lag from catch to processing, method of processing, storage temperature, etc. that also influence the rate of spoilage. During spoilage, the breakdown of various components and formation of new compounds responsible for the off- flavor, off odor, discoloration, and texture damage of the fish meat takes place [5]. Therefore, certain food additives have been added to maintain the quality, and the shelf-life of fish and fishery products. The main aim is to combat microbial contamination as well as oxidation for the extension of product's shelf life. Generally, lipid oxidation leads to quality deterioration, and some of them can be detected by organoleptic evaluation, but microbial contamination especially pathogenic microorganisms mostly do not produce sensory deterioration, which act as a challenge for food safety. It emphasizes the importance of the application of antimicrobials in the preservation techniques [6]. In the case of fish and fishery products, preservation techniques draw more scientific attraction, since they represent internationally traded products. Even though many strategies have been developed to prevent chemical and microbial spoilage by chemical preservation, there is still a need for the use of natural preservatives, considering consumer safety. Thus, various researches and efforts have been made to invent more natural alternative solutions in the field of food preservation.

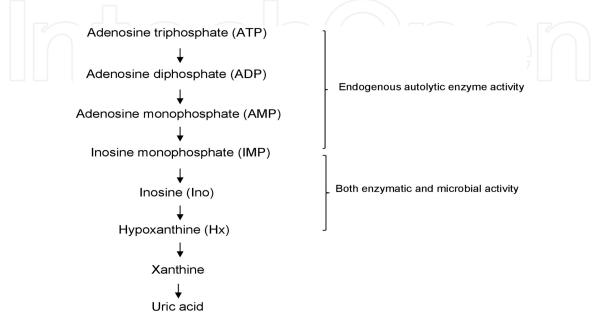
#### 2. Quality changes in fish and fishery products

Fish and fishery products deteriorate rapidly as a consequence of various biochemical breakdowns and microbial activities on the chemical composition of meat. The spoilage involves autolysis or self-digestion of these compounds by digestive enzymes or by free radicals [7]. The major spoilage process is lipid degradation, which mainly occurs through oxidation or hydrolysis. The oxidation could be various types such as photo-oxidation, thermal oxidation, enzymatic oxidation, and auto-oxidation. It can also be accelerated by prooxidants within the body such as hemoglobin, myoglobin, cytochrome c, etc. The process involves the reaction of unsaturated fatty acids of triglycerides with atmospheric oxygen to form unstable primary products like free fatty acids (FFAs), dienes, and peroxides and secondary products like aldehydes, ketones, alcohols, hydrocarbons, volatile organic acids, trienes, epoxy compounds and carbonyls [8]. The process of lipid hydrolysis or lipolysis is breaking down of triglycerides into FFAs by the action of enzyme lipases. Accumulation of these FFAs stimulates protein denaturation, texture damage, and drip loss by the formation of protein-lipid cross-linkages [9]. Generally, protein denaturation in fish occurs mainly by the action of proteolytic enzymes in the muscle (cathepsins) and the intestinal tract (trypsins), which results in muscle solubilization and leads to undesirable texture damage. End products like amino acids, peptides, amines, H<sub>2</sub>S, ammonia, indole, etc. will be formed and will all act as a medium for microbial growth. Microbial breakdown of amino acids will lead to bitterness, souring, bad odor, sliminess, etc. of the flesh [10].

For fish and fishery products, gram-negative bacteria like *Shewanella*, *Photo-bacterium*, *Pseudomonas*, *Moraxella*, *Acinetobacter*, *Flavobacterium*, Aeromonadaceae, and Vibrionaceae and the gram-positive bacteria such as *Bacillus*, *Micrococcus*, *Lactobacillus*, *Clostridium*, and *Corynebacterium* are considered as the major spoilers [3, 10]. The spoilage resulting in off-flavors is due to the formation of specific alcohols, aldehydes, acids, ketones, and sulfur and nitrogen compounds. One of the other major non-nitrogen compounds formed is trimethylamine (TMA) by

the action of several spoilage bacteria on TMAO, an osmoregulant, present in fish (mostly marine and some freshwater fish), and cause a high (positive) redox potential (Eh) in the flesh. Under anoxic conditions, many of the spoilage bacteria utilize TMAO as a terminal hydrogen acceptor, thus allowing them to grow, and resulting in the formation of TMA. TMA reacts with lipids in the muscle to produce the off odor of low-quality fish. This could be a reason for rapid spoilage occurring in seafood than other muscle foods [11]. Thus, microbial spoilage can be determined by TMA level in the product. In the case of shrimps, at above 10°C, indole-positive organisms such as Aeromonas cause subsequent conversion of tryptophan to indole, which is associated with the off-odor of decomposition of shrimp. Thus high levels of indole in the flesh is an indicator of high temperature in the chilled storage process [12]. Clams and oysters undergo fermentative type spoilage also [13]. Generally, the microbial contamination in the fish mainly occurs through microbes associated with the habitat, invasion during processing, handling, and long-term storage. Growth of spoilers differs by habitats like freshwater or marine, temperate or tropical water, and storage or processing conditions. The microbial and chemical stability of food during processing and storage will be determined by the available water for microbial growth, called water activity  $(a_w)$ . Yeast requires  $a_w$  of minimum 0.7 for their growth, and except *Staphylococcus aureus*, most bacteria require at least  $0.9 a_w$  to grow [14]. Thus, it can be said that microbiologically stable fish product is with an aw less than 0.6 [15]. Thus, the water content in the product should be minimum to prevent microbial spoilage. Moreover, pathogenic microbes of public health concern are also taken into consideration as they can produce hazardous toxins. Some of these are; toxin produced by *Clostridium botulinum* (botulinum toxin) in processed food, Scombrotoxin as a result of the microbial conversion of histidine to histamine. Bacteria involved in this process include Morganella morganii, Klebsiella pneumoniae, Hafnia alvei, Pseudomonas putrefaciens, and Clostridium perfringens. Shellfishes can accumulate various algal toxins like brevetoxins, okadaic acids, domoic acids, saxitoxins, etc., and cause serious illness to humans [16].

Another important spoilage mechanism is post-mortem nucleotide catabolism, resulting in ATP depletion and subsequent formation of hypoxanthine (Hx) (**Figure 1**). The breakdown products do not affect the safety but sensory quality undergoes some changes [17, 18]. Based on these compounds formed, the freshness can be expressed. The ratio of inosine (Ino) and Hx to total nucleotides and their



**Figure 1.** Nucleotide catabolism as a result of autolysis and putrefaction.

catabolic derivatives will give the K value, an indicator of loss of freshness [19]. The ratio of Hx to the total of Inosine monophosphate (IMP), Ino, and Hx will give the H value, an indicator of Hx accumulation (bitterness), and its limit for human consumption has been suggested as 60% [20]. Another quality indicator is the F value. It is the ratio of IMP to the total of IMP, Ino, and Hx, and fish with F-value of 10% and higher is considered unacceptable [21]. Thus there is a huge need for the use of additives in the food industry. Application of food additives and low-temperature preservation leads to diminution of most of the spoilage process to a greater extent.

#### 3. Role of chemical additives and natural alternative solutions

By definition, additives are the substances that are added to maintain or improve the safety, freshness, taste, texture, or appearance of food. Generally additives can occur in fish and fishery products during production, processing, storage, packaging, and transportation. Additives can be of two types; Synthetic or chemical, and natural additives. Some of them are listed in **Table 1**.

#### 3.1 Chemical additives

Among chemical additives, the most common and widely used chemical is sodium chloride (NaCl). Salt drying and brining is the most traditional as well as an effective method of food preservation, and several studies have been made to explore all the preservation properties of NaCl. In Nile tilapia fillets, NaCl improved the weight and minimized drip loss [34], showed weight gain in white shrimp (Litopenaeus vannamei) [35], and had anti-melanotic activity along with the shelf-life extension in shrimp (*Xyphopenaeus kroyeri*) [36]. Like salt, sugar is also easily available and is a widely used additive for seafood products. Sugar treatment can significantly reduce pH value and decrease volatile bases like total volatile base nitrogen (TVB-N) [37]. It also showed a cryoprotectant action in frozen surimi (wet protein concentrate) and other products [38, 39], protection of myofibrillar protein [40], decrease the accumulation of biogenic amines in sausages and dry-cured grass carp [41, 42], and prevention of protein denaturation in minced fish meat [43]. The combination of both sugar and salt could also delay spoilage and improve many sensory qualities [37]. The product 'gravad' traditionally manufactured in Nordic countries is prepared by such a combination of sugar and salt [44]. Additives such as table salt and organic acids like acetic acid or citric acid in the Marination technique not only prevent microbial growth but also improve organoleptic properties of fish and fishery products [45]. In seafood, the addition of organic acids provides great preservative action as an antimicrobial agent. Acetic acid and lactic acid, either single-use or combination had a growth-inhibiting effect against pathogens like *Listeria monocytogenes* and *Escherichia coli* [46]. The inhibitory effect of these acids against *L. monocytogenes* was also reported from mussels [47]. Generally, the addition of citric acid showed a positive impact on TVB-N accumulation, toughness, and pH, but a negative impact on the texture and cooking yield of refrigerated shrimp. In such a case, sodium citrate helps to improve cooking yield and texture by preventing excessive pH drop [48]. Sodium or potassium lactate is also considered a good additive for seafood products. It showed shelf-life extension in minced fish products [49], antibacterial effect in sliced salmon [50], in cold-smoked salmon [51], and in catfish fillets [52].

