

---

---

## PROBIOTICS IN AQUATIC ANIMAL HEALTH MANAGEMENT

---

---

S. Ezhil Nilavan and Toms C Joseph

Microbiology, Fermentation & Biotechnology Division,

E-mail: theezhilnilavan@gmail.com

Aquaculture is a major food-producing activity that helps to cater for the needs of an ever-increasing populace. Nevertheless, infectious diseases have become a serious issue in aquaculture, resulting in significant financial losses for the industry. Treatment with costly chemotherapy medications has a detrimental effect on the aquatic ecosystem. As a result, there is an increasing worry about finding alternatives that are safe, non-antibiotic-based, and environmentally acceptable prophylactic approaches. Probiotics are a possible alternative to antibiotics for controlling infectious agents and treating disorders. Growth promotion, better metabolism, improved immunological response, and water quality maintenance are all advantages of probiotics. Probiotics help the fishes fight diseases and promote well being since they have antibacterial, antifungal, and antiviral capabilities. Probiotics help the fishes fight diseases and promote well being since they have antibacterial, antifungal, and antiviral capabilities. Probiotics are a unique concept in fish farming, and their effectiveness in an aquatic setting is still to be well investigated. This review presents current information about using probiotics, including selection criteria, kinds of probiotics utilized in fish farming, the mechanisms underlying, and probiotics administration methods.

### **Definition of probiotic?**

Probiotic is originally a Greek term, where 'Pro' means benefit and 'bios' means life. During 1907, a Russian analyst named Elie Metchnikoff noted that Bulgarian labourers lived long lives because they ate fermented dairy products. Lilly and Stillwell (1965) used the term "probiotic" to describe unknown growth-promoting chemicals produced by ciliated protozoan. In 1974, Parker defined probiotics as organism and substances add to intestinal balance.

In 1992, Fuller redefined the definition as a live microbial feed supplement that beneficially affects the host by improving the intestinal microbiological balance. According to a joint working group of the United Nations Food and Agriculture Organization (FAO) and the World Health Organization (WHO), probiotics are live microorganisms which, when administered in adequate amounts, confer a health benefit on the host (FAO/ WHO,2001). By releasing compounds such as bacteriocins and other inorganic compounds, probiotics defend the host body from harmful microorganisms.

Probiotics has gained increasing prominence as an alternate to antibacterial drugs in the aquaculture sector for increasing productivity and preventing disease. Whenever probiotics are fed to the fish, they have a positive effect on the fish host. Dietary intake of probiotics aids in the modification of the intestinal tract's microbes balance, as well as the immune modulation and also offers several nutritional advantages (Kesarcodi-Watson et al., 2008). Probiotics have a wide range of applications in aquaculture, in addition to their health and growth-promoting characteristics. Because of the complex link between an aquatic organism and its surroundings, the notion of probiotic use in fish culture has been developed to encompass water quality enhancement by directly introducing probiotics in ponds. For these reasons probiotics are defined as "water additives" (Moriarty, 1998). It is assumed that microorganisms that improve water quality also improve the health of aquatic animals, and various commercial products labelled as "probiotics" have attempted to capitalize on this theory. The research and potential application of probiotics in aquaculture has continued to grow during last couple of decades. Representatives of roughly 20 bacterial genera have recently been recognized as prospective probiotic candidates, with *Bacillus* spp and the *Lactobacillus* spp (LAB) group representing the bulk of promising species (Knipe et al., 2020).

#### **Characteristics of an ideal probiotic**

The vital role of probiotics is to establish or to maintain a health intestinal microbial flora in the fish (Thirumurugan and Vignesh, 2015). The following are the characteristics of an ideal probiotic.

- They should offer a beneficial effect on growth, maturation, and immunity against pathogens
- Probiotics should have no negative consequences for the host.
- Antibiotic resistance should never be a feature of probiotics; instead, they must be able to maintain inherited features.

Probiotics should have the following characteristics in order to be used as an effective feed probiotic,

- ✓ Withstand acidic conditions
- ✓ Resistant to gastric secretions
- ✓ Attach to the epithelium of the digestive tract
- ✓ Antagonism towards pathogenic microorganisms
- ✓ Immune system stimulation
- ✓ Increase in gut movement
- ✓ Able to survive in mucus
- ✓ Probiotics should have fermentative activity, resistance to drying, and viability in food during transport and storage.