Many other compounds, including phosphates, carbonates, and sulfites, are used as major seafood additives. Phosphate compounds especially, polyphosphates (PP) have been widely used in fish and fishery products as cryoprotectant [38],

Additive function	Categories	Examples	Application	Referenc
To maintain palatability and wholesomeness (preservatives)	Antimicrobial agents	Benzoates, Sorbates, NaCl $\mathrm{NO}_3^-, \mathrm{NO}_2^-$ Organic acids, EOs	Surimi/minced fish products dried, salted or cured fish Fish fillets	[22, 23]
_	Antioxidants	BHA, BHT, TBHQ, PG, Ca/Na propionate, vit E, Ascorbate, Citric acid, Erythorbic acid EOs	Fish oil Fish fillets	[24–26]
To enhance the appeal of foods	Flavor enhancers	MSG, CaCl <sub>2</sub> , Citric acid, Disodium guanylate/ inosinate	Ready to cook or ready to eat products	[25]
-	Sweeteners	Sucrose, lactose, glucose, fructose, glycerol, sorbitol. Acesulfame K, Aspartame, Sodium cyclamate, Saccarin, Sucralose, Neotam, Neohesperidine	Crab meat Fish fillets Marinades	[27, 28]
	Colorants	Carmine, carmosine, caramel, paprika, annatto dye Iron oxides and hydroxides, Ponceau, Cochineals, Oleoresin of turmeric, TiO <sub>2</sub> , FD&C Yellow, Astaxanthin	Paste products, pre-cooked crustaceans, salmon substitutes, surimi, fish roe and smoked fish.	[28, 29]
	Texturizing agents; (Emulsifiers/Stabilizers/ Water-binding agents)	Polysorbates, DATEM, Agar, Alginates, Carrageenan, gum, Calcium stearate, Lecithin, Yeast, Ammonium alum,	Fish and shrimp paste/mince products, Surimi	[25, 30]
To aid in the processing	Moisture control	CaO, Calcium stearate, STPP	Processed products	[25]
	Reduce thaw drip	Na <sub>2</sub> CO <sub>3</sub> , SHMP, STPP, TSPP	Frozen: clams, crab, fillets, lobster, shrimp and minced fish	[31]
	Prevent cracking of glaze	Na <sub>3</sub> PO <sub>4</sub> , Na <sub>2</sub> HPO <sub>4</sub>	Frozen products	[31]
	Anticaking agents	Ca <sub>3</sub> (PO4) <sub>2</sub> , Na <sub>2</sub> SiO <sub>3</sub> , Ca <sub>2</sub> SiO <sub>4</sub> , MgCO <sub>3</sub> , Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	Paste/minced products	[25]
	Prevent discolouration/ blackspot/browning	Na <sub>2</sub> SO <sub>3</sub> 4-Hexylresorcinol, EDTA	Crustaceans	[32]
	Packaging gases	O <sub>2</sub> , CO <sub>2</sub> , N <sub>2</sub> , Ar, O <sub>3</sub> , N <sub>2</sub> O, He, SO <sub>2</sub> , Cl	Fish and fishery products	[33]
	Cryoprotectants	РР	Paste products, fillets, Frozen crustaceans	[28]

**Table 1.**Lists of additives used in fish and fishery products.

gel strength, and flavor enhancer [53], for providing higher cooking yield [31], improving weight, and reducing drip loss [34, 54], modifying texture, color, and reducing cooking loss [55–57], improving quality of fillet [58], minimizing drip loss in shrimp [59, 60], drip loss in sea robin (*Prionotus punctatus*) and pink cuskeel (Genypterus brasiliensis) fillet [31], and weight gain in kutum (Rutilus frisii) fillets [61]. Sodium hexametaphosphate (SHMP) or tripolyphosphate (STPP), or pyrophosphate- tribasic/ tetrabasic (TSPP) are the major phosphate compounds used in processing. Among carbonates, sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) sodium bicarbonate (NaHCO<sub>3</sub>), and magnesium carbonate (MgCO<sub>3</sub>) have been widely used. Weight gain is observed when white shrimp are treated with Na<sub>2</sub>CO<sub>3</sub> and NaHCO<sub>3</sub> [35]. The addition of NaHCO<sub>3</sub> also provides the highest expansion volume for yellow pike conger crackers [62]. Sulfites have been widely used as additives due to their desirable technical properties like preventing melanosis or discoloration. The most predominant sulfiting agent is sodium sulfite used to prevent melanosis in crustaceans like shrimp, lobster, crab, crayfish, etc. [32]. Nitrite is another chemical commonly used as an antimicrobial agent, and effective against *C. botulinum* and its toxin production [63]. A combination of nitrite and sorbic acid would also give the best result as it can inhibit most yeasts. A combination of sorbic acid with benzoic acid could preserve brined shrimp [64]. Moreover, additives such as flavor enhancers, sweeteners, colorants, etc. are used to enhance the appeal of the food. Monosodium glutamate (MSG), calcium chloride (CaCl<sub>2</sub>), and Disodium guanylate/ inosinate are the major flavor enhancers. Commonly used sweetening agents are saccharin, sucralose, glycerol, acesulfame potassium, aspartame, sodium cyclamate, neotam, and neohesperidine. Widely used colorants include carmine, carmosine, caramel, paprika, annatto dye, iron oxides and hydroxides, ponceau, cochineals, titanium dioxide ( $TiO_2$ ), FD&C Yellow, and astaxanthin (Table 1). Butylated hydroxyl toluene (BHT), butylated hydroxyanisole (BHA), tertbutylhydroquinone (TBHQ), propyl gallate (PG), and sodium acetate are widely used synthetic antioxidants to prevent lipid oxidation through free radicals scavenging, breaking chain reactions, peroxide decomposition, and decreasing oxygen concentrations and thereby increasing the shelf life [50]. But preservatives include sulfites, nitrates, benzoates, sorbates, formaldehyde, and others that may possess carcinogenic side effects. Thus nowadays the use of chemical preservatives in food industries steadily decreases and consumers are turning to the use of natural additives.

#### 3.2 Natural additives

#### 3.2.1 Plant-derived products

The use of plant-derived natural compounds such as essential oils, plant extracts, hydrocolloids, phenolic compounds, etc. is very popular in seafood preservation. Their strong antimicrobial and antioxidant activities present great potential for use in the food industry [64–66].