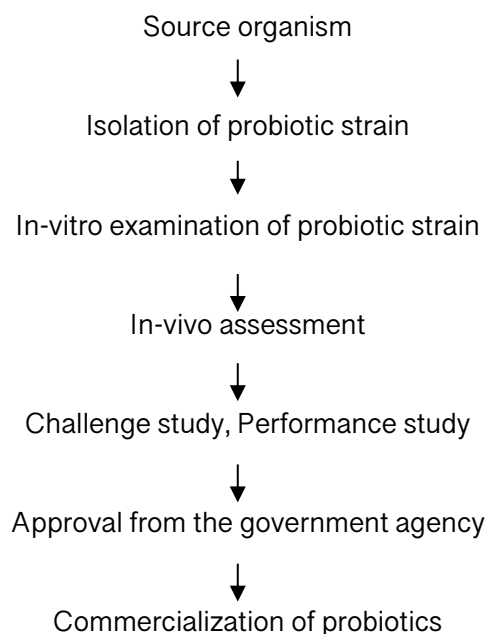
Organisms obtained from various sources are submitted to a series of tests in order to determine their suitability as ideal probiotic. The screening procedure includes Gram's reaction, in-vitro assessment of antagonistic characters, tolerance to acids and bile and susceptibility to antimicrobial drugs. If all of these criteria are met, they are considered a promising probiotic for use in fish culture.

### **Sources of bacterial probiotics**

Bacteria can be found in humans, animals, soil, sediment, aquatic environment and different numbers of bacteria ( $10^2$  - $10^{11}$  CFU/g) were observed in different environments (Liu et al., 2010). Bacteria from atmosphere, soil and anthropogenic activities can enter the aquatic system and alter the microbial load in the water, which further leads to the colonization of different bacteria in the gastrointestinal tract of aquatic organisms. The microbial load in the GIT of aquatic animals is normally  $10^2$  - $10^9$  CFU/g (Kim et al., 2007). Probiotic candidates potential has been evaluated in a variety of settings, including semi-intensive culture systems, intensive fish farms, and natural water bodies (Chantharasophon et al., 2011), where microbes obtained from outside of the hosts are referred to as "allochthonous or exogenous," and the ones recovered from the host are referred to as "autochthonous or indigenous" (Ringo et al., 2016).

### **Selection of probiotics**

The selection process of probiotics can be represented as follows



### **Types of probiotics**

Probiotics are grouped into two categories based on their mechanism of action. They are gut probiotics and water probiotics. Gut probiotics are normally administered through feed which

helps to improve the gut Microflora. Water probiotics are administered in the aquatic environment which intake all nutrients from the water and the harmful bacteria is eliminated from the system due to lack of nutrients.

<b>Types</b>	<b>Description</b>
Non-viable probiotics	Probiotics with dead microorganisms
Freeze dried probiotics	These probiotics will rapidly die upon leaving refrigeration
Fermented probiotics	These are probiotics which are produced through fermentation
Viable probiotics	These are live microorganisms , have a protocol to be counting, and to be very stable and efficacious

### **Probiotics in aquaculture**

Fish raised in an aquaculture facility are highly influenced by the microorganisms in the surrounding water (Verschuere et al., 2010). Eukaryotes and commensal bacteria thrive in the aquaculture habitat, while opportunistic pathogens grow under favourable environmental condition (Moraity 1998). Opportunistic pathogens such as *Vibrio* spp invade the host through the gut and invade fish through the gills and skin (Weber et al., 2010).

The Firmicutes phylum contains some of the most investigated probiotic candidates, such as LAB (lactic acid-producing bacteria) and *Bacillus* spp (Amoa et al., 2019, Azad et al., 2019, Balcazar et al., 2008, Venkat et al., 2004). Lactic acid bacteria can survive acidic pH and bile salts, allowing them to live in gastrointestinal tract despite not being acclimated to the aquatic environment (Merrifield et al., 2010). These bacteria can colonize the intestinal mucus, whereupon they aid in the digestion and absorption of food, boosting the fish's growth and development.

### **Mode of administration of probiotics**

Probiotics in aquaculture could be given by a variety of ways, including feed, injections, and direct exposure to water. Probiotics can be used alone or in combinations (Hai et al., 2015).

#### ***Feed additives, water additives and injection***

Incorporation of probiotic combinations into the feed is by far the most typical way of probiotics administration. Melo et al., 2021 stated that 92.8 % of probiotics are given as feed, followed by direct addition to water (4.8%) and in live food (1.8%) in fish culture systems. In aquaculture; probiotics such as bacterial strains, yeast, and extracted compounds are commonly used as food supplements. Dietary supplementation of probiotic strains of *Lactobacillus plantarum* has resulted in better growth and increased immunity in *Pangasius larnaudii* (Silarudee et al., 2019). Sahandi et al., 2019 reported that *Bifidobacterium* strains given as feed additive has improved the growth and nutrient utilization in rainbow trout fry.

There are several reports suggesting that probiotics can also be administered through water as an additive (Gopi et al., 2016, Gupta et al., 2016). In the sea bream, probiotic *Vibrio lentus* administered through water at a concentration of  $10^6$  CFU/ ml significantly altered gene expression, including immune response, cell proliferation adhesion, Reactive Oxygen Species, and iron transfer (Schaek et al., 2017). In addition to the above methods probiotics can also be given as injection. Injection of *Enterobacter* sp through intramuscular route enhanced the immunity in rainbow trout (Laptra et al., 2014).

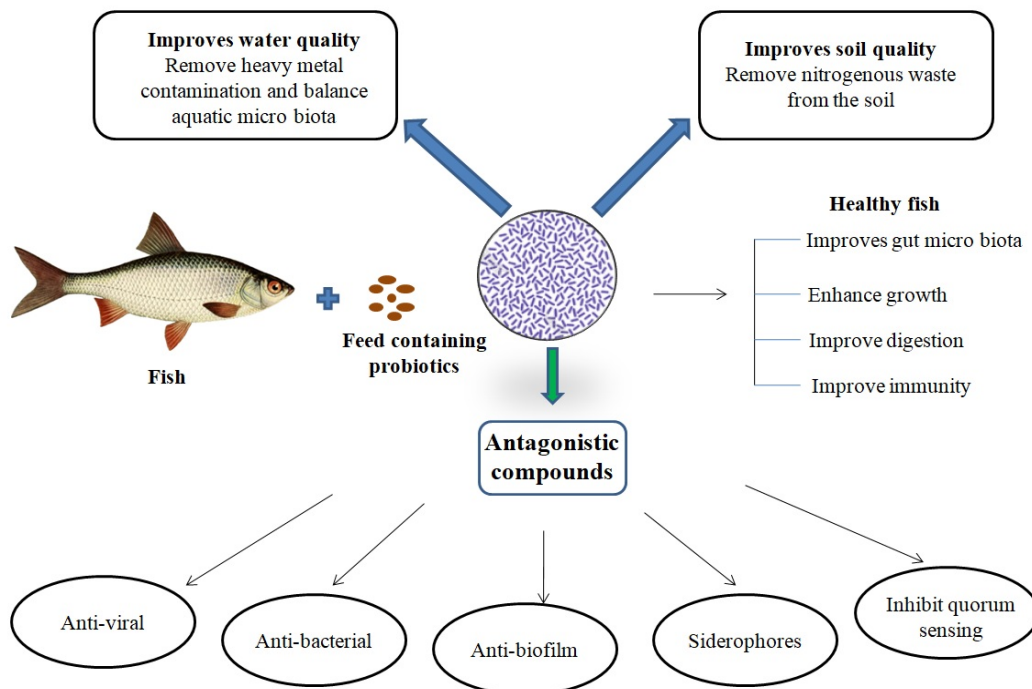
#### **Single and combinations of probiotics**

Probiotics come in a variety of forms, including multi-strain probiotics, probiotics with bioactive compounds, and probiotics with fermented products. The majority of research on probiotics in aquaculture has concentrated on single probiotics, while probiotic combinations are more effective. Multi-strain probiotics have the benefit of being more sensitive to pathogenic organisms and active against a variety of hosts (Pannu et al., 2014). Multi-strain probiotic has a positive effect on the growth and survival of *Labeo rohita* fingerlings (Jha et al., 2014).

#### **Beneficial effects and mode of action of probiotics in aquaculture: Figure 1**

The threat of disease development inside the aquaculture sector stimulates probiotic research and analysis to build more sustainable aquaculture. With the increasing public awareness over the use of antibiotics, that's not surprise that the probiotics for aquaculture is growing at a quick pace.