Plant extracts and essential oils can be derived from plant petals, leaves, fruits, peels, stems, roots, and xylems and their antioxidant effects are due to volatile organic compounds, terpenoid, and phenolic components in the plant. The inhibitory effects of essential oil on gram-negative bacteria are less than that of grampositive bacteria as their lipopolysaccharide cell wall of gram-negative bacteria blocked the invasion of hydrophobic oils into the cell membrane [67]. Using essential oils (EOs) and plant extracts to extend shelf-life and maintain the quality of fish and fishery products has been reported frequently. Some of the recent studies of their application in fish and fishery products are represented in **Table 2**. However strong odor and taste, high volatility, complex chemical composition,

Plant derived compounds	Preservative properties	Product	Referen
Essential oils			
Rosemary	Retarded microbial growth, delay chemical deterioration, sensory qualities, and extend the shelf life for 14 days during storage.	Bonito Fish Patties	[68]
Rosemary and basil	Inhibited the formation of TVB-N and lipid oxidation products during storage	Atlantic Mackerel	[69]
Rosemary, laurel, thyme, and sage	Antimicrobial, antioxidant properties, and also enhanced the organoleptic quality	Rainbow trout fillet	[70]
Rosemary, cinnamon fennel and cardamom	The counts of <i>S. aureus, E. coli</i> and <i>Bacillus cereus</i> were reduced	Carp fingers	[71]
Green pepper	Inhibited the growth of wild type strain of <i>P. aeruginosa</i> and attenuated its virulence properties	Fish-based products	[72]
Orange leaf	Enriched with Gelatin film showed shelf- life extension of 10 days	Shrimps	[73]
Carvacrol, bergamot and grapefruit	Improved the quality of fresh fish and extended the shelf-life up to 4 days.	Seabream	[74]
Oregano, thyme, and star anise	Inhibited microbial growth and delaying lipid oxidation	Grass carp	[75]
Ginger	Significant reduction in the TVBN and lipid oxidation	Cobia steaks	[76]
Clove, cumin, and spearmint	Retarded sensory deterioration and formation of biogenic amines.	Red drum fillets	[77]
Black cumin	Higher sensory quality, and extended shelf life	Fresh fish fillets	[78]
limonene	Maintained spoilage bacteria at a lower level and extended the shelf-life of 15 days	Gilthead sea bream fillets	[79]
Allyl Isothiocyanate	Extended the shelf-life by maintain specific spoilage organisms at a lower level	Gilthead sea bream	[80]
Oregano	Activity against Listeria spp.	Salmon	[81]
Fennel	Coating with chitosan nanoparticles reduced the PV, TVBN, TBARS and microbial count	Huso huso fish fillets	[82]
Satureja thymbra leaves	Reduced peroxide value and eliminates secondary oxidation products	Gilthead seabream fillets	[83]
Lemon	Lowered accumulation of histamine, and improved sensory characteristics	Salted sardines	[84]
cinnamon bark	Inhibited <i>Aeromonas</i> and <i>Shewanella</i> , and reduced the accumulation of TVBN, putrescine, cadaverine	Grass carp	[85]
Plant extracts			
Cumin seed and Wild mint leaf	Retarded microbial growth, chemical deterioration, and improved sensory characteristics	Rainbow trout fillets	[86]
Shallot fruit and ajwain seed	Delayed lipid oxidation and microbiological spoilage	Semi-fried coated rainbow trout fillets	[87]

Plant derived compounds	Preservative properties	Product	Reference
Allium paradoxum and Eryngium caucasicum trauve.	Significantly delayed oxidative deterioration and maintain lower bacterial growth	Silver carp fillets	[88]
<i>Punica granatum</i> peels and <i>Hibiscus sabdariffa</i> calyxes	Colorant, preservative, and antimicrobial action	Burger and surimi	[89]
Nothopanax sutellarium leaf	Potential to preserve the fresh fish during transportation	Nile tilapia	[90]
Mint leaf and citrus peel	Retarded the quality changes and extended the shelf life	Indian mackerel	[91]
<i>Syzygium australe</i> and <i>S.</i> <i>luehmannii</i> fruit and leaf	Inhibition to <i>Shewanella</i> spp.	Fresh and cold storage fish	[92]
Kakadu plum	Inhibited bacterial growth for 15 days at 4°C	Fish fillets	[93]
Clove, Sage and kiwifruit peel	Antioxidant and antimicrobial potential, and extend the shelf life	Fish fingers	[94]
Pomegranate, rosemary, and olive	Delayed the lipid oxidation, and microbial count	Fish patties	[95]
<i>Plectranthus amboinicus</i> leaf	Improved the color, rehydration and water activity	Fish oil fortified soup powder	[96]
Green Tea Leaves and Fenugreek Seeds	Decreased TVBN, TBARS, total bacterial count, and pH	Shrimp	[97]
Basil leaf	Antimicrobial effect and longest shelf life	Mullet fillet	[98]

#### Table 2.

Summary of some of the recent studies of application of essential oils, and plant extracts in fish and fishery products.

low bioavailability and stability, and factors affecting chemical compositions like plant genetic variability, extraction techniques, etc. are some limitations for the application of these phytogenic additives [99]. Like other plant-derived products, seaweed and algal extracts are emerging as a rich source of natural antioxidants, along with many nutritional values. The three important widely used hydrocolloids are; agar-agar, align, and Carrageenan. As thickening agent agar-agar is used mainly in fish paste products. Carrageenan is used to enhance the gelling property of fish mince [100–102], and organoleptic properties of mussels and squids [103]. The sodium salt of alginic acid is widely used as a stabilizer and thickener in coating films. Sodium alginate coating with rosemary extract reduced the accumulation of biogenic amines and bacterial count in Abalone (Haliotis discus hannai) [104]. Coating with gingerol delayed lipid oxidation, protein degradation, nucleotide breakdown, and inhibited microbial growth in Seabream (*Pagrosomus major*) [105]. Coating with tea polyphenols had significantly lowered the levels of TVB-N, lipid oxidation, and protein decomposition in Japanese Sea Bass (*Lateolabrax japonicas*) fillets [106]. The use of alginate-calcium film coating with Citrus wilsonii extract delays the deterioration and results in a higher sensory score for L. vannamei [107]. Significant reduction in the TVB-N, TMA, and thiobarbituric acid reactive substance (TBARS) has been detected during chilled storage with the presence of Gracilaria gracilis extract in shrimp [108], G. verrucosa extract in Indian mackerel [109], and extracts of Hypnea musciformis and A. muscoides in black tiger shrimp [110]. Similarly, seaweeds like Sargassum kjellmanianurn [111], and Grateloupia

*filicina* [112] exhibits a good antioxidant activity and prevent lipid oxidation in fish oil. Extracts of seaweeds such as *Fucus vesiculosus* inhibited the hemoglobinmediated lipid oxidation in washed cod muscle and cod protein isolates [113], and extracts of *Durvillaea antarctica* (cochayuyo/ulte), *Pyropia columbina* (red luche), and *Ulva lactuca* (sea lettuce) improved the lipid and sensory qualities in canned salmon [114]. Some phenolic compounds like flavonoids, phenolic acids, hydroxycinnamic acid, and lignans are also used as plant derived natural additives [115]. In surimi-derived products, several hydrocolloids like konjac enhance the gelling property [116]. Other products like starch [101, 102], gums such as garrofin, guar, xanthan [117], etc. also provide a gelling effect and assure elasticity of the product. Iota carrageenan and xanthan had a cryoprotective effect too [118]. Other plantderived products such as soybean protein, wheat gluten, and starch are also used as additives for fish-paste products [119].

#### 3.2.2 Animal-derived products

Nowadays, animal-derived products like chitosan, gelatin, and whey proteins are widely used as food additives. Chitosan is a natural polymer obtained from chitin, a component of the exoskeleton of shellfish and fungal cell walls. Gelatin is a protein derived from the raw collagen of animal body parts. Whey protein is one of the two proteins, other than casein, found in milk. The bioactive coating of food products with these compounds provides antioxidant and antimicrobial properties and can thereby increase the shelf life of the product. Direct addition of compounds into the packaging materials also provided more potent preservative action [120]. Some of the recent studies on the application of chitosan, gelatin, and whey protein as edible coatings in fish and fish product preservation are represented in **Table 3**. But in moist environments, edible films and coating showed relatively low stiffness and strength, thus limited their use in specific conditions. Another animal-derived product, bioactive peptide (specific protein fragments) showed antimicrobial [137], and antioxidant activities [138]. In fish paste products, the products like plasma hydrolysate, plasma protein, ovomucoid, egg albumin, egg white, etc. were added as additives for improving strength [119]. The binding effect of egg whites and hydrolyzed beef plasma proteins in surimi gels [139], gel enhancing effect of bovine plasma powder, and egg white powder in arrow tooth surimi [140] were also reported.