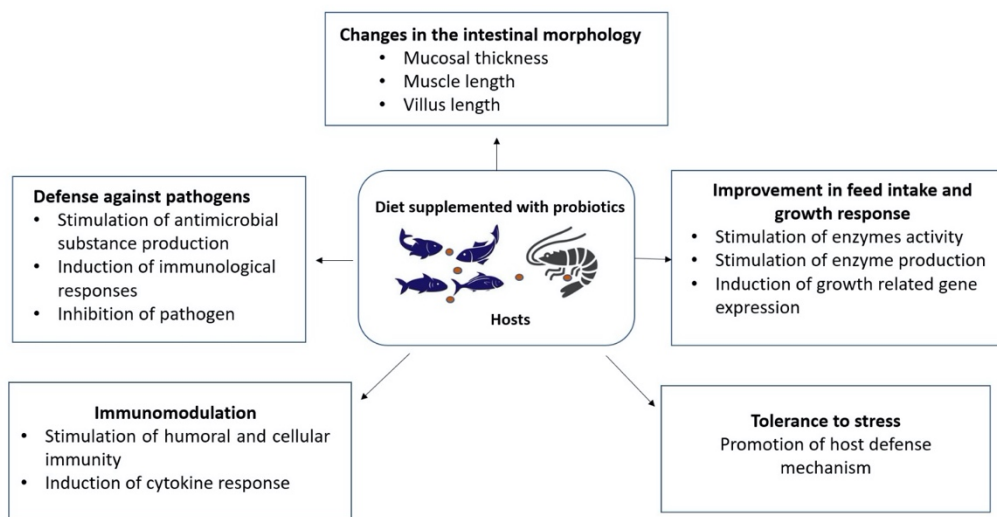
Probiotics have now been recommended by the Food and Agricultural Organization (FAO) for improving aquatic environmental quality by reducing mortality (Subasinghe, 2005). *Bacillus*, *Lactobacillus*, and *Bifidobacterium* are by far the most often used probiotic bacteria. Different species of *Lactobacillus*, *Bifidobacterium* and *Streptococcus* are used as probiotic in aquaculture which include *L. acidophilus*, *L. casei*, *L. fermentum*, *L. plantarum*, *L. salivarius*, *B. bifidum*, *B. lactum*, *B. breve*, *S. boulardii*, *S. thermophiles* and *S. cremonis* (Reda et al., 2018).



### Application of probiotics in Indian Aquaculture: Table 1

In the Indian aquaculture industry, intensive and semi-intensive farming practices have emerged one of the most practical and viable choices for meeting the nutritional needs of a rapidly growing population. Furthermore, the use of new techniques, such as the administration of probiotics, has increased total production and quality (Bandyopadhyay et al., 2015). *Bacillus* spp., *Lactobacillus* spp., *Bifidobacterium* spp., *Enterococcus* spp., *Streptomyces* spp., *Carnobacterium* spp., and yeast are the most often employed probiotic bacteria in aquaculture today (Van Doan et al., 2020).

According to studies, Gram +ve bacteria (*Bacillus* species) have been used as probiotics to improve the water quality. Gram positive bacteria, particularly *Bacillus* species, were shown to be highly efficient at converting organic materials to CO<sub>2</sub>, slime, or microbial biomass. Gram - positive appear to do superior than Gram negative in investigations. Producers can also manage the development of gaseous and particulate organic carbon during the growth period by ensuring a high standard of probiotics inside the production pond, according to the researchers (Mohapatra et al., 2013). Nitrifying probiotic bacteria are advantageous because they can substantially increase the microbial content in the water and improve the water quality by removing ammonia and nitrate toxicity (Zorriehzahra et al. 2016). Temperatures, acidity, dissolved oxygen, ammonia, and hydrogen sulphide in rearing water were also determined to be of higher quality after the administration of probiotics. Probiotics provide a favourable and healthy environment in aquatic systems for prawn and shrimp larval rearing (Banerjee et al. 2010).



Overview of beneficial effects of probiotics on fish and shellfish

**Table 1. Probiotic species used in finfish aquaculture, source and beneficial effects to the host species**

Probiotic species	Host	Beneficial effects	Reference
<i>Bacillus amyloliquefaciens</i> <i>COFCAU-P1</i>	<i>Labeo rohita</i>	Disease resistance against <i>A. hydrophila</i>	Khan et al., 2022
<i>Bacillus amyloliquefaciens</i> <i>Bacillus subtilis</i> <i>Bacillus megaterium</i>	<i>Labeo rohita</i>	Increased survival against <i>A. hydrophila</i> infection	Saravanan et al., 2021
<i>Saccharomyces cerevisiae</i>	<i>Labeo rohita</i>	Growth performance, haematological parameters, improved feed utilization	Jahan et al., 2021
<i>Lactobacillus fermentum</i>	<i>Cirrhinus mrigala</i>	Better growth, haematological parameters, improved feed utilization	Krishnaveni et al., 2021
<i>Bacillus methylotrophicus</i> <i>Bacillus licheniformes</i>	<i>Labeo rohita</i>	Increased survival against <i>A. hydrophila</i> infection	Mukherjee et al., 2019
<i>Bacillus amyloliquefaciens</i>	<i>Labeo rohita</i>	Increased antibody concentration, stress reduction	Nandi et al., 2018

<i>Bacillus subtilis</i> <i>Lactobacillus rhamnosus</i>	<i>Labeo rohita</i>	Enhanced feed digestibility	Munirasu et al., 2017
<i>Bacillus subtilis</i> <i>Terribacillus saccharophilus</i>	<i>Labeo rohita</i>	Increased growth and immunity	Kalarani et al., 2016
<i>Saccharomyces cerevisiae</i>	<i>Labeo rohita</i>	Increased growth and immunity	Bandopadhyay et al., 2015
<i>Bacillus subtilis</i> <i>FPTB13</i>	<i>Catlacatla</i>	Immunomodulation and disease resistance	Sangama et al., 2015
<i>Bacillus subtilis</i> <i>Pseudomonas aeruginosa</i> <i>Lactobacillus plantarum</i>	<i>Labeo rohita</i>	Highest survival rate against <i>A. hydrophila</i> infection	Giri et al., 2014
<i>Bacillus subtilis</i> <i>Lactobacillus lactis</i> <i>Saccharomyces cerevisiae</i>	<i>Labeo rohita</i>	Increased survival against <i>A. hydrophila</i> infection	Mohapatra et al., 2014
<i>Bacillus cereus</i>	<i>Penaeus monodon</i>	Growth promoter	Navinchandran et al., 2014
<i>Lactobacillus plantarum</i> VSG3	<i>Labeo rohita</i>	Improved growth, immunity and disease resistance	Giri et al., 2013
<i>Bacillus amyloliquefaciens</i>	<i>Catlacatla</i>	Improved growth, immunity and disease resistance	Das et al., 2013
<i>Lactobacillus rhamnosus</i>	<i>Oncorhynchus mykiss</i>	Improved Blood parameters	Panigrahi et al., 2010
<i>Bacillus</i> NL 110 <i>Vibrio</i> NE 17	<i>Macrobrachium rosenbergii</i>	Increased growth and immunity	Mujeeb et al., 2010

## Conclusion

Substantial research on the efficacy and actions of probiotic strains, many aspects remain unanswered. Additional and future research could focus on gut bacteria transcriptome and proteome profiling, host/microbe interactions, interactions among gut microorganisms, gut immune status, antioxidant status, antagonistic activity, and knowledge on the side effects of



probiotics. Aquaculture is indeed one of the world's fastest-growing industries, accounting for around 90% of worldwide production. Aquaculture offers a vital supply of nutritious food for human consumption; however diseases in the fish farming business have a negative impact on the nation's socioeconomic status and economic development.

Because antimicrobial agents used in therapeutic strategies have side effects including residual toxic effects, emerging antimicrobial resistance, immune system suppression, and reduced customer desire for drug-treated fishery products available in the market, non-antibiotic-based, eco friendly alternatives are in high demand for aquatic animal health management.

Probiotics are an excellent alternative sustainable option of beneficial microorganisms with strong antimicrobial activity, immunostimulatory abilities of boosting health and wellbeing to enhance growth and yield, strengthen the immune function, hinder QS as a new anti-infective approach, mitigate the adverse affects of reactive oxygen species (ros stressors, and greater resistance. In order to recommend potent therapeutic, bacteria-based approaches to enhance the health, production, and economic growth of the aquaculture sector, an interactive approach among academics, researchers, growers, and fish sector owners is needed to concentrate and start exploring the specific elements of bacteria host interactions bestowing the potential significant improvements in various immune function triggered by different bacterial species. . The synthesis of probiotics ought to be feasible on a broad scale with low operating costs. They ought not to be regarded as just a 'magic elixir,' but instead as a source of nourishment.

#### **Reference:**

- Amoah, K., Huang, Q. C., Tan, B. P., Zhang, S., Chi, S. Y., Yang, Q. H., & Dong, X. H. (2019). Dietary supplementation of probiotic *Bacillus coagulans* ATCC 7050, improves the growth performance, intestinal morphology, microflora, immune response, and disease confrontation of Pacific white shrimp, *Litopenaeus vannamei*. *Fish & shellfish immunology*, *87*, 796-808.
- Araujo, C., Munoz-Atienza, E., Poeta, P., Igrejas, G., Hernandez, P. E., Herranz, C., & Cintas, L. M. (2016). Characterization of *Pediococcus acidilactici* strains isolated from rainbow trout (*Oncorhynchus mykiss*) feed and larvae: safety, DNA fingerprinting, and bacteriocinogenicity. *Diseases of aquatic organisms*, *119*(2), 129-143.
- Azad, M. A. K., Islam, S. S., Sithi, I. N., Ghosh, A. K., Banu, G. R., Bir, J., & Huq, K. A. (2019). Effect of probiotics on immune competence of giant freshwater prawn *Macrobrachium rosenbergii*. *Aquaculture Research*, *50*(2), 644-657.