#### 3.2.3 Microbial-derived products

Bacteriocin, a major bacterial-derived bio preservative (mostly from *Lactobacillus*) has potent antimicrobial properties. The mode of action is interfering cell wall synthesis of bacteria by pore formation and squeezing out of the inner material thereby restricting their growth [141]. Along with this antimicrobial action, other properties like nontoxicity, active in a wide range of pH and temperature, etc. making them generally recognized as safe (GRAS) additive [142]. The most common bacteriocins produced by *Lactobacillus* are Nicin, lacticin, pediocin, etc. Many bacteriocins are known to be more effective against endospore-forming bacteria. Bacteriocins were used to reduce the counts of *Salmonella* and *Vibrio* spp. in marine fishes and loligo [143], *Listeria inaqua*, and *Pseudomonas* spp. in fish homogenates [144], and aerobic and anaerobic bacteria in cold smoked salmon [145]. A novel bacteriocin BCC7293 from *Weissella hellenica* showed activity against *L. monocytogenes, S. aureus, P. aeruginosa, A. hydrophila, E. coli*, and *S. Typhimurium* in Pangasius fillets [146]. Bacteriocin FGC-12 and DY4–2 produced by *Lactobacillus plantarum* showed some inhibitory effect on *Vibrio parahaemolyticus* in shrimp [147], and *Pseudomonas fluorescens* in turbot fillet

Fish product	Preservative action	Referen
Chitosan-based		
Grass carp	Inhibited cathepsin activities and thereby retarded the proteolysis	
Olive flounder	Coating combined with clove oil improved the quality and shelf life was extended by 6 days	
White shrimp	Coating combined with pomegranate peel extract reduced the melanosis, TVBN and microbial count	
Mackerel	Coating combined with gallic acid decreased microbial growth, protein decomposition, biogenic amine formation, lipid oxidation and nucleotide breakdown and shelf life was extended	
Fish burgers	The chitosan film containing lactoperoxidase system suppress the increase of <i>Pseudomonas</i> spp. and <i>Shewanella</i> spp. and TBARS value	
Mori Fish	Spoilage reduction when coated with rosemary extract on storage	
Gelatin-based		
Rainbow trout fillets	Coating incorporating oregano essential oil minimized the formation of volatile bases, oxidation products, and the growth of total and psychrotrophic bacteria	
Shrimp	Fish gelatin reduced lipid oxidation and extend shelflife	
Shrimp	Coating enriched with orange leaf essential oil extend shelf life about 10 days	
Tilapia fillets	Combined with grape seed extract reduced the formation of undesirable metabolites like TMA and histidine significantly	
Chitosan-gelatin-b	ased	
Golden- pomfret fillets	Inhibits myofibril degradation	[130]
White shrimp	Decreased the total and psychrotrophic bacteria and increased the shelf-life	
Salmon fillet	Incorporated with garlic and lime extracts extended the shelf life by antibacterial and antioxidant activity	
Minced trout fillet	Incorporated with grape seed extract <i>Ziziphora clinopodioides</i> essential oil reduced the spoilage	[133]
Whey protein-base	ed	
Pike-Perch	Coating with lactoperoxidase system and $\alpha$ -tocopherol extended the shelflife significantly	[134]
Rainbow trout fillet	Reduced microbial growth, and TVB-N and TBA	[135]
Rainbow trout fillet	Inhibited Shewanella and P. fluorescens and shelf life was extended	[136]

#### Table 3.

Some of the recent studies on the application of chitosan, gelatin, and whey protein as edible coatings in fish and fish product preservation.

[148] respectively. Bacteriocin LJR1produced by *Pediococcus pentosaceus* showed activity against *L. monocytogenes* in white shrimp [149]. Bacteriocin GP1 produced by *Lactobacillus rhamnosus* active against Coliforms, *Aeromonas*, and *Vibrio* spp. in fish fillets [150]. The combination of bacteriocins with other preservation techniques usually results in better action. Microencapsulated *Ziziphora clinopodioides* essential oil and Nisin showed the strongest effect on preserving the sensorial quality of fish

burgers [151]. However the use of bacteriocin is limited due to its high cost. Another microbial-derived product kojic acid, a natural product of many fungi like *Aspergillus* and *Penicillium*, has certain anti-enzymatic browning and antibacterial effects, especially against gram-negative bacteria [152]. A combination of kojic acid and tea polyphenols showed an antibacterial effect against spoilage bacteria in refrigerated seabass (*Lateolabrax japonicas*) [153].  $\varepsilon$ -Polylysine is another microbial-derived product with excellent preservative properties. It was isolated originally from bacteria *Streptomyces albulus*. Treatment with  $\varepsilon$ -Polylysine lowered the TVB-N, putrescine, cadaverine, and hypoxanthine and extended the shelf-life of shrimp [154]. The addition of  $\varepsilon$ -polylysine chitosan and carrageenan showed shelf life extension of Chinese shrimp (*Fenneropenaeus chinensis*) [155], and chitosan-based coatings combined with  $\varepsilon$ -polylysine and rosmarinic acid contributed to the reduction of TVB-N, TMA, and ATP-related compounds in Half-smooth tongue sole fillets [156]. A combination of plant, animal, and microbial-derived products showed the strongest preservative action than the independent use.

#### Conclusions

As a perishable food commodity, most of the world's supply of fish and fishery products are lost through chemical and microbial spoilage than other reasons like improper storage, handling and processing damage. Thus, the increasing demand for good quality fish products has intensified the search for applications of additives in preservation strategies. It is well known that none of the additives offer complete protection against spoilage, but can improve the quality of fish as well as shelf life to a greater extent. By considering the potential health hazards associated with chemicals as well as consumer preference, application of natural products from cheap and underutilized resources enabling food safety holds promise.

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### References

[1] FAO yearbook: Fishery and aquaculture statistics 2017. Rome: Food and Agriculture Organization of the United Nations. 2019:50-52

[2] FAO. The state of world fisheries and aquaculture 2020. Rome: Food and Agriculture Organization of the United Nations. 2020:60-61. https://doi. org/10.4060/ca9229en

[3] Gram L, and Huss HH. Microbiological spoilage of fish and fish products. International journal of food microbiology. 1996;*33*:121-137.

[4] Sriket C. Proteases in fish and shellfish: Role on muscle softening and prevention. International Food Research Journal. 2014; 21:433-445

[5] Baird-Parker TC. The production of microbiologically safe and stable foods. The microbiological safety and quality of food. 2000;1:3-18.

[6] Mahmud A, Abraha B, Samuel M, Abraham W, Mahmud E. Fish preservation: A multi-dimensional approach. MOJ Food Processing and Technology. 2018;6:303-310.

[7] FAO. Post-harvest changes in fish. In: FAO Fisheries and Aquaculture Department, Food and Agriculture Organization, Rome, Italy. 2005

[8] Ghaly AE, Dave D, Budge S, Brooks MS. Fish spoilage mechanisms and preservation techniques. American journal of applied sciences. 2010;7:859.

[9] Mackie IM. The effects of freezing on flesh proteins. Food Reviews International. 1993;9:575-610.

[10] Gram L, Dalgaard P. Fish spoilage bacteria — Problems and solutions. Current Opinion in Biotechnology.2002;13:262-266. [11] Gram L, Trolle G, Huss HH. Detection of specific spoilage bacteria from fish stored at low (0 C) and high (20 C) temperatures. International journal of food microbiology. 1987;4:65-72.

[12] Miget RJ. Microbiology of crustacean processing: shrimp, crawfish, and prawns. In Microbiology of marine food products. Springer, Boston, MA.1991:65-87.

[13] Fieger EA, Novak AF. Microbiology of shellfish deterioration. Fish as food. 1961:561-611.

[14] Ratkowsky DA, Olley J,McMeekin TA, Ball A. Temperature and bacteria. Journal of Bacteriology.1982;149:1-5.

[15] Abbas KA, Saleh AM, Mohamed A, Lasekan O. The relationship between water activity and fish spoilage during cold storage: A review. Journal of Food Agriculture and Environment. 2009:86-90.

[16] Özogul F, Hamed I. Marine-based toxins and their health risk. InFood Quality: Balancing Health and Disease. Academic Press. 2018:109-144.

[17] Masniyom P. Deterioration and shelf-life extension of fish and fishery products by modified atmosphere packaging. Songklanakarin Journal of Science & Technology. 2011;33.

[18] Visciano P, Schirone M, Tofalo R, Suzzi G. Biogenic amines in raw and processed seafood. Frontiers in microbiology. 2012;3:188.

[19] Howgate P. A review of the kinetics of degradation of inosine monophosphate in some species of fish during chilled storage. International journal of food science & technology.2006;41:341-353.

[20] Luong JHT, Male KB, Masson C, Nguyen AL. Hypoxanthine ratio determination in fish extract using capillary electrophoresis and immobilized enzymes. Journal of Food Science. 1992;57:77-81. doi: 10.1111/ j.1365-2621.1992.tb05429.

[21] Kuley E, Özogul F, Özogul Y. Effects of aluminium foil and cling film on biogenic amines and nucleotide degradation products in gutted sea bream stored at 2±1 C. European Food Research and Technology. 2005;221:582-591.