- Balcázar, J. L., Vendrell, D., de Blas, I., Ruiz-Zarzuela, I., Muzquiz, J. L., & Girones, O. (2008). Characterization of probiotic properties of lactic acid bacteria isolated from intestinal microbiota of fish. *Aquaculture*, *278*(1-4), 188-191.
- Bandyopadhyay, P., Mishra, S., Sarkar, B., Swain, S. K., Pal, A., Tripathy, P. P., & Ojha, S. K. (2015). Dietary *Saccharomyces cerevisiae* boosts growth and immunity of IMC *Labeo rohita* (Ham.) juveniles. *Indian Journal of Microbiology*, *55*(1), 81-87.
- Banerjee, S., Khatoon, H., Shariff, M., & Yusoff, F. M. (2010). Enhancement of *Penaeus monodon* shrimp postlarvae growth and survival without water exchange using marine *Bacillus pumilus* and periphytic microalgae. *Fisheries Science*, *76*(3), 481-487.
- Chantharasophon, K., Warong, T., Mapatsa, P., & Leelavatcharamas, V. (2011). High potential probiotic *Bacillus* species from gastro-intestinal tract of Nile Tilapia (*Oreochromis niloticus*).
- Das, A., Nakhro, K., Chowdhury, S., & Kamilya, D. (2013). Effects of potential probiotic *Bacillus amyloliquifaciens* [corrected] FPTB16 on systemic and cutaneous mucosal immune responses and disease resistance of catla (*Catla catla*). *Fish and Shellfish Immunology*, *35*(5), 1547–1553.
- FAO/WHO, 2001. Report on Joint FAO/WHO Expert Consultation on Evaluation of Health and Nutritional Properties of Probiotics in Food Including Powder Milk with Live Lactic Acid Bacteria. 1–4 October, Cordoba, Argentina
- Giri, S. S., Sukumaran, V., & Oviya, M. (2013). Potential probiotic *Lactobacillus plantarum* VSG3 improves the growth, immunity, and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish and Shellfish Immunology*, *34*(2), 660–666.
- Gobi, N., Malaikozhundan, B., Sekar, V., Shanthi, S., Vaseeharan, B., Jayakumar, R., & Nazar, A. K. (2016). GFP tagged *Vibrio parahaemolyticus* Dahv2 infection and the protective effects of the probiotic *Bacillus licheniformis* Dahb1 on the growth, immune and antioxidant responses in *Pangasius hypophthalmus*. *Fish & Shellfish Immunology*, *52*, 230-238.
- Gupta, A., Gupta, P., & Dhawan, A. (2016). *Paenibacillus polymyxa* as a water additive improved immune response of *Cyprinus carpio* and disease resistance against *Aeromonas hydrophila*. *Aquaculture Reports*, *4*, 86-92.
- Hai, N. V. (2015). The use of probiotics in aquaculture. *Journal of applied microbiology*, *119*(4), 917-935.
- Harikrishnan, R., Balasundaram, C., & Heo, M. S. (2010). Effect of probiotics enriched diet on *Paralichthys olivaceus* infected with lymphocystis disease virus (LCDV). *Fish & Shellfish Immunology*, *29*(5), 868-874.