[22] Syamdidi M. The Use of Chemical Additives for Fisheries Product Preservation. Squalen Bulletin of Marine and Fisheries Postharvest and Biotechnology. 2013;7:79-87.

[23] Desai MA, Soni KA, Nannapaneni R, Schilling MW, Silva JL. Reduction of *Listeria monocytogenes* in raw catfish fillets by essential oils and phenolic constituent carvacrol. Journal of Food Science 2012;77:16-22.

[24] Kaitaranta JK. Control of lipid oxidation in fish oil with various antioxidative compounds. Journal of the American Oil Chemists Society.1992;69:810-813.

[25] Taylor SL, Nordlee JA. Chemical additives in seafood products. Clinical reviews in allergy. 1993;11:261-291.

[26] Veeck AP, Klein B, Ferreira LF, Becker AG, Heldwein CG, Heinzmann BM, Baldisserotto B, Emanuelli T. Lipid stability during the frozen storage of fillets from silver catfish exposed in vivo to the essential oil of *Lippia alba* (Mill.) NE Brown. Journal of the Science of Food and Agriculture. 2013;93:955-960.

[27] Patel S. Plant essential oils and allied volatile fractions as multifunctional additives in meat and fish-based food products: a review. Food Additives & Contaminants: Part A. 2015;32: 1049-1064.

[28] Fishery Products Additives Regulations Legal Notice No 65/2003

[29] Jeyakumari A. Surimi and surimi based products. Central Institute of Fisheries Technology, Cochin. 2014.

[30] Lee CM. Ingredient and formulation technology for surimi-based products. Surimi Technology. 1992:273-302.

[31] Gonçalves AA, Ribeiro JL. Optimization of the freezing process of red shrimp (*Pleoticus muelleri*) previously treated with phosphates. International Journal of refrigeration. 2008;31:1134-1144.

[32] Gonçalves AA, de Oliveira AR. Melanosis in crustaceans: A review. LWT-Food Science and Technology. 2016;65:791-799.

[33] DeWitt CA, Oliveira A. Modified atmosphere systems and shelf life extension of fish and fishery products. Foods. 2016;5:48

[34] Gonçalves AA, Souza MA, Regis RC. Effects of different levels of food additives on weight gain, cookrelated yield loss, physicochemical and sensorial quality of Nile tilapia fillets (*Oreochromis niloticus*). International Food Research Journal. 2018;25.

[35] Chantarasuwan C, Benjakul S, Visessanguan W. Effects of sodium carbonate and sodium bicarbonate on yield and characteristics of Pacific white shrimp (*Litopenaeus vannamei*). Food science and technology international. 2011;17:403-414.

[36] Fossati AA, Bergmann GP, Ribeiro LA, Júnior DP, Schneider TM, Kindlein L. Effects of different additives on colorimetry and melanosis prevention of Atlantic seabob shrimp (*Xyphopenaeus kroyeri*) stored under refrigeration. International Journal of Fisheries and Aquaculture. 2016;8:74-80.

[37] Fan H, Luo Y, Yin X, Bao Y, Feng L. Biogenic amine and quality changes in lightly salt-and sugar-salted black carp (*Mylopharyngodon piceus*) fillets stored at 4 C. Food Chemistry. 2014;159:20-28.

[38] Sych J, Lacroix C, Adambounou LT, Castaigne F. The effect of lowor nonsweet additives on the stability of protein functional properties of frozen cod surimi. Journal of Food Science and Technology. 1991;26: 185-197.

[39] Sultanbawa Y, Li-Chan EC. Cryoprotective effects of sugar and polyol blends in ling cod surimi during frozen storage. Food research international. 1998;31:87-98.

[40] Lee CM. Surimi process technology.Food technology (Chicago).1984;38:69-80.

[41] Bover-Cid S, Izquierdo-Pulido M, Vidal-Carou MC. Changes in biogenic amine and polyamine contents in slightly fermented sausages manufactured with and without sugar. Meat Science. 2001;57:215-221.

[42] Zhang J, Liu Z, Hu Y, Fang Z, Chen J, Wu D, Ye X. Effect of sucrose on the generation of free amino acids and biogenic amines in Chinese traditional dry-cured fish during processing and storage. Journal of food science and technology. 2011;48:69-75.

[43] Toyoda K. The surimi manufacturing process. Surimi technology. 1992:79-112.

[44] Lyhs U, Lahtinen J, Fredriksson-Ahomaa M, Hyytiä-Trees E, Elfing K, Korkeala H. Microbiological quality and shelf-life of vacuumpackaged 'gravad'rainbow trout stored at 3 and 8 C. International Journal of Food Microbiology. 2001;70:221-230 [45] Kilinc B, Cakli S. Chemical, enzymatical and textural changes during marination and storage period of sardine (*Sardina pilchardus*) marinades. European Food Research and Technology. 2005;221:821-827.

[46] Young KM, Foegeding PM. Acetic, lactic and citric acids and pH inhibition of *Listeria monocytogenes* Scott A and the effect on intracellular pH. The Journal of applied bacteriology. 1993;74:515-520.

[47] Conner DE, Scott VN, Bernard DT. Growth, inhibition, and survival of *Listeria monocytogenes* as affected by acidic conditions. Journal of Food Protection. 1990;53:652-655.

[48] Agrafioti PT, Katsanidis E. Effects of additives on the selected quality attributes and cooking yield of squid: Modelling and optimization. International Journal of food properties. 2012;15:579-589.

[49] Birkeland S, Rotabakk BT. Effects of Additives and Packaging Method on Quality and Microbiological Characteristics in Mild Thermal Processed Fish Mince. Journal of Aquatic Food Product Technology.
2014;23:368-384.

[50] Sallam KI. Antimicrobial and antioxidant effects of sodium acetate, sodium lactate, and sodium citrate in refrigerated sliced salmon. Food control. 2007;18:566-575.

[51] Vogel BF, Ng YY, Hyldig G, Mohr M, Gram L. Potassium lactate combined with sodium diacetate can inhibit growth of *Listeria monocytogenes* in vacuum-packed cold-smoked salmon and has no adverse sensory effects. Journal of food protection. 2006;69:2134-2142.

[52] Williams SK, Rodrick GE, West RL. Sodium lactate affects shelf life and consumer acceptance of fresh catfish (*Ictalurus nebulosus*, *marmoratus*) fillets

under simulated retail conditions. Journal of Food Science. 1995;60:636-639.

[53] Hui YH. Handbook of food science, technology, and engineering. CRC press. 2006.

[54] Wangtueai S, Tongsiri S, Maneerote J, Supaviriyakorn W. Effect of phosphate on frozen Nile tilapia fillets. Food and Applied Bioscience Journal. 2014;2:203-215

[55] Lampila LE. Functions and uses of phosphates in the seafood industry. Journal of Aquatic Food Product Technology. 1993;1:29-41.

[56] Chang CC, Regenstein JM. Water uptake, protein solubility, and protein changes of cod mince stored on ice as affected by polyphosphates. Journal of Food Science. 1997;62:305-309.

[57] Gonçalves AA. Phosphates for seafood processing. Phosphates: Sources, Properties, and Applications. Hauppauge. 2012:83-112.

[58] Kilinc B, Cakli S, Cadun A, Sen B. Effects of phosphate dip treatments on chemical, microbiological, color, textural, and sensory changes of rainbow trout (*Onchorhyncus mykiss*) fillets during refrigerated storage. Journal of Aquatic Food Product Technology. 2009;18:108-119.

[59] Moawad RK, Ashour MM, Mohamed GF, El-Hamzy EM. Effect of food grade trisodium phosphate or water dip treatments on some quality attributes of decapitated white marine shrimp (*Penaeus* spp.) during frozen storage. Journal of Applied Sciences Research. 2013;9:3723-3734.

[60] Carneiro CD, Mársico ET, Ribeiro RD, Conte Junior CA, Álvares TS, De Jesus EF. Quality Attributes in Shrimp Treated with Polyphosphate after Thawing and Cooking: A Study Using Physicochemical Analytical Methods and Low-Field 1 H NMR. Journal of Food Process Engineering. 2013;36:492-499.

[61] Etemadian Y, Shabanpour B, Mahoonak AS, Shabani A. Combination effect of phosphate and vacuum packaging on quality parameters of *Rutilus frisii* kutum fillets in ice. Food Research International. 2012;45:9-16.