- Jahan, N., Islam, S. M., Rohani, M. F., Hossain, M. T., & Shahjahan, M. (2021). Probiotic yeast enhances growth performance of rohu (*Labeo rohita*) through upgrading hematology, and intestinal microbiota and morphology. *Aquaculture*, *545*, 737243.
- Jha, D. K., Bhujel, R. C., & Anal, A. K. (2015). Dietary supplementation of probiotics improves survival and growth of Rohu (*Labeo rohita* Ham.) hatchlings and fry in outdoor tanks. *Aquaculture*, *435*, 475-479.
- Kalarani, V., Sumathi, V., Roshan, J. K., Sowjanya, D., & Reddy, D. C. (2016). Effect of dietary supplementation of *Bacillus subtilis* and *Terribacillus saccharophilus* on innate immune responses of a tropical freshwater fish, *Labeo rohita*. *Journal of Clinical and Cellular Immunology*, *7*, 395.
- Kesarcodi-Watson A, Kaspar H, Lategan MJ and Gibson L, 2008. Probiotics in aquaculture: the need, principles and mechanisms of action and screening processes. *Aquaculture*, *274*(1): 1-14, doi: 10.1016/j.aquaculture.2007.11.019
- Khan, M. I. R., Kamilya, D., Choudhury, T. G., & Rathore, G. (2022). Dietary administration of a host-gut derived probiotic *Bacillus amyloliquefaciens* COFCAU\_P1 modulates immune-biochemical response, immune-related gene expression, and resistance of *Labeo rohita* to *Aeromonas hydrophila* infection. *Aquaculture*, *546*, 737390.
- Kim, D. H., Brunt, J., & Austin, B. (2007). Microbial diversity of intestinal contents and mucus in rainbow trout (*Oncorhynchus mykiss*). *Journal of applied microbiology*, *102*(6), 1654-1664.
- Knipe H, Temperton B, Lange A, Bass D and Tyler CR, 2020. Probiotics and competitive exclusion of pathogens in shrimp aquaculture. *Rev Aquac*, 1-29, doi: 10.1111/raq.12477
- Krishnaveni, G., Vignesh, S., Vidhyalakshmi, N., Vijay, V., & Ramesh, U. (2021). Effects of dietary supplementation of *Lactobacillus fermentum* URLP18 on growth, innate immunity and survival against *Aeromonas hydrophila* ATCC 7966 challenge in freshwater fish *Cyprinus carpio* (common carp). *Aquaculture Research*, *52*(3), 1160-1176..
- Krishnaveni, G., Vignesh, S., Vidhyalakshmi, N., Vijay, V., & Ramesh, U. (2021). Effects of dietary supplementation of *Lactobacillus fermentum* URLP18 on growth, innate immunity and survival against *Aeromonas hydrophila* ATCC 7966 challenge in freshwater fish *Cyprinus carpio* (common carp). *Aquaculture Research*, *52*(3), 1160-1176.
- LaPatra, S. E., Fehring, T. R., & Cain, K. D. (2014). A probiotic *Enterobacter* sp. provides significant protection against *Flavobacterium psychrophilum* in rainbow trout (*Oncorhynchus mykiss*) after injection by two different routes. *Aquaculture*, *433*, 361-366.

- Liu, K. F., Chiu, C. H., Shiu, Y. L., Cheng, W., & Liu, C. H. (2010). Effects of the probiotic, *Bacillus subtilis* E20, on the survival, development, stress tolerance, and immune status of white shrimp, *Litopenaeus vannamei* larvae. *Fish & shellfish immunology*, *28*(5-6), 837-844.
- Melo-Bolívar, J. F., Ruiz Pardo, R. Y., Hume, M. E., & Villamil Diaz, L. M. (2021). Multistrain probiotics use in main commercially cultured freshwater fish: A systematic review of evidence. *Reviews in Aquaculture*, *13*(4), 1758-1780.
- Merrifield, D. L., Dimitroglou, A., Foey, A., Davies, S. J., Baker, R. T., Børgwald, J., & Ringø, E. (2010). The current status and future focus of probiotic and prebiotic applications for salmonids. *Aquaculture*, *302*(1-2), 1-18.
- Mohapatra, S., Chakraborty, T., Prusty, A. K., Prasad, K. P., & Mohanta, K. N. (2014). Dietary multispecies probiotic supplementation enhances the immunohematological responses and reduces mortality by *Aeromonas hydrophila* in *Labeo rohita* fingerlings. *Journal of the World Aquaculture Society*, *45*(5), 532-544.
- Moriarty DJW, 1998. Control of luminous *Vibrio* species in penaeid aquaculture ponds. *Aquaculture*, *164*: 351-358, doi: 10.1016/S0044-8486(98)00199-9
- Moriarty, D. J. W. (1998). Control of luminous *Vibrio* species in penaeid aquaculture ponds. *Aquaculture*, *164*(1-4), 351-358.
- Mujeeb Rahiman, K. M., Jesmi, Y., Thomas, A. P., & Mohamed Hatha, A. A. (2010). Probiotic effect of *Bacillus* NL110 and *Vibrio* NE17 on the survival, growth performance and immune response of *Macrobrachium rosenbergii* (de Man). *Aquaculture research*, *41*(9), e120-e134.
- Munirasu, S., Ramasubramanian, V., & Arunkumar, P. (2017). Effect of probiotics diet on growth and biochemical performance of freshwater fish *Labeo rohita* fingerlings. *Journal of Entomology and Zoology Studies*, *5*(3), 1374–1379.
- Nandi, A., Banerjee, G., Dan, S. K., Ghosh, K., & Ray, A. K. (2018). Evaluation of *in vivo* probiotic efficiency of *Bacillus amyloliquefaciens* in *Labeo rohita* challenged by pathogenic strain of *Aeromonas hydrophila* MTCC 1739. *Probiotics and Antimicrobial Proteins*, *10*(2), 391–398.
- NavinChandran, M., Iyapparaj, P., Moovendhan, S., Ramasubburayan, R., Prakash, S., Immanuel, G., & Palavesam, A. (2014). Influence of probiotic bacterium *Bacillus cereus* isolated from the gut of wild shrimp *Penaeus monodon* in turn as a potent growth promoter and immune enhancer in *P. monodon*. *Fish and Shellfish Immunology*, *36*(1), 38–45.
- Panigrahi, A., Kiron, V., Satoh, S., & Watanabe, T. (2010). Probiotic bacteria *Lactobacillus rhamnosus* influences the blood profile in rainbow trout *Oncorhynchus mykiss* (Walbaum). *Fish Physiology and Biochemistry*, *36*(4), 969–977.