[62] Peranginangin R, Fawzya YN, Sugiyono S, Muljanah I. Food additives and effect of thickness on fish crackers quality.1997:106-114

[63] Tompkin RB, Branen AL, Davidson PM. Antimicrobials in food. Nitrite. Davidson, PM. 2005:169-236.

[64] Einarsson H, Lauzon HL.
Biopreservation of brined shrimp (*Pandalus borealis*) by bacteriocins from lactic acid bacteria. Applied and Environmental Microbiology.
1995;61:669-676.

[65] Benkeblia, N. Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*). LWT-Food Science and Technology. 2004;37:263-268. https:// doi.org/10.1016/j.lwt.2003.09.001

[66] Chouliara E, Karatapanis A, Savvaidis I, Kontominas M. Combined effect of oregano essential oil and modified atmosphere packaging on shelf-life extension of fresh chicken breast meat, stored at 4 °C. Food Microbiology. 2007;24:607-617.

[67] Bajpai VK, Rahman A, Kang SC. Chemical composition and inhibitory parameters of essential oil and extracts of *Nandina domestica* Thunb. to control food-borne pathogenic and spoilage bacteria. International Journal of Food Microbiology. 2008;125:117-122.

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[68] Guran HS, Oksuztepe G, Coban OE, Incili GK. Influence of different essential oils on refrigerated fish patties produced from bonito fish (*Sarda sarda* Bloch, 1793). Czech Journal of Food Sciences. 2015;33:37-44.

[69] Karoui R, Hassoun A. Efficiency of rosemary and basil essential oils on the shelf-life extension of Atlantic mackerel (*Scomber scombrus*) fillets stored at 2 C. Journal of AOAC International. 2017;100:335-344.

[70] Ozogul Y, Yuvka İ, Ucar Y, Durmus M, Kösker AR, Öz M, Ozogul F. Evaluation of effects of nanoemulsion based on herb essential oils (rosemary, laurel, thyme and sage) on sensory, chemical and microbiological quality of rainbow trout (*Oncorhynchus mykiss*) fillets during ice storage. Lebensmittel-Wissenschaft & Technologie. 2017;75:677-684.

[71] Abdeldaiem MH, Ali HG, Ramadan MF. Impact of different essential oils on the characteristics of refrigerated carp (*Cyprinus carpio*) fish fingers. Journal of Food Measurement and Characterization. 2017;11:1412-1420.

[72] Myszka K, Olejnik A, Majcher M, Sobieszczańska N, Grygier A, Powierska-Czarny J, Rudzińska M. Green pepper essential oil as a biopreservative agent for fish-based products: Antimicrobial and antivirulence activities against *Pseudomonas aeruginosa* KM01. Lebensmittel-Wissenschaft & Technologie. 2019;108:6-13.

[73] Alparslan Y, Yapıcı HH, Metin C, Baygar T, Günlü A, Baygar T. Quality assessment of shrimps preserved with orange leaf essential oil incorporated gelatin. LWT-Food Science and Technology. 2016;72:457-466.

[74] Navarro-Segura L, Ros-Chumillas M, López-Cánovas AE, García-Ayala A, López-Gómez A. Nanoencapsulated essential oils embedded in ice improve the quality and shelf life of fresh whole seabream stored on ice. Heliyon. 2019;5:1804.

[75] Huang Z, Liu X, Jia S, Zhang L, Luo Y. The effect of essential oils on microbial composition and quality of grass carp (*Ctenopharyngodon idellus*) fillets during chilled storage. International journal of food microbiology. 2018;266:52-59.

[76] Remya S, Mohan CO, Venkateshwarlu G, Sivaraman GK, Ravishankar CN. Combined effect of O2 scavenger and antimicrobial film on shelf life of fresh cobia (*Rachycentron canadum*) fish steaks stored at 2 C. Food Control. 2017;71:71-78.

[77] Cai L, Cao A, Li Y, Song Z, Leng L,
Li J. The effects of essential oil
treatment on the biogenic amines
inhibition and quality preservation of
red drum (*Sciaenops ocellatus*) fillets.
Food Control. 2015;56:1-8.

[78] Ozpolat E, Duman M. Effect of black cumin oil (*Nigella sativa* L.) on fresh fish (*Barbus grypus*) fillets during storage at 2±1 C. Food Science and Technology. 2017;37:148-152.

[79] Giarratana F, Muscolino D, Beninati C, Ziino G, Giuffrida A, Panebianco A. Activity of R (+) limonene on the maximum growth rate of fish spoilage organisms and related effects on shelf-life prolongation of fresh gilthead sea bream fillets. International journal of food microbiology. 2016;237:109-113.

[80] MuSColino D, GiArrAtAnA F, BEninAti C, Ziino G, GiuFFriDA A, PAnEBiAnCo A. Effects of allyl isothiocyanate on the shelf-life of gilthead sea bream (*Sparus aurata*) fillets. Czech Journal of Food Sciences. 2016;34:160-165.

[81] Pedrós-Garrido S, Clemente I, Calanche JB, Condón-Abanto S,

Beltrán JA, Lyng JG, Brunton N, Bolton D, Whyte P. Antimicrobial activity of natural compounds against *Listeria* spp. and their effects on sensory attributes in salmon (*Salmo salar*) and cod (*Gadus morhua*). Food Control. 2020;107:106768.

[82] Maghami M, Motalebi AA,
Anvar SA. Influence of chitosan
nanoparticles and fennel essential oils
(*Foeniculum vulgare*) on the shelf life of *Huso huso* fish fillets during the storage.
Food science & nutrition.
2019;7:3030-3041.

[83] Choulitoudi E, Bravou K, Bimpilas A, Tsironi T, Tsimogiannis D, Taoukis P, Oreopoulou V. Antimicrobial and antioxidant activity of *Satureja thymbra* in gilthead seabream fillets edible coating. Food and bioproducts processing. 2016;100:570-577.

[84] Alfonzo A, Martorana A, Guarrasi V, Barbera M, Gaglio R, Santulli A, Settanni L, Galati A, Moschetti G, Francesca N. Effect of the lemon essential oils on the safety and sensory quality of salted sardines (*Sardina pilchardus* Walbaum 1792). Food Control. 2017;73:1265-1274.

[85] Huang Z, Liu X, Jia S, Luo Y. Antimicrobial effects of cinnamon bark oil on microbial composition and quality of grass carp (*Ctenopharyngodon idellus*) fillets during chilled storage. Food Control. 2017;82:316-324.

[86] Raeisi S, Quek SY, Ojagh SM, Alishahi AR. Effects of Cumin (*Cuminum cyminum* L.) Seed and Wild Mint (*Mentha Longifolia* L.) Leaf Extracts on the Shelf Life and Quality of Rainbow Trout (*Oncorhynchus Mykiss*) Fillets Stored at 4C±1. Journal of Food Safety. 2016;36:271-281.

[87] Raeisi S, Sharifi-Rad M, Quek SY, Shabanpour B, Sharifi-Rad J. Evaluation of antioxidant and antimicrobial effects of shallot (*Allium ascalonicum* L.) fruit and ajwain (*Trachyspermum ammi* (L.) Sprague) seed extracts in semi-fried coated rainbow trout (*Oncorhynchus mykiss*) fillets for shelf-life extension. LWT-Food Science and Technology. 2016;65:112-121.

[88] Raeisi S, Ojagh SM, Sharifi-Rad M, Sharifi-Rad J, Quek SY. Evaluation of *Allium paradoxum* (MB) G. Don. and *Eryngium caucasicum* trauve. Extracts on the shelf-life and quality of silver carp (*Hypophthalmichthys molitrix*) fillets during refrigerated storage. Journal of Food Safety. 2017;37:12321.

[89] Tayel AA, Almabady NA, Sorour NM, Diab AM. Application of natural plant extracts as colorants, preservatives, and anti-listerial agents in processed fish products. Journal of Food Safety. 2018;38:12435.

[90] Safrida S, Hardania DI, Khairil K, Muhammad N, Nur YI. Evaluation natural preservatives of *Nothopanax scutellarium* merr. leaf extract in physical characteristics of nile tilapia (*Orechromis niloticus*). InIOP Conference Series: Earth and Environmental Science. IOP Publishing 2021;667:12060

[91] Viji P, Binsi PK, Visnuvinayagam S, Bindu J, Ravishankar CN, Gopal TK. Efficacy of mint (*Mentha arvensis*) leaf and citrus (*Citrus aurantium*) peel extracts as natural preservatives for shelf life extension of chill stored Indian mackerel. Journal of food science and technology. 2015;52:6278-6289.

[92] Murhekar S, Wright MH, Greene AC, Brownlie JC, Cock IE. Inhibition of *Shewanella* spp. growth by *Syzygium australe* and *Syzygium luehmannii* extracts: natural methods for the prevention of fish spoilage. Journal of food science and technology. 2017;54:3314-3326.

[93] Wright MH, Shalom J, Matthews B, Greene AC, Cock IE. *Terminalia ferdinandiana* Exell: Extracts inhibit *Shewanella* spp. growth and prevent fish spoilage. Food microbiology. 2019;78:114-122.

[94] Abdel-Wahab M, El-Sohaimy SA, Ibrahim HA, El-Makarem HS. Evaluation the efficacy of clove, sage and kiwifruit peels extracts as natural preservatives for fish fingers. Annals of Agricultural Sciences. 2020;65:98-106.

[95] Martínez L, Castillo J, Ros G, Nieto G. Antioxidant and antimicrobial activity of rosemary, pomegranate and olive extracts in fish patties. Antioxidants. 2019;8:86.

[96] Lekshmi RG, Jayathilakan K, Sarika K, Priya ER, Greeshma SS, Sultana K, Tejpal CS, Mathew S. Effect of *Plectranthus amboinicus* leaf extract on the quality attributes of microencapsulated fish oil fortified soup powder. Krishi. 2019

[97] Hatab S, Lin K, Miao W, Chen M, Lin J, Deng S. Potential Utilization of Green Tea Leaves and Fenugreek Seeds Extracts as Natural Preservatives for Pacific White Shrimp During Refrigerated Storage. Foodborne pathogens and disease. 2018;15:498-505.

[98] Shofiani S, Rostini I, Afrianto E. The Effect of Concentrations of Basil Leaves Extract as Natural Preservatives in Mullet Fillet on Bacterial Growth in Low-Temperature Storage. Asian Journal of Fisheries and Aquatic Research. 2020:30-41.

[99] Stevanović ZD, Bošnjak-Neumüller J, Pajić-Lijaković I, Raj J, Vasiljević M. Essential oils as feed additives—future perspectives. Molecules. 2018;23:1717.

[100] Borderías J, Montero P, Marti de Castro MA. Gelificación de serrín de merluza (*Merluccius australis*)/Gelling of hake (*Merluccius australis*) sawdust. Food Science and Technology International. 1996;2:293-299. [101] Gómez-Guillén C, Borderías AJ, Montera P. Thermal gelation properties of two different composition sardine (*Sardina pilchardus*) muscles with addition of non-muscle proteins and hydrocolloids. Food Chemistry. 1997;58:81-87.

[102] Park JW. Ingredient technology and formulation development. Food Science and Technology-New York-Marcel Dekker-. 2000:343-392.

[103] Guldas M, Hecer C. Influences of the selected additives on the weight loss and organoleptic properties of marinated mussels and squids. Acta Veterinaria Brno. 2013;81:263-267.

[104] Hao R, Liu Y, Sun L, Xia L, Jia H, Li Q, Pan J. Sodium alginate coating with plant extract affected microbial communities, biogenic amine formation and quality properties of abalone (*Haliotis discus hannai* Ino) during chill storage. LWT-Food Science and Technology. 2017;81:1-9.

[105] Cai L, Wang Y, Cao A, Lv Y, Li J. Effect of alginate coating enriched with 6-gingerol on the shelf life and quality changes of refrigerated red seabream (*Pagrosomus major*) fillets. RSC Advances. 2015;5:36882-36889.

[106] Nie X, Wang L, Wang Q, Lei J, Hong W, Huang B, Zhang C. Effect of a sodium alginate coating infused with tea polyphenols on the quality of fresh japanese sea bass (*Lateolabrax japonicas*) fillets. Journal of food science. 2018;83:1695-1700.

[107] Liu X, Jia Y, Hu Y, Xia X, Li Y, Zhou J, Liu Y. Effect of *Citrus wilsonii* Tanaka extract combined with alginatecalcium coating on quality maintenance of white shrimps (*Litopenaeus vannamei* Boone). Food control. 2016;68:83-91.

[108] Balti R, Mansour MB, Zayoud N, Le Balc'h R, Brodu N, Arhaliass A, Massé A. Active exopolysaccharides

based edible coatings enriched with red seaweed (*Gracilaria gracilis*) extract to improve shrimp preservation during refrigerated storage. Food Bioscience. 2020;34:100522.

[109] Arulkumar A, Paramasivam S, Miranda JM. Combined effect of icing medium and red alga *Gracilaria verrucosa* on shelf life extension of Indian Mackerel (*Rastrelliger kanagurta*). Food and Bioprocess Technology. 2018;11:1911-1922.

[110] Arulkumar A, Satheeshkumar K, Paramasivam S, Rameshthangam P, Miranda JM. Chemical Biopreservative Effects of Red Seaweed on the Shelf Life of Black Tiger Shrimp (*Penaeus monodon*). Foods. 2020;9:634.

[111] Yan XJ. Quantitative determination of phlorotannins from some Chinese common brown seaweeds. Studia Marina Sinica. 1996;37:61-65.

[112] Athukorala Y, Lee KW, Song C, Ahn CB, Shin TS, Cha YJ, Shahidi F, Jeon YJ. Potential antioxidant activity of marine red alga *Grateloupia filicina* extracts. Journal of Food Lipids. 2003;10:251-265.

[113] Wang T, Jónsdóttir R, Kristinsson HG, Thorkelsson G, Jacobsen C, Hamaguchi PY, Ólafsdóttir G. Inhibition of haemoglobin-mediated lipid oxidation in washed cod muscle and cod protein isolates by *Fucus vesiculosus* extract and fractions. Food Chemistry. 2010;123:321-330.

[114] Ortiz J, Vivanco JP, Aubourg SP. Lipid and sensory quality of canned Atlantic salmon (*Salmo salar*): Effect of the use of different seaweed extracts as covering liquids. European Journal of Lipid Science and Technology. 2014;116:596-605.

[115] Maqsood S, Benjakul S, Shahidi F. Emerging role of phenolic compounds as natural food additives in fish and fish products. Critical reviews in food science and nutrition. 2013;53: 162-179.

[116] Park JW. Temperature-tolerant fish protein gels using konjac flour. Journal of Muscle Foods. 1996;7:165-174.

[117] Montero P, Hurtado JL, Pérez-Mateos M. Microstructural behaviour and gelling characteristics of myosystem protein gels interacting with hydrocolloids. Food Hydrocolloids. 2000;14:455-461.

[118] Da Ponte DJ, Roozen JP, Pilnik W. Effects of iota carrageenan, carboxymethyl cellulose and xanthan gum on the stability of formulated minced fish products. International Journal of Food Science & Technology. 1987;22:123-133.

[119] Bashir KM, Kim JS, An JH, Sohn JH, Choi JS. Natural food additives and preservatives for fish-paste products: a review of the past, present, and future states of research. Journal of Food Quality. 2017.

[120] Ahmed J, Mulla M, Arfat YA. Mechanical, thermal, structural and barrier properties of crab shell chitosan/ graphene oxide composite films. Food Hydrocolloids. 2017;71:141-148.

[121] Yu D, Regenstein JM, Zang J, Xia W, Xu Y, Jiang Q, Yang F. Inhibitory effects of chitosan-based coatings on endogenous enzyme activities, proteolytic degradation and texture softening of grass carp (*Ctenopharyngodon idellus*) fillets stored at 4 C. Food chemistry. 2018;262:1-6.

[122] Li XP, Zhou MY, Liu JF, Xu YX, Mi HB, Yi SM, Li JR, Lin H. Shelf-life extension of chilled olive flounder (*Paralichthys olivaceus*) using chitosan coatings containing clove oil. Journal of food processing and preservation. 2017;41:13204. [123] Yuan G, Lv H, Tang W, Zhang X, Sun H. Effect of chitosan coating combined with pomegranate peel extract on the quality of Pacific white shrimp during iced storage. Food Control. 2016;59:818-823.

[124] Wu C, Li Y, Wang L, Hu Y, Chen J, Liu D, Ye X. Efficacy of chitosan-gallic acid coating on shelf life extension of refrigerated Pacific mackerel fillets. Food and Bioprocess Technology. 2016;9:675-685.

[125] Ehsani A, Hashemi M, Afshari A, Aminzare M, Raeisi M, Zeinali T. Effect of different types of active biodegradable films containing lactoperoxidase system or sage essential oil on the shelf life of fish burger during refrigerated storage. LWT. 2020;117:108633.