- Pannu, R., Dahiya, S., Sabhlok, V. P., Kumar, D., Sarsar, V., & Gahlawat, S. K. (2014). Effect of probiotics, antibiotics and herbal extracts against fish bacterial pathogens. *Ecotoxicology and Environmental Contamination*, *9*(1), 13-20.
- Reda, R. M., Selim, K. M., El-Sayed, H. M., & El-Hady, M. A. (2018). In vitro selection and identification of potential probiotics isolated from the gastrointestinal tract of Nile tilapia, *Oreochromis niloticus*. *Probiotics and antimicrobial proteins*, *10*(4), 692-703.
- Ringø, E. Z. Z. V., Zhou, Z., Vecino, J. G., Wadsworth, S., Romero, J., Krogh, A., & Merrifield, D. L. (2016). Effect of dietary components on the gut microbiota of aquatic animals. A never-ending story. *Aquaculture nutrition*, *22*(2), 219-282.
- Sahandi, J., Jafaryan, H., Soltani, M., & Ebrahimi, P. (2019). The use of two Bifidobacterium strains enhanced growth performance and nutrient utilization of Rainbow Trout (*Oncorhynchus mykiss*) fry. *Probiotics and antimicrobial proteins*, *11*(3), 966-972.
- Sangma, T., & Kamilya, D. (2015). Dietary Bacillus subtilis FPTB13 and chitin, single or combined, modulate systemic and cutaneous mucosal immunity and resistance of catla, *Catla catla* (Hamilton) against edwardsiellosis. *Comparative Immunology, Microbiology and Infectious Diseases*, *43*, 8-15.
- Saravanan, K., Sivaramakrishnan, T., Praveenraj, J., Kiruba-Sankar, R., Haridas, H., Kumar, S., & Varghese, B. (2021). Effects of single and multi-strain probiotics on the growth, hemato-immunological, enzymatic activity, gut morphology and disease resistance in Rohu, *Labeo rohita*. *Aquaculture*, *540*, 736749.
- Subasinghe, R. P. (2005). Epidemiological approach to aquatic animal health management: opportunities and challenges for developing countries to increase aquatic production through aquaculture. *Preventive veterinary medicine*, *67*(2-3), 117-124.
- Thirumurugan R, Vignesh V (2015) Probiotics: live boon to aquaculture. In: Advances in marine and brackishwater aquaculture. Springer India, pp 51–61
- Van Doan, H., Hoseinifar, S. H., Ringø, E., Ángeles Esteban, M., Dadar, M., Dawood, M. A., & Faggio, C. (2020). Host-associated probiotics: a key factor in sustainable aquaculture. *Reviews in Fisheries Science & Aquaculture*, *28*(1), 16-42.
- Venkat, H. K., Sahu, N. P., & Jain, K. K. (2004). Effect of feeding Lactobacillus-based probiotics on the gut microflora, growth and survival of postlarvae of *Macrobrachium rosenbergii* (de Man). *Aquaculture Research*, *35*(5), 501-507.
- Verschuere, L., Rombaut, G., Sorgeloos, P., & Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology and molecular biology reviews*, *64*(4), 655-671.
- Weber, B., Chen, C., & Milton, D. L. (2010). Colonization of fish skin is vital for *Vibrio anguillarum* to cause disease. *Environmental microbiology reports*, *2*(1), 133-139.

Zorriehzahra, M. J., Delshad, S. T., Adel, M., Tiwari, R., Karthik, K., Dhama, K., & Lazado, C. C. (2016). Probiotics as beneficial microbes in aquaculture: an update on their multiple modes of action: a review. *Veterinary quarterly*, 36(4), 228-241.

\*\*\*\*\*