[126] Nawaz T, Fatima M, Shah SZ, Afzal M. Coating effect of rosemary extract combined with chitosan on storage quality of mori (*Cirrhinus mrigala*). Journal of Food Processing and Preservation. 2020;44:14833.

[127] Hosseini SF, Rezaei M, Zandi M, Ghavi FF. Effect of fish gelatin coating enriched with oregano essential oil on the quality of refrigerated rainbow trout fillet. Journal of Aquatic Food Product Technology. 2016;25:835-842.

[128] Mirzapour-Kouhdasht A, Moosavi-Nasab M. Shelf-life extension of whole shrimp using an active coating containing fish skin gelatin hydrolysates produced by a natural protease. Food science & nutrition. 2020;8:214-223.

[129] Zhao X, Wu JE, Chen L, Yang H.
Effect of vacuum impregnated fish gelatin and grape seed extract on metabolite profiles of tilapia (*Oreochromis niloticus*) fillets during storage. Food chemistry.
2019;293:418-428.

[130] Feng X, Bansal N, Yang H. Fish gelatin combined with chitosan coating inhibits myofibril degradation of golden pomfret (*Trachinotus blochii*) fillet during cold storage. Food Chemistry. 2016;200:283-292.

[131] Farajzadeh F, Motamedzadegan A, Shahidi SA, Hamzeh S. The effect of chitosan-gelatin coating on the quality of shrimp (*Litopenaeus vannamei*) under refrigerated condition. Food Control. 2016;67:163-170.

[132] Thaker M, Hanjabam MD, Gudipati V, Kannuchamy N. Protective Effect of Fish Gelatin-Based Natural Antimicrobial Coatings on Quality of Indian Salmon Fillets during Refrigerated Storage. Journal of Food Process Engineering. 2017;40:12270.

[133] Kakaei S, Shahbazi Y. Effect of chitosan-gelatin film incorporated with ethanolic red grape seed extract and *Ziziphora clinopodioides* essential oil on survival of *Listeria monocytogenes* and chemical, microbial and sensory properties of minced trout fillet. LWT-Food Science and Technology. 2016;72:432-438.

[134] Shokri S, Ehsani A. Efficacy of whey protein coating incorporated with lactoperoxidase and  $\alpha$ -tocopherol in shelf life extension of Pike-Perch fillets during refrigeration. LWT-Food Science and Technology. 2017;85:225-231.

[135] Erbay EA, Dağtekin BB, Türe M, Yeşilsu AF, Torres-Giner S. Quality improvement of rainbow trout fillets by whey protein isolate coatings containing electrospun poly (ε-caprolactone) nanofibers with *Urtica dioica* L. extract during storage. Lwt. 2017;78:340-351.

[136] Shokri S, Ehsani A, Jasour MS. Efficacy of lactoperoxidase systemwhey protein coating on shelf-life extension of rainbow trout fillets during cold storage (4 C). Food and Bioprocess Technology. 2015;8:54-62

[137] Mosquera M, Gimenez B, Ramos S, Lopez-Caballero ME, Gomez-Guill ' en, MdC, Montero P. Antioxidant, ACE-Inhibitory, and antimicrobial activities of peptide fractions obtained from dried giant squid tunics. Journal of Aquatic Food Product Technology. 2016;25:444-455. https://doi.org/10.1080/10498850.2 013.819543

[138] Irshad I, Kanekanian A, Peters A, Masud T. Antioxidant activity of bioactive peptides derived from bovine casein hydrolysate fractions. Journal of Food Science and Technology.
2015;52:231-239.

[139] Park JW. Functional protein additives in surimi gels. Journal of Food Science. 1994;59:525-527.

[140] Wasson DH, Reppond KD, Babbitt JK, French JS. Effects of additives of proteolytic and functional properties of arrowtooth flounder surimi. Journal of Aquatic Food Product Technology. 1993;1:147-165.

[141] Sharma G, Dang S, Gupta S, Gabrani R. Antibacterial activity, cytotoxicity, and the mechanism of action of bacteriocin from *Bacillus subtilis* GAS101. Medical Principles and Practice. 2018;27:186-192.

[142] Woraprayote W, Malila Y, Sorapukdee S, Swetwiwathana A, Benjakul S, Visessanguan W. Bacteriocins from lactic acid bacteria and their applications in meat and meat products. Meat Science. 2016;120:118-132.

[143] Ashwitha A, Thamizharasan K, Vithya V, Karthik R. Effectiveness of bacteriocin from *Bacillus subtilis* (KY808492) and its application in biopreservation. Journal of Fisheries Sciences com. 2017;11:36-42

[144] Schelegueda LI, Gliemmo MF, Campos CA. Antimicrobial synergic effect of chitosan with sodium lactate, nisin or potassium sorbate against the bacterial flora of fish. Journal of Food Research. 2012;1:272-281.

[145] Ye M, Neetoo H, Chen H. Effectiveness of chitosan-coated plastic films incorporating antimicrobials in inhibition of *Listeria monocytogenes* on cold-smoked salmon. International Journal of Food Microbiology. 2008;127:235-240.

[146] Woraprayote W, Pumpuang L, Tosukhowong A, Zendo T, Sonomoto K, Benjakul S, Visessanguan W. Antimicrobial biodegradable food packaging impregnated with Bacteriocin 7293 for control of pathogenic bacteria in pangasius fish fillets. Lebensmittel-Wissenschaft & Technologie. 2018;89:427-433.

[147] Lv X, Du J, Jie Y, Zhang B, Bai F, Zhao H, Li J. Purification and antibacterial mechanism of fish-borne bacteriocin and its application in shrimp (*Penaeus vannamei*) for inhibiting *Vibrio parahaemolyticus*. World Journal of Microbiology and Biotechnology. 2017;33:1-2.

[148] Lv X, Ma H, Sun M, Lin Y, Bai F, Li J, Zhang B. A novel bacteriocin DY4-2 produced by *Lactobacillus plantarum* from cutlassfish and its application as bio-preservative for the control of *Pseudomonas fluorescens* in fresh turbot (*Scophthalmus maximus*) fillets. Food Control. 2018;89:22-31.

[149] Ladha G, Jeevaratnam K. Characterization of purified antimicrobial peptide produced by *Pediococcus pentosaceus* LJR1, and its application in preservation of white leg shrimp. World Journal of Microbiology and Biotechnology. 2020;36:1-2

[150] Sarika AR, Lipton AP, Aishwarya MS. Biopreservative efficacy of bacteriocin GP1 of *Lactobacillus rhamnosus* GP1 on stored fish filets. Frontiers in nutrition. 2019;6:29. [151] Shahinfar R, Khanzadi S, Hashami M, Azizzadeh M, Bostan A. The effect of *Ziziphora clinopodioides* essential oil and nisin on chemical and microbial characteristics of fish burger during refrigerated storage. Iranian Journal of Chemistry and Chemical Engineering (IJCCE). 2017;36:65-75.

[152] Wu Y, Shi YG, Zeng LY, Pan Y, Huang XY, Bian LQ, Zhu YJ, Zhang RR, Zhang J. Evaluation of antibacterial and anti-biofilm properties of kojic acid against five food-related bacteria and related subcellular mechanisms of bacterial inactivation. Food Science and Technology International. 2019;25:3-15.

[153] Wang R, Hu X, Agyekumwaa AK, Li X, Xiao X, Yu Y. Synergistic effect of kojic acid and tea polyphenols on bacterial inhibition and quality maintenance of refrigerated sea bass (*Lateolabrax japonicus*) fillets. Lebensmittel-Wissenschaft & Technologie. 2021;137:110452.

[154] Jia S, Liu Y, Zhuang S, Sun X, Li Y, Hong H, Lv Y, Luo Y. Effect of ε-polylysine and ice storage on microbiota composition and quality of Pacific white shrimp (*Litopenaeus* vannamei) stored at 0° C. Food microbiology. 2019;83:27-35.

[155] Zhang Z, Xia G, Yang Q, Fan X, Lyu S. Effects of chitosan-based coatings on storage quality of Chinese shrimp. Food science & nutrition. 2019;7:4085-4094.

[156] Li NA, Liu W, Shen Y, Mei J, Xie J. Coating effects of  $\varepsilon$ -polylysine and rosmarinic acid combined with chitosan on the storage quality of fresh halfsmooth tongue sole (*Cynoglossus semilaevis* Günther) fillets. Coatings. 2019;9:273.



